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13384-002001**TRANSMITTAL LETTER TO THE UNITED STATES  
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CONCERNING A FILING UNDER 35 U.S.C. 371**U.S. APPLICATION NO. (If Known, see 37 CFR  
1.5)

09/889874

INTERNATIONAL APPLICATION NO.  
PCT/GB00/00219INTERNATIONAL FILING DATE  
24 January 2000PRIORITY DATE CLAIMED  
22 January 1999TITLE OF INVENTION  
BIOLOGICAL CONTROL OF NEMATODES

APPLICANT(S) FOR DO/EO/US

James Alan Wynne Morgan, Paul Jarrett, Debbie Ellis and Margaret Anne Ousley

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to promptly begin national examination procedures (35 U.S.C. 371(f)).
4. ☒ The US has been elected by the expiration of 19 months from the priority date (PCT Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
  - a. ☐ is attached hereto (required only if not communicated by the International Bureau).
  - b. ☒ has been communicated by the International Bureau.
  - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
  - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
  - b. ☐ have been communicated by the International Bureau.
  - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
  - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)) (unsigned).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

**Items 11 to 16 below concern other documents or information included:**

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☐ A **FIRST** preliminary amendment.  
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☐ Other items or information:

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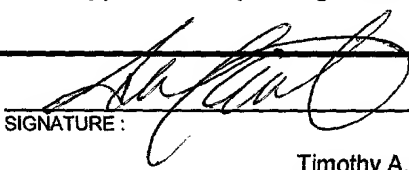
7-23-2001

Signature

Samantha Bell

Typed Name of  
Person Signing

Samantha Bell

U.S. APPLICATION NO. (IF KNOWN) <b>09/889874</b>		INTERNATIONAL APPLICATION NO. PCT/GB00/00219		ATTORNEY'S DOCKET NUMBER 13384-002001			
17. <input checked="" type="checkbox"/> The following fees are submitted:				CALCULATIONS PTO USE ONLY			
<b>Basic National Fee ( 37 CFR 1.492(a)(1)-(5) ):</b>  Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO ..... <b>\$1000</b>  International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO ..... <b>\$860</b>  International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO ..... <b>\$710</b>  International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) ..... <b>\$690</b>  International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) ..... <b>\$100</b>  <div style="text-align: right;"><b>ENTER APPROPRIATE BASIC FEE AMOUNT =</b></div>							
						<b>\$860.00</b>	
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Surcharge of <b>\$130</b> for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				<b>\$0.00</b>			
Claims	Number Filed	Number Extra	Rate				
Total Claims	34 - 20 = 14	4	x \$18	<b>\$72.00</b>			
Independent Claims	3 - 3 = 0	0	x \$80	<b>\$0.00</b>			
MULTIPLE DEPENDENT CLAIMS(S) (if applicable)			+ \$270	<b>\$270.00</b>			
<b>TOTAL OF ABOVE CALCULATIONS =</b>				<b>\$1,202.00</b>			
<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				<b>\$0.00</b>			
<b>SUBTOTAL =</b>				<b>\$1,202.00</b>			
Processing fee of <b>\$130</b> for furnishing the English Translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f))				<b>\$0.00</b>			
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Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +				<b>\$0.00</b>			
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<b>NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b) must be filed and granted to restore the application to pending status.</b>							
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Timothy A. French FISH & RICHARDSON P.C. 225 Franklin Street Boston, MA 02110-2804 (617) 542-5070 phone (617) 542-8906 facsimile			<div style="text-align: center;">             SIGNATURE:         </div> <div style="text-align: center;">           NAME            Timothy A. French         </div> <div style="text-align: center;">           REGISTRATION NUMBER            30,175         </div>				

14/PRTS

1

BIOLOGICAL CONTROL OF NEMATODES

## TECHNICAL FIELD

The present invention relates to methods and materials for controlling nematodes.

## PRIOR ART

Several thousand species of nematodes, sometimes called eel worms, are known. Numerous nematodes attack and parasitize humans and animals and cause disease. Additionally, several hundred species are known to feed on living plants. Certain of these are reviewed by Agrios in "Plant Pathology - 3rd Ed" Pub Academic Press Inc, see Chapter 15 therein.

Methods of controlling nematodes and their associated diseases include cultural practices; biological methods, e.g. use of resistant varieties; physical methods, e.g. heat; and use of chemical agents.

Patent application WO 92/19739 (Mycogen) relates to genes and gene fragments from *Bacillus thuringiensis* which have nematocidal activity. These generally encode crystal toxins from particular strains.

Patent application EP 0 303 426 (Mycogen) also relates to strains of *B. thuringiensis* which have nematocidal activity.

Patent application EP 0 171 381 (Monsanto) relates to particular soil bacteria which are capable of proliferating in an environment which is infested with

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nematodes such as pseudomonads which colonise the surface of plant roots. The basis for the controlling activity appears to stem from glycosidase enzymes which are hypothesised to directly inhibit the nematodes.

Notwithstanding these disclosures, there is an ongoing requirement for materials which have nematocidal activity, for instance for use in crop protection or nematode-mediated disease control.

Patent application PCT/WO 99/22598 (University of Reading) published 14 May 1999 claims a biopesticide for the control of insect pests or plant parasitic nematodes or both, which comprises as an effective agent a species of bacteria which is a symbiont of an entomopathogenic nematode.

#### DISCLOSURE OF THE INVENTION

The present inventors have established that species of bacteria which in nature are associated symbiotically with entomopathogenic nematodes, can in fact be utilised to control nematodes, and in preferred forms of the invention, to kill them. The bacteria themselves can be employed, or nematode control agents can be used which are derived from such bacteria. In one aspect of the invention, the present invention employs bacteria which are engineered and thus not naturally occurring, or nematode control agents which are derived from natural or non-natural bacteria.

It has been reported that certain bacterial species such as *Xenorhabdus* and *Photorhabdus* can be used to control insects, see e.g. PCT/WO 98/08388 of MAFF, PCT/WO 97/17432 of WARF, and PCT/WO 99/42589 of Novartis. An effect against nematodes had not previously been demonstrated.



The symbiotic bacteria used in the present invention are isolatable from nematodes or the insects which the nematodes attack, and differ fundamentally in terms of life-style and activity from those soil bacteria such as *B. thuringiensis* or pseudomonads which have previously been suggested as being nematocidal.

Indeed, *prima facie*, it seems highly unlikely that nematode symbiotes might possess nematocidal activity. However, in the light of the present disclosure, a number of possible explanations for the observed activity can be tentatively proposed. Firstly, in order to protect a nutrient supply from a dead insect, the bacteria might produce anti-nematocides to prevent saprophytic nematodes gaining access. Alternatively, to become a symbiont, the bacterial strains may have once been pathogens of these nematodes and evolved towards a less hostile symbiotic relationship. The nematocidal activity may be an evolutionary throwback from the original pathogenic relationship, in which case it may be expected to be widely present amongst bacteria which have evolved in this way.

A first aspect of the present invention is the use of bacterial strains to control a target nematode, characterised in that in nature the bacterial strain is associated symbiotically with an entomopathogenic nematode.

As discussed in more detail below, the bacterial strains may be used in the methods of the present invention *per se*, or they may be used as a source of nematode control agent. The nematode control agent can be derived directly, or be prepared and utilised through recombinant DNA techniques, optionally via a host cell.

The target nematode will generally be different to the nematode with which the bacterial strain is found symbiotically in nature.

By means of the present invention employing bacteria or a nematode control agent, it becomes possible to control nematodes, in the sense of, to prevent or retard the effect that the nematode has on other organisms such as animals or more preferably plants, or to reduce the number of nematodes or nematode eggs in an area of interest, or to alleviate or cure a disease caused by nematodes. Control may be at the level of larval nematodes or nematode eggs, or may inhibit the motion, feeding or infectivity of adult nematodes. Nematocidal control may be employed to kill the nematode target. Such controlling activity can be assessed as shown in the Examples below.

#### PREFERRED EMBODIMENTS

The present invention provides a composition for the control of parasitic nematodes which comprises as an effective agent a species of bacteria which is a symbiont of an entomopathogenic nematode, or engineered bacteria having such activity, or a nematode control agent derived from natural or engineered bacteria.

Correspondingly, the present invention also provides a method of nematode control employing such a composition.

The bacterial species is typically of the genera *Xenorhabdus* or *Photorhabdus*, preferably the genus *Xenorhabdus*, for instance the species *Xenorhabdus bovienii*. Examples of particularly preferred bacteria include:

*Xenorhabdus bovienii* strain H31 deposited with NCIMB under accession number NCIMB 40985 on 20 January 1999;

*Xenorhabdus bovienii* strain I73 deposited with NCIMB under accession number NCIMB 40986 on 05 November 1998; and

*Xenorhabdus* strain C42 deposited with NCIMB under accession number NCIMB 41004 on 05 November 1998.

The nematode control agent can be a peptide derived from a symbiont of an entomopathogenic nematode or an engineered bacterium has functional activity against a nematode. The peptide nematode control agent can be produced from a nucleic acid derived from a symbiont of an entomopath nematode or an engineered bacterium and which encodes such a peptide. The peptide can be an oligopeptide or a polypeptide, notably a protein. In one version, the nematode control agent is a toxin with toxic activity against nematodes, but the nematode control agent can have other activity.

The nucleic acids of this invention can be employed in a method of producing a peptide comprising the step of causing or allowing the expression from a nucleic acid of this invention in a suitable host cell.

The nucleic acid can comprise a natural nucleotide sequence or a degeneratively equivalent sequence, and functional variants thereof. Variants include homologous variants encoding a peptide which is a nematode control agent, the nucleic acid having 70% or more DNA sequence identity and/or the peptide having 70% or more amino acid sequence identity. Especially preferred nucleic acids in p 13-1f and p 14-2f and variants thereof.

The present invention extends to nucleic acids having a sequence which is a derivative by way of addition, insertion, deletion or substitution of one or more nucleotides. The nucleic acid can contain longer expressed sequences such that the nematode control agent is expressed as a fusion protein.

Nucleic acids complementary to the nucleic acid encoding a nematode

[illegible]

A method provided by this invention comprises the steps of:

- The hybridisation conditions can be selected to allow the identification of sequences having 70% or more sequence identity with the probe.

In one embodiment, the method comprises use of two primers to amplify a nucleic acid encoding a nematode control agent, at least one of the primers having a conserved nucleotide sequence of at least 15 nucleotides.

A method is further made possible by this invention comprising the steps of:

- (a) providing a preparation of nucleic acid from a bacterium,
- (b) providing a pair of nucleic acid molecule primers, at least one of which is a primer,
- (c) contacting nucleic acid in said preparation with said primers under

conditions for performance of PCR,

- (d) performing PCR and determining the presence or absence of an amplified PCR product.

Additionally, the invention provides a recombinant vector comprising a nucleic acid of this invention. The vector is preferably capable of replicating in a suitable host such as *E. coli* or in *Xenorhabdus*. The vector can be a baculovirus. In a preferred feature, the nucleic acid is operably linked to a promoter or other regulatory element for transcription in a host cell.

Vectors can further comprise any one or more of the following: a terminator sequence; a polyadenylation sequence; an enhancer sequence; a marker gene; a sequence encoding pesticidal material derived from *Bacillus thuringiensis*.

The vector can be a plant vector.

The vector of this invention can be introduced into a cell. Thus, a method for transforming a plant cell comprises the step of causing or allowing recombination between the vector and the plant cell genome to introduce the nucleic acid into the genome. The nucleic acid can be incorporated into chloroplast DNA, or into mitochondrial DNA.

Host cells comprising a vector are also part of this invention. The host cell can be a plant cell, which may be in a plant.

To this end, a method for producing a transgenic plant comprises the step of regenerating a plant from the transformed cell. In turn, plants of this invention extend to the progeny of such plants.

Examples of plants of this invention include crop species which can be

A method of influencing or affecting the toxicity of a cell such as a plant cell is provided where the method includes causing or allowing expression of a heterologous nucleic acid of this invention within the cells.

The present invention extends to control of helminthiasis in humans and other animals including domesticated animals such as swine, sheep, horses, cattle, goats, dogs, cats and poultry. The nematodes to be controlled include *Haemonchus*, *Trichostrongylus*, *Ostertagia*, *Nematodirus*, *Cooperia*, *Ascaris*, *Bunostomum*, *Oesophagosromuni*, *Chabberia*, *Trichuris*, *Strongylus*, *Trichonema*, *Dictyocaulus*, *Capillaria*, *Heterkis*, *Toxocara*, *Ascaridia*, *Oxyuris*, *Ancylostoma*, *Uncinaria*, *Toxascaris*, *Caenorhabditis* and *Parascaris*.

The compositions of this invention can be used in conjunction with *Bacillus*

*thuringiensis* or pesticidal materials derived therefrom.

In a further aspect, there is provided an antibody or fragment thereof, or a polypeptide comprising the antigen-binding domain of the antibody, capable of specifically binding a peptide of this invention.

Such an antibody or fragment can be obtained by immunising a mammal with the peptide, and is useful in a method of identifying and/or isolating a nematode control agent comprising the step of screening candidate polypeptides with a polypeptide comprising the antigen-binding domain of the antibody of claim.

Some further aspects of preferred embodiments of the invention will now be discussed.

#### Bacterial strains

These can be derived from any entomopathogenic nematode. Preferred species are *Xenorhabdus* and *Photorhabdus*.

Potential sources of bacteria for use in the methods of the present invention may be identified by any preferred method. For instance, entomopathogenic nematodes can be isolated using an insect baiting technique such as that described by Bedding & Akhurst (1975) *Nematologia* 21: 215-227. Bacteria from nematodes identified as being pathogenic to the insect are isolated, cultured, and used as a source of nematocidal agent, e.g. by analogy with the methods used in the Examples below. Preferably *Xenorhabdus* or *Photorhabdus* species are used.

The preferred bacterial strains include ones which have the characteristics of

strain C42, I73 or H31 isolated by the present inventors. This *Xenorhabdus* strain has the following characteristics: rod shaped; motile; non-bio luminescent; blue on NBTA; produces antibiotics; resistant to ampicillin; forms circular colonies; has convex morphology; white colour.

This strain was presumptively identified as belonging to the genera *Xenorhabdus* since it was isolated from an insect killed by an entomopathogenic nematode and had the above characteristics. The strain has been deposited at the NCIMB (23 St Machar Drive, Aberdeen, AB24 3RY, Scotland) by the applicants under accession number NCIMB 41004 on 20 January 1999.

Further preferred strains of the present invention are two strains of *X. bovienii* designated H31 and I73 which have also been deposited under the terms of the Budapest Treaty at the NCIMB under the accession numbers NCIMB 40985 and 40986 respectively. These share characteristics of C42 in that they are rod-shaped; motile; non-bioluminescent; blue on NBTA; produce antibiotics; resistant to ampicillin; form circular colonies; and have convex morphology. The strains were identified as belonging to the species *X. bovienii* when compared to the *X. bovienii* type strain T228 using Restriction Analysis of the complete 16S rRNA gene and partial sequence analysis.

#### Target nematodes and diseases

The group of diseases described generally as helminthiasis is due to infection of an human or other animal host with parasitic worms known as helminths. Helminthiasis is a prevalent and serious economic problem in domesticated animals such as swine, sheep, horses, cattle, goats, dogs, cats and poultry. Among the helminths, the group of worms described as nematodes causes



widespread and often at times serious infection in various species of animals. The most common genera of nematodes infecting the animals referred to above are *Haemonchus*, *Trichostrongylus*, *Ostertagia*, *Nematodirus*, *Cooperia*, *Ascaris*, *Bunostomum*, *Oesophagostomum*, *Chabertia*, *Trichuris*, *Strongylus*, *Trichonema*, *Dictyocaulus*, *Capillaria*, *Heterakis*, *Toxocara*, *Ascaridia*, *Oxyuris*, *Ancylostoma*, *Uncinaria*, *Toxascaris*, *Caenorhabditis* and *Parascaris*. Certain of these, such as *Nematodirus*, *Cooperia*, and *Oesophagostomum*, attack primarily the intestinal tract, while others, such as *Dictyocaulus* are found in the lungs. Still other parasites may be located in other tissues and organs of the body.

The bacteria and encoded toxins of the invention may be used as nematocides for the control of the nematodes and diseases discussed above. More preferably, however, they are used to control soil and plant parasitic nematodes. Particular crop species which can be protected include tomatoes, potatoes, sugar beet, barley, soybean, peanut, onion, rye, wheat, corn, banana, raspberry, beans. Decorative and other plants may also be treated e.g. rose.

Target nematodes may be selected from the genera *Aphelenchoides*, *Anguina*, *Bursaphelenchus*, *Criconemella*, *Meloidogyne*, *Ditylenchus*, *Globodera*, *Helicotylenchus*, *Heterodera*, *Pratylenchus*, *Radopholus*, *Roteltylenchus*, *Tylenchus*, *Trichodorus*, *Xiphenema*. A further organism used in certain of the Examples below is *Caenorhabditis elegans*. Other target organisms and plants are discussed by Agrios in "Plant Pathology - 3rd Ed" Pub Academic Press Inc, see Chapter 15 therein.

As stated above, the target nematode will generally be different to that with which the bacterial strain is found in nature.

### Methods of use of bacteria

The bacteria may be used in any appropriate method which brings them into contact with the target nematode, preferably such that they, or their products, are ingested or absorbed by the target nematode.

In particular, regarding plants, the bacteria may be formulated in a variety of ways so as to enhance stability. For instance they may be employed in admixture with substrates to protect the cells.

The mixture can be spread over, ploughed into or otherwise mixed with nematode infected or potentially infected soil.

Regarding animals, bacteria intended for enteric inoculation can be mixed with carrier material that is suitable for ingestion by the intended animals.

### Isolation of agent

Nematode control agents of the present invention, which may be proteinaceous, or nucleic acids encoding them, may be isolated and/or purified from the C42, I73 or H31 bacteria described above, in substantially pure or homogeneous form, or free or substantially free of other materials from the bacterial strain of origin. Where used herein, the term "isolated" encompasses all of these possibilities.

Methods of purifying proteins from heterogenous mixtures are well known in the art, e.g. selective precipitation, proteolysis, ultrafiltration with known molecular weight cut-off filters, ion-exchange chromatography, gel filtration, etc. A particularly useful initial technique in this regard is ultracentrifugation. Further methods which are known to be suitable for

protein purification are disclosed in "Methods in Enzymology Vol 182 - Guide to Protein Purification" Ed. M P Deutscher, Pub. Academic Press Inc. Other references which outline techniques commonly used by those of ordinary skill in the art include "Protein Purification - principles and practice" Pub. Springer-Verlag, New York Inc (1982), and by Harris & Angal (1989) "Protein purification methods - a practical approach " Pub. O.U.P. UK.

Nematocidal activity may be assessed using a spread assay as discussed below.

The C42, I73 or H31 agent may be wholly or partially synthetic. In particular they may be recombinantly produced from nucleic acid sequences which are not found together in nature (do not run contiguously) but which have been ligated or otherwise combined artificially.

For instance, in the Examples below, nucleic acid encoding toxin(s) from I73 has been expressed in hosts cells using a vector system. Amino acid sequences of 38 different putative I73 toxin(s) are set out in sequence Annex 1. These sequences are based on the nucleic acid sequence set out in Fig 2 ('chrim5'), a cosmid clone derived from I73 genomic DNA which conferred nematocidal activity upon *E. coli* cells into which it was introduced (i.e. significantly reduced nematode larval growth and development, and feeding). As detailed below, the entire amino acid sequence as set out in each case may not be required for nematocidal activity. In particular the portion up to the first Met in each sequence may be omitted, as may other portions which may not contribute to the nematocidal activity. Thus, not all the proteins or genes may be required for nematocidal activity, and usually there will be one or more principal proteins, though others may play supporting roles such as in enhancing the activity or encoding other nematocidal activities.

Thus isolated nematocidal agents comprising a polypeptide containing all, or a nematocidal fragment, of any of the depicted I73 sequences, form one aspect of the present invention. Preferred agents include those encoded by p14-2f and p13-1f. Other active variants of these sequences are also encompassed as described below.

Candidate agents for use in this invention to control nematodes extend to those from the bacteria described in PCT/WO 99/22598, as well as the insecticidal toxins and bacteria of PCT/WO 99/42589, PCT/WO 98/08388 and PCT/WO 97/17432, the disclosures of which are incorporated by reference.

#### Nucleic acids and variants

In one aspect of the present invention there is provided a nucleic acid molecule encoding a nematode control agent of the present invention, for example a toxin, as described above.

The nucleic acid may be derived from the sequence shown in Fig 2 or the complement (or degenerate equivalent) thereof. This sequence (cHRIM5) was itself derived from I73 and identified by its unexpected nematocidal activity. Regions of this sequence believed to correspond to genes of the present invention are described in Fig 3. Isolated nucleic acids comprising one or more of these regions which encode a nematocidal activity are particularly preferred.

In the light of the present disclosure, further nucleic acids of the present invention may be isolated using PCR or southern blotting or other techniques well known to those skilled in the art. This requires the use of two primers to specifically amplify target nucleic acid, so preferably two

nucleic acid molecules with sequences characteristic of the C42, H31 or most preferably an I73 toxin isolated as above are employed. Using RACE, PCR, only one such primer may be needed (see "PCR protocols: A Guide to Methods and Applications", Eds. Innis et al, Academic Press, New York, (1990)).

Thus a method involving use of PCR in obtaining nucleic acid according to the present invention may include:

- (a) providing a preparation of bacterial nucleic acid,
- (b) providing a pair of nucleic acid molecule primers suitable for PCR, at least one of said primers being a primer based on a toxin from C42, H31 or I73,
- (c) contacting nucleic acid in said preparation with said primers under conditions for performance of PCR,
- (d) performing PCR and determining the presence or absence of an amplified PCR product. The presence of an amplified PCR product may indicate identification of a variant.

In a further aspect of the present invention there are disclosed nucleic acids which are variants of the C42, I73 or H31 toxin. A variant nucleic acid molecule shares homology (or identity) with all or part of the C42, H31, or most preferably I73 sequence discussed above.

Preferably sequence comparisons are made using FASTA and FASTP (see Pearson & Lipman, 1988. Methods in Enzymology 183: 63-98). Parameters are set, using the default matrix blosum62, as follows:

Gapopen (penalty for the first residue in a gap): -12 for proteins / -16 for DNA

Gapext (penalty for additional residues in a gap): -2 for proteins / -4 for DNA

KTUP word length: 2 for proteins / 6 for DNA.

16

Homology (similarity or identity) may be at the nucleotide sequence and/or encoded amino acid sequence level. Preferably, the nucleic acid and/or amino acid sequence shares at least about 70%, 75%, 80%, or 85% homology, most preferably at least about 90%, 95%, 96%, 97%, 98% or 99% homology.

Another method for assessing homology at the nucleic acid level is by hybridization screening. One common formula for calculating the stringency conditions required to achieve hybridisation between nucleic acid molecules of a specified sequence homology is shown in Molecular Cloning: a Laboratory Manual: 2nd edition, Sambrook et al, 1989, Cold Spring Harbor Laboratory Press:

$T_m = 81.5^{\circ}\text{C} + 16.6\text{Log} [\text{Na}^+] + 0.41 (\% \text{ G+C}) - 0.63 (\% \text{ formamide}) - 600/\text{\#bp}$   
in duplex

As an illustration of the above formula, using  $[\text{Na}^+] = [0.368]$  and 50-% formamide, with GC content of 42% and an average probe size of 200 bases, the  $T_m$  is  $57^{\circ}\text{C}$ . The  $T_m$  of a DNA duplex decreases by 1 -  $1.5^{\circ}\text{C}$  with every 1% decrease in homology. Thus, targets with greater than about 75% sequence identity would be observed using a hybridization temperature of  $42^{\circ}\text{C}$ . Such a sequence would be considered substantially homologous to the nucleic acid sequence of the present invention.

Variants of the present invention can be artificial nucleic acids.

Alternatively they may be novel, naturally occurring, nucleic acids, isolatable using the information disclosed herein. Thus a variant may be a distinctive part or fragment (however produced) corresponding to a portion of the C42, I73 or H31 toxin. The fragments may encode particular functional parts of the agent or they may be used for probing for, or amplifying, sequences corresponding to C42, I73 or H31 toxin. Sequence variants which occur naturally may include homologs of the C42, I73 or H31 toxin from other

bacteria, including nematode-symbionts. Artificial variants (derivatives) may be prepared by those skilled in the art, for instance by site directed or random mutagenesis (i.e. nucleotide addition, deletion or substitution, optionally to lead to amino acid addition, deletion or substitution) or by direct synthesis. Preferably the variant nucleic acid is generated either directly or indirectly from an original nucleic acid encoding the C42, I73 or H31 toxin.

Changes may be desirable for a number of reasons, including introducing or removing the following features. Sites which are required for pre- or post-translation modification. Changes for codon usage preferences to enhance gene expression in different organisms. Leader or other targeting sequences (e.g. membrane or golgi locating sequences) may be added to the expressed protein to determine its location following expression. All of these may assist in efficiently cloning and expressing an active polypeptide in recombinant form. Other desirable mutation may be random or site directed mutagenesis in order to alter the activity (e.g. host specificity) or stability of the encoded polypeptide. Changes may be by way of conservative variation, i.e. substitution of one hydrophobic residue such as isoleucine, valine, leucine or methionine for another, or the substitution of one polar residue for another, such as arginine for lysine, glutamic for aspartic acid, or glutamine for asparagine. Also included are active (nematocidal) variants having non-conservative substitutions.

Variant nucleic acids encompass all of these possibilities. When used in the context of polypeptides or proteins they indicate the encoded expression product of the variant nucleic acid i.e. variants of C42, I73 or H31 toxin e.g. variants of the I73 toxin sequences disclosed hereinafter.

Vectors and production of host cells

In one aspect of the present invention, the nucleic acid encoding the nematode control agent is provided in the form of a recombinant and preferably replicable vector.

Generally speaking, those skilled in the art are well able to construct vectors and design protocols for recombinant gene expression. Suitable vectors can be chosen or constructed, containing appropriate regulatory sequences, including promoter sequences, terminator fragments, polyadenylation sequences, enhancer sequences, marker genes and other sequences as appropriate. For further details see, for example, Sambrook et al (1989) *supra*.

The permitted vectors include, *inter alia*, any plasmid, cosmid, phage or *Agrobacterium* binary vector in double or single stranded linear or circular form which may or may not be self transmissible or mobilizable, and which can transform a prokaryotic or eukaryotic host either by integration into the cellular genome or exist extrachromosomally, e.g. an autonomous replicating plasmid with an origin of replication. Illustratively integration can occur into chloroplast DNA or into mitochondrial DNA.

Preferably the nucleic acid in the vector is under the control of, and operably linked to, an appropriate optionally inducible promoter or other regulatory elements for transcription in a host cell such as a microbial, e.g. bacterial, yeast, filamentous fungal or plant cell. The vector may be a bi-functional expression vector which functions in multiple hosts. In the case of genomic DNA, this may contain its own promoter or other regulatory elements and in the case of cDNA this may be under the control of an appropriate promoter or other regulatory elements for expression in the host cell. The vectors and host cells into which they are introduced may be used to clone or otherwise



identify nucleic acids according to the invention.

The agent may be used as part of a viral vector which is itself pathogenic to nematodes.

Also of interest in the present context are nucleic acid constructs which operate as plant vectors. Specific procedures and vectors previously used with wide success upon plants are described by Guerineau and Mullineaux (1993) (Plant transformation and expression vectors. In: Plant Molecular Biology Labfax (Croy RRD ed) Oxford, BIOS Scientific Publishers, pp 121-148). Suitable vectors may include plant viral-derived vectors (see e.g. EP-A-194809). Suitable promoters which operate in plants include the Cauliflower Mosaic Virus 35S (CaMV 35S). Other examples are disclosed at page 120 of Lindsey & Jones (1989) "Plant Biotechnology in Agriculture" Pub. OU Press, Milton Keynes, UK.

#### Host cells

The toxin genes or gene fragments encoding the nematocidal agents of the subject invention may be introduced into a host cell, microbial, animal or plant. Expression of the toxin gene in the host cell results, directly or indirectly, in the intracellular production and maintenance of the nematocide.

Thus the present invention also provides methods comprising introduction of such a construct into a plant cell or a microbial cell and/or induction of expression of a construct within a cell, by application of a suitable stimulus e.g. an effective exogenous inducer.

Hosts may be used to assay the activity of particular sequences or

fragments. Hosts can also be used to generate quantities of toxin which can be employed in situ in suitable treated cells, or alternatively with suitable hosts, e.g., *Pseudomonas* viable microbes can be applied to the sites of nematodes where they will proliferate and where they or their products can be ingested by the nematodes. Higher organisms, preferably plants, can also be engineered with the toxin. The result in each case is a control of the nematodes. A host may be selected that can tolerate harsh environmental conditions and then grow when they improve, as illustrated by *Bacillus* species where the spores can exist under environmental extremes.

Characteristics of interest for use as a nematocide microcapsule i.e. a vehicle for the active agent include protective qualities for the nematocide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to nematodes for ingestion; ease of killing and fixing without damage to the toxin; and the like.

#### Treated host cells

Where the cell is treated, the cell will usually be intact and be substantially proliferative form when treated, rather than in a spore form, although in some instances spores may be employed. Treatment of the microbial cell, e.g. a microbe containing the bacterial toxin gene or gene fragment, can be by chemical or physical means, or by a combination of chemical and/or physical means, so long as the technique does not deleteriously affect the properties of the toxin, nor diminish the cellular capability in protecting the toxin.

#### Viable hosts

Where the toxin gene or gene fragment is introduced via a suitable vector into a microbial host, and said host is applied to the environment in a living state, it is preferable that microorganism hosts are selected which are known to occupy the phytosphere (phylloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment (crop and other insect habitats) with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the nematocide from environmental degradation and inactivation.

A large number of microorganisms are known to inhabit the phylloplane (the surface of the plant leaves) and/or the rhizosphere (the soil surrounding plant roots) of a wide variety of important crops. These microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., genera *Pseudomonas*, *Erwinia*, *Serratia*, *Klebsiella*, *Xanthomonas*, *Streptomyces*, *Rhizobium*, *Rhodopseudomonas*, *Methylophilus*, *Agrobacterium*, *Acetobacter*, *Lactobacillus*, *Arthrobacter*, *Azotobacter*, *Leuconosroc*, and *Alcaligenes*; fungi, particularly yeast, e.g., genera *Saccharomyces*, *Cryptococcus*, *Kluyveromyces*, *Sporobolomyces*, *Rhodororula*, and *Aureobasidium*.

#### Plants as hosts

Nucleic acid encoding the nematocides of the present invention can be introduced into plant cells using any suitable technology, such as a disarmed Ti-plasmid vector carried by *Agrobacterium* exploiting its natural gene transfer ability (EP-A-270355, EP-A-0116718, NAR 12(22) 8711 - 87215 1984), particle or microprojectile bombardment (US 5100792, EP-A-444882,

EP-A-434616) microinjection (WO 92/09696, WO 94/00583, EP 331083, EP 175966, Green et al. (1987) Plant Tissue and Cell Culture, Academic Press), electroporation (EP 290395, WO 8706614 Gelvin Debeyser) other forms of direct DNA uptake (DE 4005152, WO 9012096, US 4684611), liposome mediated DNA uptake (e.g. Freeman et al. Plant Cell Physiol. 29: 1353 (1984)), or the vortexing method (e.g. Kindle, PNAS U.S.A. 87: 1228 (1990d). Physical methods for the transformation of plant cells are reviewed in Oard, 1991, Biotech. Adv. 9: 1-11.

*Agrobacterium* transformation is widely used by those skilled in the art to transform dicotyledonous species. It has also been used with filamentous fungi (see de Groot et al, 1998, Nature Biotechnology 16: 839-842).

Recently, there has also been substantial progress towards the routine production of stable, fertile transgenic plants in almost all economically relevant monocot plants (see e.g. Hiei et al. (1994) The Plant Journal 6, 271-282)). Microprojectile bombardment, electroporation and direct DNA uptake are preferred where *Agrobacterium* alone is inefficient or ineffective. Alternatively, a combination of different techniques may be employed to enhance the efficiency of the transformation process, e.g. bombardment with *Agrobacterium* coated microparticles (EP-A-486234) or microprojectile bombardment to induce wounding followed by co-cultivation with *Agrobacterium* (EP-A-486233).

Generally speaking, following transformation, a plant may be regenerated, e.g. from single cells, callus tissue or leaf discs, as is standard in the art. Almost any plant can be entirely regenerated from cells, tissues and organs of the plant. Available techniques are reviewed in Vasil et al., Cell Culture and Somatic Cell Genetics of Plants, Vol I, II and III, Laboratory Procedures and Their Applications, Academic Press, 1984, and Weissbach and

Weissbach, Methods for Plant Molecular Biology, Academic Press, 1989.

The generation of fertile transgenic plants has been achieved in the cereals: rice, maize, wheat, oat, and barley (reviewed in Shimamoto, K. (1994) Current Opinion in Biotechnology 5, 158-162.; Vasil, et al. (1992) Bio/Technology 10, 667-674; Vain et al., 1995, Biotechnology Advances 13 (4): 653-671; Vasil, 1996, Nature Biotechnology 14 page 702).

#### Combination nematocides

In further embodiments of the invention, bacteria associated with entomopathogenic nematodes or the toxins or products discussed above are used in conjunction with other nematocidal bacteria such as *B. thuringiensis* strains (e.g. from WO 92/19739) or pesticidal materials derived therefrom.

#### Materials for use in the present invention

The present invention also embraces materials for use in the methods above. These materials include the novel bacterial strains which are associated symbiotically with an entomopathogenic nematode and which are capable of controlling a target nematode. In particular the invention encompasses strain C42, I73 or H31 in isolated or substantially isolated form, or strains having the characteristics of C42, I73 or H31 (including nematocidal activity assessed as below).

Also embraced are compositions and formulations of these bacteria. These may comprise or consist of wettable powders, granules or dusts, mixed with various inert materials, such as inorganic minerals (phyllosilicates, carbonates, sulfates, phosphates, methylcellulose, xanthan gum and the like) or botanical materials (powdered corncobs, rice hulls, walnut shells,

peat moss, vermiculite, soil, seeds, other plant tissue and the like). The formulations may include spreader-sticker adjutants, stabilizing agents or surfactants. Liquid formulations may be aqueous-based or non-aqueous and employed as foams, gels, suspensions, emulsifiable concentrates, or the like. The ingredients may include rheological agents, surfactants, emulsifiers, dispersants, or polymers.

Bacteria may be mixed with other material while in freeze-dried form, encapsulated in biodegradable or water-soluble material, or otherwise treated to prolong their viability or decrease their levels of metabolic activity during handling. If desired, the carrier material may contain assimilatable nutrient sources to support proliferation of the bacteria.

Also included are purified or substantially purified nematocidal agents (particularly proteinaceous agents) isolated or isolatable from the strains or host cells discussed above.

Thus the invention further discloses nematocidal compositions comprising one or more agents as described above. Such compositions preferably further comprise other nematocidal materials from other *Xenorhabdus* species or non-*Xenorhabdus* species. These other materials may be chosen such as to have complementary properties to the agents described above, or act synergistically with it.

Toxins of the invention for use with animals can be adapted to be administered orally in a unit dosage form such as a capsule, bolus or tablet, or as a liquid drench when used as an anthelmintic in mammals, and in the soil to control plant nematodes. The drench is normally a solution, suspension or dispersion of the active ingredient, usually in water, together with a suspending agent such as bentonite and a wetting agent or like

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excipient. Generally, the drenches also contain an antifoaming agent. Drench formulations generally contain from about 0.001 to 0.5% by weight of the active compound. Preferred drench formulations may contain from 0.01 to 0.1% by weight, the capsules and boluses comprise the active ingredient admixed with a carrier vehicle such as starch, talc, magnesium stearate, or dicalcium phosphate. Where it is desired to administer the toxin compounds in a dry, solid unit dosage form, capsules, boluses or tablets containing the desired amount of active compound usually are employed. These dosage forms are prepared by intimately and uniformly mixing the active ingredient with suitable finely divided diluents, fillers, disintegrating agents and/or binders such as starch, lactose, talc, magnesium stearate, vegetable gums and the like. Such unit dosage formulations may be varied widely with respect to their total weight and content of the antiparasitic agent, depending upon the factors such as the type of host animal to be treated, the severity and type of infection and the weight of the host.

When the active compound is to be administered via an animal feedstuff, it is intimately dispersed in the feed or used as a top dressing or in the form of pellets which may then be added to the finished feed or, optionally, fed separately. Preferably, a carrier for feed administration is one that is, or may be, an ingredient of the animal ration. Suitable compositions include feed premixes or supplements in which the active ingredient is present in relatively large amounts and which are suitable for direct feeding to the animal or for addition to the feed either directly or after an intermediate dilution or blending step. Typical carriers or diluents suitable for such compositions include, for example, distillers' dried grains, corn meal, citrus meal, fermentation residues, ground oyster shells, wheat shorts, molasses solubles, corn cob meal, edible bean mill feed, soya grits, crushed limestone and the like.

Alternatively, the antiparasitic compounds may be administered to animals parenterally, for example, by intraluminal, intramuscular, intratracheal, or subcutaneous injection, in which event the active ingredient is dissolved or dispersed in a liquid carrier vehicle. For parenteral administration, the active material is suitably admixed with an acceptable vehicle, preferably of the vegetable oil variety, such as peanut oil, cotton seed oil and the like. Other parenteral vehicles, such as organic preparations using solketal, glycerol, formal and aqueous parenteral formulations, are also used. The active compound or compounds are dissolved or suspended in the parenteral formulation for administration; such formulations generally contain from 0.005 to 5% by weight of the active compound.

Further aspects of the invention include nucleic acids, vectors and host cells containing a heterologous construct according to the present invention, especially a plant or a microbial cell.

Such microbial cells may be treated as described in the methods above. Examples of chemical reagents are halogenating agents. Other suitable techniques include treatment with aldehydes, such as formaldehyde and glutaraldehyde; anti-infectives, such as zephiran chloride and cetylpyridinium chloride; alcohols, such as isopropyl and ethanol; various histologic fixatives, such as Bouin's fixative and Helly's fixative (See: Humason, Gretchen L., Animal Tissue Techniques, W.H. Freeman and Company, 1967); or a combination of physical (heat) and chemical agents that preserve and prolong the activity of the toxin produced in the cell when the cell is administered to the host animal. The method of inactivation or killing retains at least a substantial portion of the bio-availability or bioactivity of the nematode control agent.



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In all of the compositions discussed above, the nematocide concentration may vary widely depending upon the nature of the particular formulation, particularly whether it is a concentrate or to be used directly. The nematocide will be present in at least 1% by weight and may be 100% by weight. The dry formulations will have from about 1-95% by weight of the nematocide while the liquid formulations will generally be from about 16% by weight of the solids in the liquid phase. The formulations will generally have from about  $10^2$  to about  $10^{10}$  cells/mg, more preferably  $10^7$  to about  $10^9$  cells/mg. These formulations will be administered at about 50 mg (liquid or dry) to 1 kg or more per hectare. The formulations can be applied to the environment of the nematodes, e.g., plants, soil or water, by spraying, dusting, sprinkling, or the like.

In addition to the above the invention includes plant cells which have been transformed with the genes of the present invention, and plants which include such plant cells.

#### EXAMPLES OF THE INVENTION

The invention will now be further described with reference to the following non-limiting Figures and Examples. Other embodiments of the invention will occur to those skilled in the art in the light of these.

#### FIGURES

Fig 1 shows the cHRIM5 cosmid vector and subclones used for sequencing, as described in Example 6.

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Fig 2 shows the sequence of cHRIM5 (1-37544 bps).

Fig 3 shows the position and orientation of ORFs in the cHRIM5 sequence.

Fig 4 shows deletions of cHRIM5 tested for nematocidal activity.

Fig 5 illustrates cloning of nematocidal activity in PLEX.

Example 1 - Source of strains C42, I73 and H31

Strain C42 was obtained using an insect entrapment method. Insects which were killed on the surface of a soil sample were observed under a microscope at high magnification. Any that contained high numbers of bacteria and not fungal hyphae were presumed to have been killed by insect parasitic nematodes. The identified presence of nematodes also aids this identification step, but it is not essential. These samples were plated on to NBTA media (see Poinar & Thomas, 1984 Nematodes p238-280 in "Laboratory guide to insect pathogens and parasites" Eds. Poiner & Thomas, Pub. Plenum Press, New York). Any colonies that developed that had characteristic features (e.g. morphology, size, colour) of *Xenorhabdus* or *Photorhabdus* strains were selected. Non-luminescent colonies were presumptively identified as *Xenorhabdus*. The identity of those having nematocidal activity as assessed in Example 3, is further confirmed using 16s rRNA sequence data (see Brunel et al 1997, Applied and Environmental Microbiology 63,2: 574-580).

I73 and H31 strains were obtained in a similar way to strain C42 but they were identified as belonging to the species *X. bovienii* when compared to the *X. bovienii* type strain T228 using Restriction Analysis of the complete 16S

rRNA gene (see Brunel et al, 1997 Applied and Environmental Microbiology: 574-580), and partial 16s ribosomal RNA sequence analysis.

### Example 2 - Cell growth and preservation

Subcultures of the *Xenorhabdus* species C42, I73 and H31 were used to inoculate three 9 cm diameter petri dishes containing L agar (10g tryptone, 5 g Yeast Extract, 5 g NaCl and 15 g agar per lt). Plates were incubated for 48 hrs at 26°C and the resulting growth harvested by scraping off bacterial cells and thoroughly resuspending in 40 mls of 5% w/v lactose. The cells were washed once by centrifugation (5000 x g for 10 mins), resuspended in 10 mls of 5% w/v lactose, dispensed into 1 ml aliquots and freeze dried (-60°C for 48 hrs ) for medium term storage at 2°C. Other stocks were re-suspended in nutrient broth containing 10% w/v glycerol (Protect) and frozen at -70°C.

### Example 3- Activity of cells against *Caenorhabditis elegans*

The bioassays were performed by allowing *C. elegans* to feed on live bacterial cell suspensions spread over the surface of Luria broth agar (Luria broth containing 1.2%w/v agar) in segmented square petri dishes (2.0 x 2.0 cm per test well). A minimum of three test wells, each containing 50-100 nematodes were used for each test. Mortalities were recorded after 3 days at 18°C.

*C. elegans* was cultured on *Escherichia coli* at 18°C on 9 cm diameter LB agar plates. Once the nematodes had colonised the complete plate they were re-subbed on a fresh plate to maintain stocks and the remainder re-suspended in 40 ml LB. The tube was allowed to stand for 15 min and the nematodes settled to the bottom. The concentrated nematodes were removed using a

sterile pipette and placed in 40 mls of fresh LB. The process was repeated 5 more times to wash the nematodes away from the *E. coli* cells. The nematodes were then diluted so that approximately 50 nematodes were present in 50  $\mu$ l of LB.

The *Xenorhabdus* cells used were cultured in LB at 30°C/100 rpm for 24 hours and 50  $\mu$ l spread on to the surface of each test well. The control *E. coli* cells were treated in a similar way but incubated at 37°C for growth. After application the wells were air dried for 30 min, and 50  $\mu$ l of the nematode suspension placed in each well. Again the wells were air dried for 30 min. Plates were incubated at 18°C with 80% relative humidity for 3 days.

*Xenorhabdus* spp. C42, H31 and I73 gave 95% mortality, as compared with no significant effect for certain other *Xenorhabdus* bacterial strains and *E. coli*. Thus these results clearly show that cells from *Xenorhabdus* C42, H31 and I73 are an effective nematocide.

#### Example 4 - Cloning of nematode active gene from I73

Total DNA was isolated from I73 using a Quiagen genomic DNA purification kit (cat no. 10243). To isolate DNA, cells were grown in Luria broth (10g tryptone, 5g yeast and 5g NaCl per lt) at 26°C with shaking at 200 rpm to an optical density of 1.5 A600. Cells were harvested by centrifugation at 4000 x g and the DNA isolated using Quiagen 100/G tips, as per manufacturer's instructions. The purified DNA was stored at -20°C in TE buffer (10 mM Tris, 1 mM EDTA, pH 8.0).

To obtain a representative I73 library, total DNA was partially digested with *Sau*3A. Approximately 25  $\mu$ g of DNA was incubated at 37°C with 0.25 units

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of enzyme. At intervals of 5, 15, 30, 45 and 60 minutes, samples were removed and heated at 65°C for 10 minutes. To determine the size of the resulting DNA fragments, the samples were separated on a 0.5% (w/v) agarose gel. The samples containing a dominant DNA fragment size of between 30 and 50 Kb were combined and treated with shrimp alkaline phosphatase (Boehringer) for 20 minutes at 37°C. The DNA was ligated into the *Bam*HI site of the Stratagene cosmid vector Supercos1 (scos) and packaged into the *Escherichia coli* strain XL Blue 1, using a Gigapack II packaging kit (Stratagene) following the manufacturer's instructions.

To identify individual cosmid clones with activity to *C. elegans*, single colonies were grown in individual wells of segmented square petri dishes on Luria agar, containing 50 µg/ml ampicillin at 30°C for 24 hours. To each well, approximately 50, mainly L4 and adult *C. elegans* larvae were added in 50 µl of Luria broth. The dishes were incubated at 18°C and examined after 6 days for nematode development.

A total of 600 clones were examined and one coded cHRIM5 was found, which caused significant reduction in larval numbers, with no live L4 and adult larvae observed compared to on average, greater than 40 in all other clones tested.

Example 5 - Activity of cHRIM5, C42, H31 and I73 against *C. elegans*

Clone cHRIM5 was grown in 50 mls LB containing 50 µg of ampicillin per ml at 30°C/200 rpm for 40 hours. C42, H31 and I73 were grown in 50 mls LB at 26°C for 48 hours/200 rpm. Cultures were centrifuged at 4000 x *g* for 10 minutes, washed once and resuspended in 5 mls of PBS (0.05 mM phosphate buffer, 0.125M NaCl). To determine activity, 300 µl of cells were added in triplicate, to 1.2 ml of PBS containing 25, mainly L4 and adult *C.*

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*elegans* larvae in multi well square dishes. As a control, an equivalent amount of XL 1 Blue *E. coli* cells containing Supercos 1 were used to determine nematode survival. The assays were incubated at 18 °C for 7 days before approximate nematode counts and observations were made.

#### Activity of cells on *C. elegans*

Cell line	No. and size of larvae/square	Cell turbidity
XL 1 Blue/Supercos 1	>100 (all stages)	Clear
XL 1 Blue/cHRIM5	<20(mainly small, L1,2 &3)	Cloudy
C42	<10	Cloudy
H31	<10	Cloudy
I73	<10	Cloudy

Thus cHRIM5, C42, H31 and I73 all gave a reduction in nematode numbers, and in particular cHRIM5 cells significantly reduced larval growth and development. All four strains caused a reduction in feeding (as indicated by the cloudy cell suspensions).

#### Example 6 - DNA and protein sequences

Plasmid and cosmid DNA for cloning was prepared using the QIAGEN midi system (tip 100, cat. No 12143). Cells were grown in Luria broth (Merck) at 37°C with shaking at 200 rpm for 18 hours. Cells were harvested by centrifugation at 6,000 x g and the DNA isolated as per manufacturers instructions. Restriction digestion (Roche, Life Technologies), dephosphorylation (Roche) and ligation (Life Technologies) were carried out using manufacturer's recommended conditions and as outlined by Sambrook et al. Transformation was accomplished using electrocompetant cells and a

BIO-RAD Gene pulser set at 12.5V cm<sup>-2</sup>. Two µl of DNA was used to electroporate 80 µl of early log phase *E. coli* DH5 alpha cells washed 3 times in sterile water (centrifugation at 6000 x g for 5 mins) and resuspended in 1/100th the original volume in 10% (v/v) glycerol. Luria agar containing either kanamycin or ampicillin at 50 µg ml<sup>-1</sup> were used to select clones where appropriate.

DNA sequence analysis of cHRIM5 was completed by sequencing a number of sub clones and primer walking, see figure 1 for the supercos vector, where the numbers are kbp. The sub clones used are as follows:

code	cHRIM5 treatment	vector used or remaining
A-380	<i>Hind</i> III digestion and self-ligation	deleted scos
B-387	<i>Bam</i> HI digestion and self-ligation	pUC 19- <i>Bam</i> HI digestion
C-381	<i>Sal</i> I- <i>Bam</i> HI digestion	scos
E-391	<i>Sal</i> I- <i>Bam</i> HI digestion	pUC 19- <i>Sal</i> I <i>Bam</i> HI digestion
F-392	<i>Sal</i> I- <i>Bam</i> HI digestion	pUC 19- <i>Sal</i> I <i>Bam</i> HI digestion

Sub clone A-380 was constructed by digesting cHRIM5 DNA with the restriction enzyme *Hind*III and re-ligating fragments, this clone contains a deletion of the insert and scos cosmid DNA as the vector. Sub clone B-387 is a *Bam*HI digestion of cHRIM5 cloned into the plasmid pUC19 also cut with *Bam*HI and dephosphorylated. Sub clone C-381 was obtained by digesting cHRIM5 DNA with *Bam*HI and re-ligating the fragments, this clone contains the scos cosmid as the vector. Clones E-391 and F-392 were obtained by cutting cHRIM5 DNA with *Sal*I and *Bam*HI and ligating these fragments into the vector pUC19 also cut with these enzymes.

Sequencing was conducted using the artificial transposon AT2 (supplied by Perkin-Elmer-Applied Biosystems, Primer Island Transposition kit, cat No.

403015) using the cosmid cHRIM5 and all sub-clones as target DNA. One  $\mu\text{g}$  of cHRIM5 DNA was incubated with the transposon AT2 for 1 hour at  $30^\circ\text{C}$  in a final volume of  $20\ \mu\text{l}$ . After incubation the reaction was stopped by adding  $5\ \mu\text{l}$  of  $0.25\text{M}$  EDTA,  $1\%$  (w/v) SDS, and heat treatment at  $65^\circ\text{C}$  for 30 mins. The DNA was desalted by dialysis against water. One  $\mu\text{l}$  of the reaction mix was used to electroporate  $80\ \mu\text{l}$  of early log phase *E. coli* DH5 alpha cells. Colonies were selected on LB media containing  $50\ \mu\text{g/ml}$  trimethoprim. Once inserted the transposon mutants were used to provide a range of positions of primer sites at random intervals throughout the clones. The two primers PI+ and PI- near the end of the transposon were used to generate sequence data. In addition standard primers for the pUC19 and scos vectors were used to generate sequence data at the ends of each clone. DNA for sequencing was prepared using the QIAGEN ion exchange media (qiawell8, cat. No. 17122). Clones were grown in  $1\ \text{ml}$  of Luria broth containing trimethoprim ( $50\ \mu\text{g ml}^{-1}$ ) for 18 hours. Cells were centrifuged at  $13,000 \times g$  for 5 mins and resuspended in  $350\ \mu\text{l}$  of buffer P1. After 5 mins  $350\ \mu\text{l}$  of buffer P2 was added and the samples incubated for 5 mins at room temperature. To this  $350\ \mu\text{l}$  of buffer P3 was added and the samples left on ice for 15 mins. After centrifugation at  $13,000 \times g$  for 15 mins the samples were loaded on the Qiagen column under vacuum, and washed with buffer QC. DNA was eluted with buffer QF ( $500\ \mu\text{l}$ ) at  $50^\circ\text{C}$  and isopropanol precipitated ( $0.8\ \text{vol}$ ). After centrifugation at  $13,000 \times g$  for 30 min, DNA was washed with  $70\%$  (v/v) ethanol and air dried for 10 mins. The final pellet was resuspended in  $10\ \mu\text{l}$  of water. Cycle sequencing reactions using the Perkin-Elmer Applied Biosystems division Big Dye reaction kit (cat No. 4303149) were prepared using standard conditions for plasmid and cosmid sequencing. Samples were analysed on ABI Automated Sequencers. DNA sequences were assembled using the DNA\* software. The complete sequence of cHRIM5 was obtained by primer walking to join the final DNA contigs together. The final sequence of cHRIM5ed2 is shown

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in Figure 2. Analysis of the DNA using the software Clone indicated a number of ORF illustrated in Figure 3 and 4. Corresponding protein sequences are also presented at Annex I.

#### Example 7 - Fragments that encode nematocidal activity

To identify smaller fragments that encoded nematocidal activity, a series of sub-cloning experiments were performed using *E. coli* DH5 alpha. Qiagen midi and miniprep methods, restriction and ligations were used as for previous examples. Nematicidal activity of all constructs was determined as described in Example 4. In Figure 4, we show the deletions of cHRIM5 tested for nematocidal activity. Restriction sites and genes are indicated. Size in base pairs indicated on the map line. A cHRIM5, B cHRIM6, C cHRIM7, D cHRIM8, E cHRIM8, F cHRIM10, G *NdeI* deletion of cHRIM8, H Approximate positions (arrows) of three AT2 transposon insertions (tn58, tn26, tn43) in cHRIM9.

The cosmid cHRIM5 (figure 4A) was digested with the enzyme *SaII* and religated. The resulting sub clone cHRIM6, illustrated in Figure 4B showed nematocidal activity. cHRIM6 was digested with the enzyme *SmaI* and religated, producing sub-clone cHRIM7 (Figure 4C). cHRIM7 was digested with *BglII* and the kanamycin resistance gene block (*nptII*, Pharmacia) cut with *BamHI* was ligated into it. After selection on LB containing kanamycin (50µg ml<sup>-1</sup>) and ampicillin (50µg ml<sup>-1</sup>) the clone was digested with *SaII* and religated, in effect creating a deletion from the *SaII* site to the *BglII* site of cHRIM6 to generate cHRIM8 (figure 4D). By cutting cHRIM8 with *NruI* a further deletion was made to create cHRIM10 (figure 4F). All the above clones maintained nematocidal activity.

Deletion of cHRIM8 with *NdeI*, removed a portion of the p14-2f gene (figure

4G), this reduced nematocidal activity. This indicates that the p14-2f gene or protein are important for nematocidal activity. Transposon mutagenesis of cHRIM9 (a clone very similar to cHRIM7 but deleted with *NarI* rather than *SmaI*) with the artificial transposon AT2 (Perkin Elmer Applied Biosystems) resulted in a number of inserts within this clone (figure 4H). Insert cHRIM9-tn43 was restriction mapped to an approximate position of bp 20,700 (on cHRIM5) within the p20-9r gene, this mutant retained nematocidal activity. This indicates that this gene is not essential for activity. Insert cHRIM9-tn58 mapped to an approximate position of bp 13,400 (on cHRIM5), within the p13-1f gene, nematocidal activity was reduced. This indicates that this gene, region of DNA or the blocking effect of the transposon in this position is important for activity. Insert cHRIM9-tn26 was restriction mapped to approximate position of bp 15,000 (on cHRIM5) within the p14-2f gene, nematocidal activity was reduced. This indicates that this gene, region of DNA or the blocking effect of the transposon in this position is important for activity.

Clone cHRIM6-tn43 was digested with *BglII* and *NofI* and cloned into the vector PLEX (Invitrogen cat. No. K450-01) cut with *BamHI* and *NofI*. The *E. coli* strain used was GI742 supplied by Invitrogen. The resulting plasmid insert (PLEX-*BglII*/tn43, Figure 5) places the p14-2f and p13-1f genes under the control of the bacteriophage Lambda  $P_L$  promoter. Figure 5 illustrates the cloning of DNA encoding nematocidal activity in the expression vector PLEX, where: A, plasmid clone; B, insert and gene locations; Tpr, trimethoprim resistance; Apr, ampicillin resistance;  $P_L$ , bacteriophage lambda  $P_L$  promoter; \*, plasmid joins to form a circular molecule; \*\*, incomplete genes. Selection of colonies on RMG media (described in the Invitrogen manual) containing ampicillin ( $50 \mu\text{g ml}^{-1}$ ) and trimethoprim ( $50 \mu\text{g ml}^{-1}$ ) prevents expression from the  $P_L$  promoter. Colonies were then cultured on LB containing Trimethoprim ( $50 \mu\text{g ml}^{-1}$ ) in  $2.0 \text{ cm}^2$  wells for

nematocidal tests. The clone was active. This indicates that genes within this fragment have nematocidal activity. The clone PLEX-*Bgl*II/tn43 was digested with *Cl*oI and religated, this resulted in a deletion of part of the p13-1f gene, this clone had reduced nematocidal activity indicating the importance of this gene.

All these results indicate that the genes and gene products of p13-1f and p14-2f are important for nematocidal activity. Other smaller genes within the *Bgl*II to *Nru*I sites of cHRIM10 and PLEX-*Bgl*II/tn43 may also be essential. In addition genes outside this region within the remaining cosmid clone (cHRIM5) may also encode products with nematocidal activity, or may enhance the nematocidal activity of genes in the smaller region (*Bgl*II-*Nru*I of cHRIM10 and PLEX-*Bgl*II/tn43).

#### Example 8 - Field trials

Activity of strains selected in accordance with the above methods, or from depositary institutions which include bacteria which in nature are associated symbiotically with entomopathogenic nematodes, may be further assessed in field trials as follows.

Symbiotic bacteria in the absence of their nematode host can be inoculated into one or more portions of a field which is infested with nematodes, or into containers containing unsterilised soil from such a field. The bacteria can be inoculated onto the roots of plants, or into seeds. Periodically treated and untreated areas or containers can be assayed for nematode larva, egg, or cyst counts and for the presence of the inoculated bacteria by methods well known to those skilled in the art. A reduction in the number of nematode counts in areas in which the symbiote bacteria are present indicates control of the nematodes otherwise found in the untreated areas or samples.

## Annex I - amino acid sequences

SEQ ID NO:1

P0-0f

ISWFATGIPTVDALLAEFWHGDKQAFPPFTCRFTHFDKQDVTLPSTEEAYWLHRA  
 LQGQPLHSEVYGDDGTAQAGIPYTVMDSRPQVRLLTGLPGNSPTVWPSVIEQRTWQYERI  
 ADDPQCHQOVVLNSDRYGFRETVDIAYPRRPKPAVSPYPDTLPATLFDSSYDEQQQQLR  
 LTFQRQHYHHLTDTEHQVLGLPDVMRSDAWGYPAAVRVPREGFTLEDLLAENSLIAPGTPL  
 TYLGHQRVAYTGTGTGTEEKPTROALVAYTETAVFDELALQAFNGTLSPEALEKKLIESGY  
 LSVPRFNTGAESAVVVARQGYTDYGGSEAFYRPLAQRRTVQIGKNTLHWDTHYCAVVRM  
 QDAAGLYTDAAYDYRFLTPVQITDANDNQQHITLTALGOVSSGRFWGTEEGTPQGYTPPE  
 DRPFTPPSSVAEALDLKPDLPVANCVMVYAPLSWMPLAHTYQEYIAGFTWQALLDAGVVTE  
 DKRVCALGFRRWVQRQGIVLNGQALADSREP VHVLT LATDRYDTPDQQLRKSVTYS DGF  
 GRLLQSAVYHAPGEAWQRAADGSLITDAKGAPLVAHTATRWAVSGRTEYDGKGQPVRTYP  
 PFFLNAWQYLSDD SARQDLNADTHRYDPLGREYQVRTAKGYLRONRLTPWFV VNE DENDT  
 LS

SEQ ID NO:2

P1-2r

YLPQRGQCDMLLVVIGIGYLNNGQEA VIIGGIRVQTRILHTDDRTVMGIPMEGVFANLH  
 RRPLSORTVKRLRPAVIGISLTGDPDRRFRGTGIEWAWN RQITRLD

SEQ ID NO:3

P2-0f

SHLPARYGGRLTTL SRKGEMTVNRGDNLHQKTPEVTVLDNRGLTVRELRYHRHFNTPPTT  
 DERITRHRFTLSGQLAHSIDPRLFDLQQT DNTVNPNTYDTALTGEVVRTRSDAGNDLI  
 LNDITGRPVLAINATEVTRTWQYENDTLPGRPLSITEQPAGEAGRITERFWAGNSQAEK  
 NSNLAGQCVRHYDTAGLNQTD SIALNGIPLSVTRQLLPDGTDADWQGNNEPAWNDRLAPE  
 NFTTLSTADATGAVLT TTTDAAGNLQRVAYDVAGLLTGSWLRLAGGTEQVIVKSLTYS AAG  
 QKLREEHGNGVVTTYTYEPETQRLVGIKTKRPOGHAQGTKVLQDLRYEYDPVGNVVKVTN  
 DAEVTRFWRNQRKVPENTYVYDSLYQLVSATGREMANIVQOSTLLPTPSLIDSSTYSNYS  
 RTYNYDRGDNLTQIRHSAPATGNSYTTDITVSDHSNRVLDLTDDPAKVDA LFTAGGHQ  
 IPLQFGQNLVWTPRGELLKVAPVVRDQGISDQESYRYDAASQRIIKTHVQQTANSSQAQS  
 TL YLPGLERHTTINGTTVKEVLHVITIGEAGRAQVRVLHWENGKPGAISNNQMRYSYDNL  
 IGSSGLEVDGDGQIIISMEEYYPYGGTAVWTARSQTEADYKTVRYSGKERDATGLYYYGYR  
 YYQPWAGSWLSADPAGTIDGLNLYRMVRNNPATLDDKNGLAPGNRYVFFFIHEDRIFRL  
 ASANVYRTEHNKSDIIAVVEDKALDSKLF TNSIEQFFKKPKGKAILKGSPDIKERLLNNI  
 VHDLSNMQVGDQLYVNAHGHS AKPFFYS DSGYSKIIMEQLQORGANYVAKDLVNKFKL PEN  
 ATIKISTCHSAEGKGAHITVTSTGTNEKMRYSSIIENKGEFSRSLAGTMENELIKLQPR

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VRGNVYGYLGATTFFYGAKNEKVIHLKDGNLTTGVHEGKLSMFTKKNRFSENIFGLKVKRS  
LTRTNFTGSGV

SEQ ID NO:4

p-2-9r

PAAEYVRDFTITCSVFPASRSQLPVSRPATSYATRCRLPAASVVVSTAPVASAVLRVVKF  
SGASRSFQAGSLFFCQSASVPSGSSWRVTDSGMPLSAILSVWFSPAVS

SEQ ID NO:5

P3-2r

QRALLNDIGHFAPGGTDQLIQAVIDIGVLRHHFLVAPEAGNLRIVRHFHHVPHRVVLIAQ  
VLQHLRPLCMSLWAFGFYANKALGLRLVGVGHHAVAVLFAQFLTRGGIRQGFHDNLLCP  
ARKFOPTASQQACYVIRHTLQVTGRIGGGQYRAGGIRRAQGGEVFRCQPVVPGGFIVSLP  
VCVRTIRQQIARDGQRYAVKRNTVRLVQSGGVIVTHALSGQVAVLLRLTVPCPKTLCDT  
ACFASRLPCDTERASG

SEQ ID NO:6

P3-6r

SDRRQTGYAYSADHYRISGRSTVCTVRAGLMNYQCWLQHAATQLSESDSPKRDAEILLGY  
VTGRSRTYLIADFDETLISSEELHQLDLSLLVRRIOGEPVAYIIGEREFWSLPFAVSPATLI  
PRPDTECLVEKALELLPDSPARILDLTGTGATIALALASERNDCYVTGVDINSDAVMLAQ  
HNAEKNAKGLAIHNVNLFQSEWFAAVGNQQQFDMIVSNPPYIDERDPHLQEGDIRFEPATA  
LIAAQNGMADLQAIVGQARHFLSPNGWLLLEHGWKQGTVVRNLFLEKGYQQIATFQDYGG  
NERITIGRWKNKETHS

SEQ ID NO:7

P3-7f

ARRAVRRCGYCTGRTESRVPSVTTRCATAMITLSAAAVWRWTVTDKLSVWKNTRTGALR  
CGRRGVRORLITRLCVTQARSGMQRGCIITATGITSRGRGAG

SEQ ID NO:8

P5-6r

WQNGSSSTTPRYLAGCYVWYPCSRILSGNAKSLLAPDGEWMKHTLKSKASGNTFTGRLI  
FTGRPTVVITDKSGANTAALTLLNAEGEPQQGIEIRQNKYLNNRIEQDHRHVKKRIRREML  
GFKSFRAQT

SEQ ID NO:9

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P5-7r

ALLFLSESRVMSLIRNAFKLLHYPVDIMAQCVRWSLTIALSLRNLEEMMAKRGIFVDHAT  
IPRWVLRRLVPLLSKAERKRKKPVGSRWRMDETYIKVKGQWKYLYRSVDTDGQTDGCDYR

SEQ ID NO:10

P6-3f

VHSPSGAVAPGKFFIENFADTFPAPLPLHPFIDACIQQGFQLLPCLIAIAHSGKQAFECV  
LLDRALQGSQCLQALVLPVGDVNGQTAHGFLIGYTQTHISTYNGLWLFITQGVRYRFV  
RQTFVCRSLSFSEDDCTN

SEQ ID NO:11

P6-3r

RTCRERPRLM DYVLT KAAEADLRATIRHTRKQWGDAQVRRYITALEQGIARLAVGQGSFK  
DMSALFPALRMAHCERHYVFCLPRENAPALIVAIFHERMDLLTRLADRLK

SEQ ID NO:12

P6-6r

PQTIICANVGLCITDKEKTMSRLTIDITDRQHQLKALAALQGKTIKQYALERLFPGMSD  
SDQAWQELKALLDTRINEGMEGKGCCKSIGEILDEELAGSDRA

SEQ ID NO:13

P7-1f

NAHFLIVSKTNVMSNQDPHNKRDSLFSAPIANLGDWSFDERVAEVFPDMVKRSIPGYSN  
IISMIGMLASRFVTPGSQIYDLGCSLGAATLSIRRSINADNCRIIAIDNSPAMIERCRRH  
IDSFKASTPVEVIEQNILDTDIQNASMVVLNFTLQFLHPDDROKILKKIYAGLKPGGVLV  
LSEKFNFEQKIGELLFNMHHDFFKRANGYSELEVSQKRSMLENVMRTDSVDTHKSRLKEV  
GFQHVVEVWFQCFNFGSLLAIKGTEQ

SEQ ID NO:14

P7-9f

TMIDFGNFYQLIAKHPLNHWLDSLPAQLSHWQKTSQHGFSSWVKILENLPEIKPSHLDL  
KNGVIAIHEPDLKSGEKARLHNILKILMPWRKGFPSLYDVEIDTEWRSDWKWERVLP  
PLEGKTVLDVCGSGYHMMWRMVGEQAQLVVGIDPTQLFLCQFEAIRKLLGNNQRAHLLPL  
GIEQLPELQAFDVTFSMGVLYHRRSPLDHLWQLKNQLVSDGELVLESVIEGDENQCLIP  
GERYAQMARNVYFIPSAKMLKVWLEKCGFVDVRIVDHAATTPDEQRRTIEWMKTESLVDFLD  
PSDHSKTIEGYAPLRAVLIARKP

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SEQ ID NO:15

P8-4r

SLQIDREKVGLDRYPQPIERLRQPCATCDNHCHSRHQVRFFLLKEYGAALAPISSQSAI  
RYQFORHTMKKGLFAMASIFSGYCGGELFHLLTDPAHESQ

SEQ ID NO:16

P9-8r

SSFRLNDLLTNSYSEGFLMIKLEICCYSSISCALVAQNAGADRIELSASPLEGGLTPSFG  
ALQQSLQRLSIPVHPVIRPRGGDFCYNMDFEAMKNDVARIRDMPGIVFGILSENGHI  
DRLMRQLMSLSGNMAVTFHRAFDNCFNPHVALEQLTELGVQRILTSGQQONAEGLTL  
KELMQASRGPIIMPAGVRVSNISKFLEAGMTEVHSSAGKIVPSTMKYRKVGVMSSDDR  
DVDEYSHYSVDGELVESMKGVMSLIKR

SEQ ID NO:17

P10-5r

YFGKNRRFVIYVTLMERNFYGLFNGEEMSHFSKISELQDLVADLAGFEQKLKQFEGHLGL  
HFEQYSADHISLRNESKIADRWKGFLOCGQLISESIINGRPICLFDLNQPIVLLDWKI  
DCVELPYPSQKHVYHQWEHVELVLPVPPEQLICEAKLLPQPLPDNFRMKESHPPKGKNE  
RLPNPILAV

SEQ ID NO:18

P10-7f

GNTVNIQVILSEKISNALIEAGAPTDSEAHVRSQAKAQFGDYQANGVMAAAKKVGIPPRQ  
LAEKVVSQDLQGIASKVEIAGPGFINIFLDKAWVAANIETTLKDEKLGITPVEPOTIVI  
DYSAPNVAQMHHVGLRSTIIGDAAARTLEFLGHKVIRANHVGDWGTQFGMLIAYLEKIQ  
NENANDMALADLEAFYREAKKHUDEDEFAIRARNYVVKLOGGDEYCRKMWRKLVDITMS  
QNQETYNRLNVTLTEKDVMGESLYNDMLPGIVADLKQRGIAVKSDGATVVYLDEFKNKEG  
EPMGVIIQKKDGGYLYTTTDIACAKYRHETLNASRVLYYIDSRQHQLMQAWAIVRKTGY  
IPESMSLEHHMFGMMLGKDGKPFKTRAGGTVRLSDLLDEAIERADTLIREKNPDMPEDEL  
KKVVEAVGIGAVKYADLSKSRRTDYVFDWONMLAFEGNTAPYMOYAYTRVSSIFKRADID  
ENSLTLPVMLNEEREQALATRLQLQFEETITTVAREGTPHVMCAXYLDLAGLFSGFYEHCP  
ILNADSEELRQSRLLKALLTAKTLKQGLDTLGIQTVERM

SEQ ID NO:19

P11-1r

AQVSNMHLGDIRCGIIDNDGLRFHWGDTSELFIFQGSFYICCNPRFIKKNIDKTWACNFN  
FAGNSLOIQLADDFCQLSRRYSHLFGSGSHHTIRLIVTKLCFGRLTDVVSFTVGWSASFNQ  
RIADFF

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SEQ ID NO:20

P12-1r

HARVGVLHIRCRVAFKGOHIIPVENIVCSTALGKICIFHRANPYRFHDEFQFVFWHIWVF  
LTNEGIRTLNRFIQIGQSYCAAGTGFEWFTIFAQHHAKHVVE

SEQ ID NO:21

P12-5r

YHASFQLCRRLHTFYSLNTQSIKTLLOSFRCCQSQLQAALAQFFAIGIQDRAVLITRE  
QTGQIVQVCTHNMWRTFTGDGSDRFFKLQAGCQCLLAFFIQHHRQCQAVFIDIRTFKDR

SEQ ID NO:22

P13-1f

FTLREDSMSDWTGVSTFNVILETGLDNCNIYANGLNMIGVIINITPTDDEGNFVDIDVT  
LNDNIKIVDYIDGSDIDGSDGWFTYGNPNEYNTIPNSQSYSLKSENSQITQIKRYVSCS  
NTSRLRTKSFSKVTTSKGVISITQNSINSSRVVINAIDATNFTDDELRTTKETRFENQ  
SYTSHKSSTNSLYVHTWTIPRSLKLQNRWEDYNNNGWTWAQSCYYKTGADGGSESTRWLA  
AGSIFPPGNYDGLWLDNDIALSGMAHKSYNVDGTINQLSFTRIIGKGFVWYNISGLDRG  
HAVIIIDQYGNKYRILFHAGYENSDFYLSSSIVY

SEQ ID NO:23

P14-2f

VYIKFLKLFERRITMSDNNEFFTQANNFSAVSGGVDPRGTGLYNIQITLGHIVGNGNLGPT  
LPLTLSYSPLNKTDIGFGIGFNGFGLSVYDRKNSLLSLSTGENYKVIETDKTVKLQKKLD  
NLRFEKDLKENCYRIIHKSGDIEVLTFGNNAFDLKVPKLLNPAGHAIYIDWNFEATQP  
RLNRIYDDLDGHDIPLLNLEYQGLIKTILTLFPGQKEGYRTELRLNRQLNSIHNFSIGN  
ENPLTWSFGYTFIGKNGILGQWITSMTAPGGLKETVNYSNNNQGHFPQSANLFLVLPYVT  
LMKQVPGAGQPAIQAESYTSNHYVGGGSGNGIWNKLDNLYGLMTEYNYGSTESRRYKDK  
EGHDQIVRIERTYNNYHLLTSECKQONGYIQTETAYYAIIGHNFDSQPSQFQLPKTKTE  
TWRSADNSYRSEITETTDESIGNPLTKVIKDKKTOKIISPSTHWEYPPAGEVDNCPPEP  
YGFTFRVKKI IQTPYDSEFKDDPEKFIQYRSLIGSQSHVTLKIEERHYSATQLLNSTLF  
QYNTDKSELGRLLKQTECTKGENGKTYSVVHKFTYTKQDDTLQOSHSTHNDFTIHRSQ  
VRSRYTGRLFSDTDTKDIVTQMSYDKLGRLLTRTLNSGTFYANTLTIDYELNNLQDDNRP  
PFVITTTDVNGNQLRNEFDGAGRHSVQCLKDSGDGKFTYIHTQQYDEQGRHHTSTYSYD  
LTNGRQOTDPKVLHLSMSKSYDNWQIANHWSYGVSEKITVDPITLTATKQLQSNNSNV  
QTGKEVTTYTPSQQPIQITLFDLQAGHLQSCHTLTRDGNDRVRKETDAIGQCTIYQYDNYN  
RVIQITLPGTIVNRKYAPFSTDTLITDIRVNGISLGQOTFDGLSRLTQSQDGGRVWAYT  
YSAGNDQCPSTVITPDGQFIHYQYQPELDDAVLQVASNEITQQFSYNPVTGALLKAVAEG  
QSLTPIYYPSGRKLMENINDMKMSYLTWTLRGLNGYTDLTGTIQKISRDTHGRVTQIKD

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SSIKTTLNYYDDLNRHIGSQVTDLATGHMLTTTVEFDGLNREIGRKLCDSSSGHTLDIQQSW  
 LKTQQLANRIVKLNGLVLRTEQYSYDSRNLNOYKCDGAECPTDKYGHISIVTONFTYDIY  
 GNITACHTTTFADGTEDHATFKFANPTDPCQLTEVHHTHPDMPDNIRLKYDKAGRVINITD  
 NHGNTENFTYDTLGRQLONGQGSVYGYDPLNRLVSQKTDTLDCELYRETMLVNEVRNGEM  
 IRLRLTGETIIAQQRASKVLLTGTDSSQSVILTSKQNLSEAYSAYGKHKSTANDASIL  
 GYNGERADPVSGVTHLNGYRSYDPTLMRFHTPDSLSPFGAGGINPYSYCLGDPINRSDP  
 SGHLSWQAWTGIGMGIAGLLLTATGGMAIAAAGGIAAAIASTSTTALAFGALSVTSDIT  
 SIVSGALEDASPKASSILGWVSMGMGAAGLAESAIGGTKLATHLGAFEDGENALLKST  
 SESSRIKWGVTRSLDREIVRNEEGOVIKDHSRGYTDNFMGKGEQAILVHGDGKDFLYHTE  
 GNMKNGKGPYTRHTPEQLVDYLKDNINVDLTQGGDKFVHLLSCYKSSGAADKMAKYINR  
 PVIAYS NKPTISQGLARIERKDFFLKSTYHSYDPRKIILGRTEKTVKPKTFRP

SEQ ID NO:24

P17-6r

LCYGHICLSGIPHRHIYIGSTYYGNRKSTVLYAAILHSVSLFYLLIAVFSASSAGYLTYG  
 LSYHTISVQFLGLSHQIFLLSTYDQSLNLLLDYQYGDSGHRNLE

SEQ ID NO:25

P17-8r

SAQCIVGKVFRISMVISDIYYSTSLIIFQPDIIIRHIWMSVVYLCQLAWVSWVGKFEQSMV  
 FCPICECGVTGGDIAIDIISKILCDYAMAFVCFRAFRTVTFILVQPIVGIVRVLFCTLOQ  
 SIQFHYISIC

SEQ ID NO:26

P18-7r

PSSLRTISLSKLLVTPHFILELSEVDLSKAFSPSSANAPRCVASLVPPLMADSANPAAPI  
 PIETHPSIEDAFGEASSAPLTIDVISDVTLAPNASAVVEVEAIAAAIPPAIAIAPPV  
 AMVSSNPAIPMPFVHACQLK

SEQ ID NO:27

P19-5f

AHCHIALFPCWHNPQYCOQHPDHHSNCHHQFKQEYPPSRQARENITLTQLPIKHTGIEAG  
 SQTNRKQOTCMFQRANESKVHQLGQNGQRDRNFYWCEDILT

SEQ ID NO:28

P19-8f

PQSTPSSQNSRQLTPAESSQHOKOKSDHIEIMIPSEAPREYREQLHKATPARNROVAPNP  
 SVFDILRDYHWKNFSPVKAAKSSLTPHFVHQKAIPLNDQRNTSMKQSLKPEMRQKLY

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FOOEEI " 42353360

SEQ ID NO:29

P20-1r

GKNCINDQGNLPDRYTQNCRPHLTDNPPYGTVTERNPRQYQHADLFQMRKLIGQLQNP  
 NNGPTQRQHWRIAIRSHKQCKNDHTDIEQCRSKSRHRKAVPCIKNCASQSQRNQKDIRK  
 RNSK

SEQ ID NO:30

P20-9r

NNTMNLKSLAAVSSMTMFSRVLGFIIRDARIIFGAGMATDAFFVAFKLPNLLRRIFAE  
 GAFSQAFVFPILAKEYKNQOGDEATRTFIAYISGMLTLILAIVSVIGVIAAPWIIYVTAPGF  
 TDTPDKFVLTRDLLRITFPYIFLISLASLAGAILNTWNRFSVPAPFAPTLNVSMIIFALF  
 VAPYCNPPVLALGWAVVAGGVQLAYQLPHLKKIGMLVLPRI SFROSAVVRVIRQMGPAI  
 LGVSVGQISLIINTIFASFLVSGSVSWMYADRLMELPSGVLGVALGTILLPSLAKSFSS  
 GNHEEYRKLMDWGLRLCFLALPCAVAGILAEPLTVSLFQYGHFSAFDAEMTORALIAY  
 CFGLMGLIVVKVLAPGFYSRQDIKTFVKIAIATLIETQLMNLAFVGPLKHAGLALSIGLA  
 ACENASMLYWQLRKRDIFTPLAGWGI FLFKLVVAIAVMVGVLAVLVWMPAWEQGNMAMR  
 LLRLMGVVIAGAGSYFAVLALMGFRLKDFAHRLGLO

SEQ ID NO:31

P21-7r

AIILIRDKLSRIFSRQISGEGMFGYRSASPKIRFITDRMVRLVYERDAYRLAEYYSENK  
 DFLKPWEPTRDGSFYQPSGWTNRLNYIAELQRQNTFNFVLLDSDEREIMGVANFTNVVR  
 GAETHSCYLGYS LAEKLOGQGLMYEALQPAIRYMORYORMHRIMANYMPHNHRSGNLLKKL  
 GFEQEGYAKNYLMIDGVWQDHVLTALTDDAWGKVGL

SEQ ID NO:32

P21-8f

WCAMSLVSQARSLGKYFLLFDNLLVVLGFFVVEPLISIRFVEQLGWAALIVGFALGLRQL  
 VQOGLGIFGGAIADRFGAKPMIVTGMLLRALGFALMAMAHEPWILLSCVLSGLGGTLFD  
 PPRAALVIKLT RPHERGRFYSILMMQDSAGAVVGALIGSWLLQYDFNIVCWIGASIFVLA  
 ALFNAWLLPAYRISTIRTPIKEGMMRVIRDORRFLYYVLTLTGYFVLSVQVMLMFPIIHE  
 ITGTPTAVKWMYAIETAISLTLLYPIARWSEKHFRLEQRLMAGLFLMSICMPPIGWVNQL  
 HTLFGLLCLFYLGVLVTADPARETLSASLSDPRARGSYMGSRLGLALGGAIGYTGGGWLY  
 DTGRDLNMPQLPWILLGLSGLITIYALHRQFNQKKIDPVMLGRH

SEQ ID NO:33

P23-1f

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KGANMKRFFLGAALVLVGLVSGCDQFKDFSINEGLMNDYLLKKVHYQKKISIPGIANANI  
 TLGDLSSQIGRODPEKIELSTQAKVOLATLLGTIQADMKI.TIKAKPVFDAEKGAIFVKGL  
 EIVDYQTTPEKAAAPVKALIPYLNLSLSEFFDTHPVYVLNPEKSKAEAAASQFAKRLEIK  
 PGKLVIGLTDK

SEQ ID NO: 34

P24-4r

QVALQHGRRLGTITLFDNLLGLNQVMNEFSIVCRILGTLENRAPQDFVLQPLITMIAEGK  
 LKQAWPLEQDEWLDRLQONSELSVMAADYHALFTGESASVAVCRSDYTDGEESEVRQFLT  
 ERGMPLSDTPADQFGSLLLAVSWLEDQAAEDEIQAQITLFDYLLPWCGQFLGKVEAHAT  
 SGFYRTLAIIVTREALQALRDELESE

SEQ ID NO: 35

P25-3r

DCMNIIFFHPSFNTDEWIOGQIARLPDAKVRQWVSGDQEPADYALVWQPPYEMLANRQGL  
 KGIFALGAGVDATFKQESKNPGTLLADVPLIRLEDTGMRQMQEYAITSVLHYFRMDEY  
 KRYQEQRLWNPIAPHNRKEFVIGVLGAGILGRSVIGKLMEFDFNVRCWSRTSKQLDSVES  
 FYGKEQLGDFLSGCKVLINLLPDTPDTRGILNLSLFSQLKSGSYVINLARGAQLVEQDLL  
 VAIDKGYIAGATLDVFAEEPLSNMHPFWTHPRINVTPHIAANTIPEAAMDVICENIRRMV  
 QGEMPTGLVDRVRGY

SEQ ID NO: 36

P26-0f

KTSQGFTSTTCSNGNVLKICGLITPCSSLIQRTYPNNMTIGIFSKESTAKNFGMGFLYYF  
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 RFSIENCFIKSG

SEQ ID NO: 37

P27-0r

YSAGCSTVLKSSLNLQCDTFNCESFVMLTLNFTSVNAKPSHIWAHYVDFDLRKKWEVDL  
 EYFQFEGEVKTGQYGRMILSGMPEIRFYLSNIEVNKEFTDQVNLPQMGIITFRHQIITDE  
 NNMACRVQVTVSFEPDANI PAVQAESFFKQGTQDLVESVLRKSVVETVSPKPNLQLVYV  
 SDIESSTAFYKTI FNAEPI FASSRYVAF PAGGEVLFAIWSGGAKPDRAIPRFSEIGIMLP  
 SGKDVDRCFEEWRKNPEIKIVQEPHTEVFGRTFLAEDPDGHIIRVCPLD

SEQ ID NO: 38

P27-8r

KGNQITMILYKGSKNYLFNQLNYDSCVLLLEVDES VN LNWDELSRAQRLLFLMEILARYH

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P2000-10300-4-285560

FFVQGVKVLAKLNI SLRTLYRDIASLOAQGAIIIEGEPGIGYVLRPGFVLPLMFTQNEIE  
ALALGANWVAKRADPQLKESANNAISKIAAVIPAELKQMLEASSLLIGPAATAVQPVVEI  
QQIROAINTRHKITLAYLDIKDIPSERTIWPFALGYFENISIVIGWCELREEFRRHFRSDR  
IMRLKIENQCYPRSRQVLLKEWRAMEKISR

P27-9f

SEQ ID NO: 40

GVMNTPQLOQRIAE EHYFTTSDNASLFYRYWPQQQANPDRAIIIFHRGHEHSGRIOHVVD  
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SVGAVLVS AWVHDYAPKIRAMILAAPAFDIKLYIPFATQGLQLMQKARGIFFVNSYVKAR  
YLTHDETRIASYNSDPLITREIAVNILLDLYQTAERVVKDAAAITLPTLLFISGSDYVVN  
KKPQH QFYQQLNTPIKEKHVMDGIFYHDTLGEKDRHLVFDKIRVFIERIFALPRYQHDYSQ  
EDTWSHSADEFRTLSTSLPCLCPKKLSYQLMRKVMSTHWGRTSEGVCIGLKTGFDSGSTL  
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P28-5bf

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P30-3f

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47

LPLAPEALQSAVCTLDKLRQQGDVFEVHCTLGLSRSAMVVAWLLKQHPEYDINTVVAILR  
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SEQ ID NO:43

P31-6f

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SEQ ID NO:44

P32-3f

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SEQ ID NO:45

P33-4r

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SEQ ID NO:46

P33-5f

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SEQ ID NO:47

P34-3f

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48

SEQ ID NO:48

P35-0r

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FPELDTSK

SEQ ID NO:49

P35-8r

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SEQ ID NO:50

P36-7r

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SEQ ID NO:51

P37-5r

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FPQGRVTDHRINLTLYRLDEVIEGKLDMLIQPIIIEYQADQLSALSEQD

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## Claims:

1. The use of a bacterial strain to control a target nematode, characterised in that in nature the bacterial strain is associated symbiotically with an entomopathogenic nematode.
2. The use according to claim 1, wherein the bacterial strain from nature is directly employed to control the nematode target, or is employed to give a recombinant bacterium employed to control the nematode target, or the natural or recombinant strain is employed as a source of a nematode control agent to control the nematode target.
3. The use according to claim 1 or 2, wherein the target nematode is not the same as the nematode with which the bacterial strain is found symbiotically in nature.
4. The use according to claim 1, 2 or 3, for control of helminthiasis in a human or a domesticated animal or the control of plant pathogen nematodes.
5. The use according to any preceding claim wherein the nematode to be controlled comprises one or more of *Haemonchus*, *Trichostrongylus*, *Ostertagia*, *Nematodirus*, *Cooperia*, *Ascaris*, *Bunostomum*, *Oesophagostomum*, *Chabertia*, *Trichuris*, *Strongylus*, *Trichonema*, *Dictyocaulus*, *Capillaria*, *Heterakis*, *Toxocara*, *Ascaridia*, *Oxyuris*, *Ancylostoma*, *Uncinaria*, *Toxascaris*, *Parascaris*, *Aphelenchoides*, *Anguina*, *Bursaphelenchus*, *Criconemella*, *Meloidigyne*, *Ditylenchus*, *Globodera*, *Heliocotylenchus*, *Heterodera*, *Pratylenchus*, *Radopholus*, *Rotelynchus*, *Tylenchus*, *Trichodorus*, *Xiphenema*, and *Caenorhabditis*.

6. A composition for the control of parasitic nematodes which comprises as an effective agent a species of bacterium which is a symbiont of an entomopathogenic nematode, or an engineered bacterium, or a nematode control agent derived from a natural or engineered bacterium.
7. A composition according to claim 6, wherein the bacterial species is of the genera *Xenorhabdus* or *Photorhabdus*,
8. A composition according to claim 7, wherein the bacterial species is of the genus *Xenorhabdus*
9. A composition according to claim 8, wherein the bacterial species is of , the species *Xenorhabdus bovienii*.
10. A composition according to claim 8, wherein the bacterial species is:  
*Xenorhabdus bovienii* strain H31 deposited with NCIMB under accession number NCIMB 40985;  
*Xenorhabdus bovienii* strain I73 deposited with NCIMB under accession number NCIMB 40986; and  
*Xenorhabdus* strain C42 deposited with NCIMB under accession number NCIMB 41004.
11. A composition according to any of claim 6, wherein the nematode control agent which is derived from a symbiont of an entomopathogenic nematode or from an engineered bacterium has functional activity against a nematode, and is a peptide.
12. A nucleic acid encoding a peptide of claim 11.
13. A nucleic acid according to claim 12, which nucleic acid comprises a



natural nucleotide sequence or a degeneratively equivalent sequence, or a functional variant thereof.

14. A nucleic acid according to claim 13, which is a homologous variant encoding a peptide which is a nematode control agent, the nucleic acid having 70% or more DNA sequence identity and/or the peptide having 70% or more amino acid sequence identity.
15. A nucleic acid according to claim 13, which is all or part of cosmid cHRIM5, in particular p 13-1f or p 14-2f, and variants thereof.
16. A nucleic acid according to claim 13, 14 or 15, wherein the variant has a sequence which is a derivative by way of addition, insertion, deletion or substitution of one or more nucleotides.
17. A nucleic acid according to any of claims 12 to 16, which is part of a longer sequence and the nematode control agent is expressed as a fusion protein.
18. A nucleic acid complementary to a nucleic acid according to any of claims 12 to 17.
19. A nucleic acid for use as a probe or primer having a nucleotide sequence of at least 15 nucleotides, which sequence is present in a nucleic acid according to any of claims 12 to 18.
20. A method for identifying or cloning a nucleic acid according to any of claim 12 for a nematode control agent, which method employs a nucleic acid probe according to claim 19.

21. A method according to claim 20, which comprises the steps of:
- (a) providing a preparation of nucleic acid from a bacterium,
  - (b) providing a probe,
  - (c) contacting nucleic acid in said preparation with said probe under conditions for hybridisation of probe to any said gene or homologue in said preparation, and,
  - (d) identifying said gene or homologue if present by its hybridisation with said probe.
22. A method according to claim 20, which comprises the use of two primers to amplify a nucleic acid encoding a nematode control agent, at least one of the primers having a conserved nucleotide sequence of at least 15 nucleotides.
23. A method according to claim 20, which comprising the steps of:
- (a) providing a preparation of nucleic acid from a bacterium,
  - (b) providing a pair of nucleic acid molecule primers, at least one of which is a primer,
  - (c) contacting nucleic acid in said preparation with said primers under conditions for performance of PCR,
  - (d) performing PCR and determining the presence of absence of an amplified PCR product.
24. A recombinant vector comprising a nucleic acid according to any of claims 12 to 17.
25. A host cell containing a vector according to claim 24 capable of replication.
26. A host cell according to claim 25 which is a plant cell.

27. A method for producing a transgenic plant which comprises the step of regenerating a plant from a plant cell according to claim 26.
28. A plant produced according to claim 27 which is a crop species which can be maize, cotton, soya, rice, *Brassica* species, tomato, potato, sugar beet, barley, soybean, peanut, onion, rye, wheat, corn, banana, raspberry, bean, or a decorative or other plant.
29. A method of producing a peptide nematode control agent comprising causing or allowing expression of a nucleic acid according to claim 12.
30. An antibody or fragment thereof, or a polypeptide comprising the antigen-binding domain of the antibody, capable of specifically binding a peptide of claim 11.

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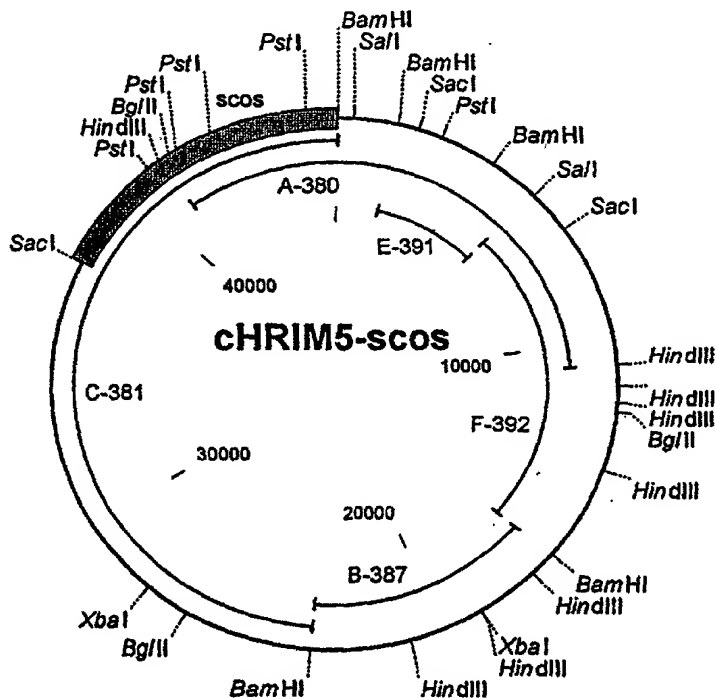

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(71) Applicant (for all designated States except US): HORTICULTURE RESEARCH INTERNATIONAL [GB/GB]; Wellesbourne, Warwick, Warwickshire CV35 9EF (GB).			
(72) Inventors; and			
(75) Inventors/Applicants (for US only): MORGAN, James, Alun, Wynne [GB/GB]; Horticulture Research International, Wellesbourne, Warwick, Warwickshire CV35 9EF (GB); JARRETT, Paul [GB/GB]; Horticulture Research International, Wellesbourne, Warwick, Warwickshire CV35 9EF (GB); ELLIS, Debbie [GB/GB]; Horticulture Research International, Wellesbourne, Warwick, Warwickshire CV35 9EF (GB); OUSLEY, Margaret, Anne [GB/GB]; Horticulture Research International, Wellesbourne, Warwick, Warwickshire CV35 9EF (GB).		Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments. With an indication in relation to deposited biological material furnished under Rule 13bis separately from the description.	
(74) Agent: RUFFLES, Graham, Keith; Marks & Clerk, 57-60 Lincoln's Inn Fields, London WC2A 3LS (GB).			

(54) Title: BIOLOGICAL CONTROL OF NEMATODES

## (57) Abstract

Nematodes can be controlled through the use of bacteria associated symbiotically with an entomopathogenic nematode. The bacteria can be employed for nematode control, or engineered to a recombinant form. Control may be achieved using material such as a peptide. The peptide can be obtained from a natural or engineered nucleic acid.



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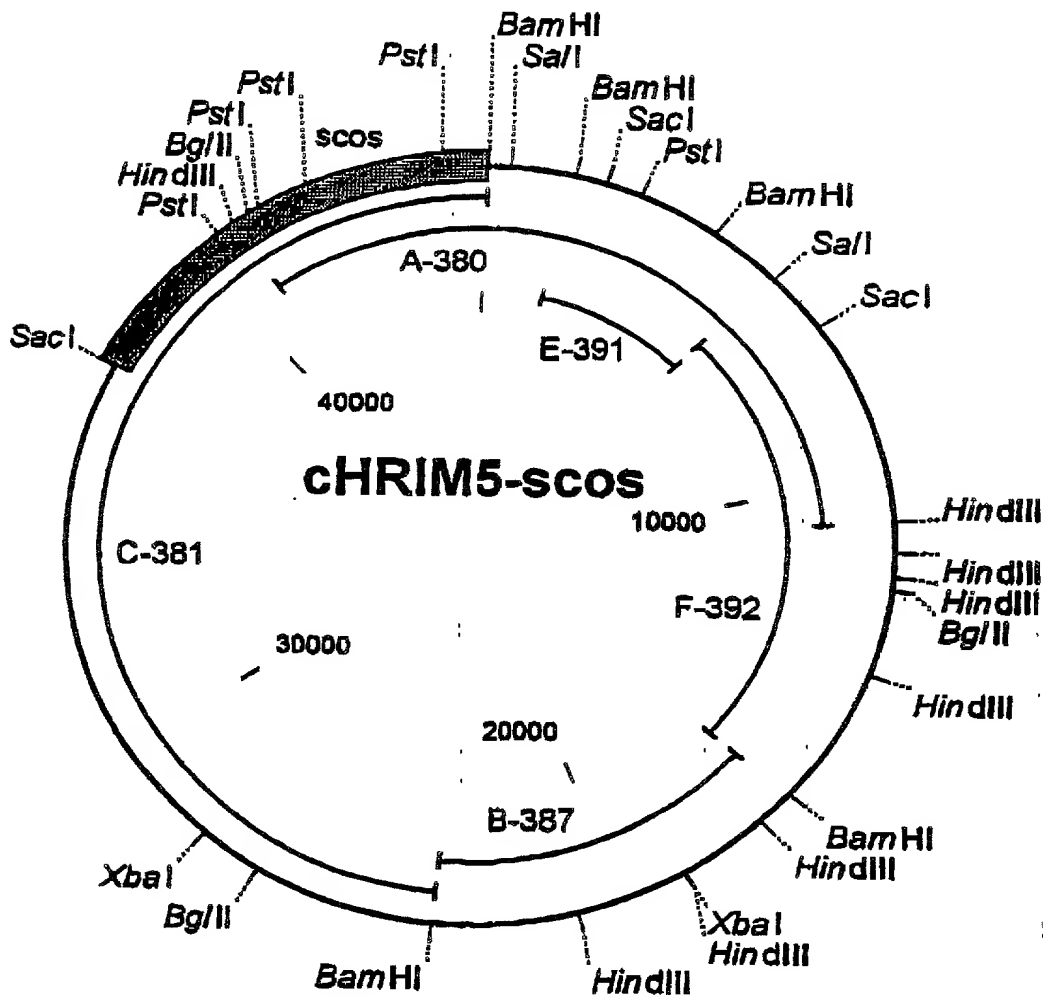


Fig. 1

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Molecule: Sequence Data  
Description: chr15ed2.seq, 37544 bps DNA

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121	ataaagaaca	ggatgttact	etecgttccct	cgacggaaga	ggottatttg	ctgcaccggg
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361	ttgccgatga	tccgcaatgc	catcagcagg	tggtgctgaa	cagtgaaccg	tacggttttc
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3901	acggcgaggga	gtcagacaga	ggctgattac	aagactgtgc	gttactcagg	caaggagcgg

Fig. 2

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chr1m5ed2.seq

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4081 aaccocggcga cactggatga taaaaacgga ctacgccccg gaaatagata tgtatttttt
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4681 gaaaataggt caattaaact acagccgggg agagttcgcg ggaatgtata tggttattct
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Fig. 2(i)

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FOOT-1236880

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chr5ed2.seq

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8401	tcgatccaac	ccaacttttt	ctctgtcaat	ttgaagcgat	cagaaagtgt	ttggggaaaca
8461	atcaacgagc	ccacctttctg	ccattgggca	togaacatt	acccgaactg	caagcctttg
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8761	gaattgtcga	tcatgcccgt	acaacacctg	atgaacagcg	ccggacagaa	tggatgaaga
8821	ccgaatcact	ggtagatttc	cttgaccat	cagatcacag	taaaacaatt	gaaggctacc
8881	ctgccccatt	gcgtgctgtc	ctcattggcc	gcaaaccata	atattgaata	aatattaatg
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12421	gccagagcgc	cctgaagtgt	gctctgctga	cagcgaaac	tttgaaagca	ggtcttgata
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Fig. 2(ii)

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MARKS AND CLERK

20 JUL 2001 19:48

NO. 4565 P. 64/101

09889874-10001



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chr15ed2.seq

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15601	gggaattacta	ccctcgggt	ggggagggtcg	ataattgccc	accagaaaccg	tatggattta
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15901	gaaaaaactta	ttctgtcgtg	cataaatttta	cctatacaaaa	acaggacgac	acgctgcaac
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Fig. 2(iii)

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Fig. 2(iv)

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Fig. 2(v)

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chr1m5ed2.seq

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Fig. 2(vi)

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Fig. 2(vii)



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Fig. 2(viii)

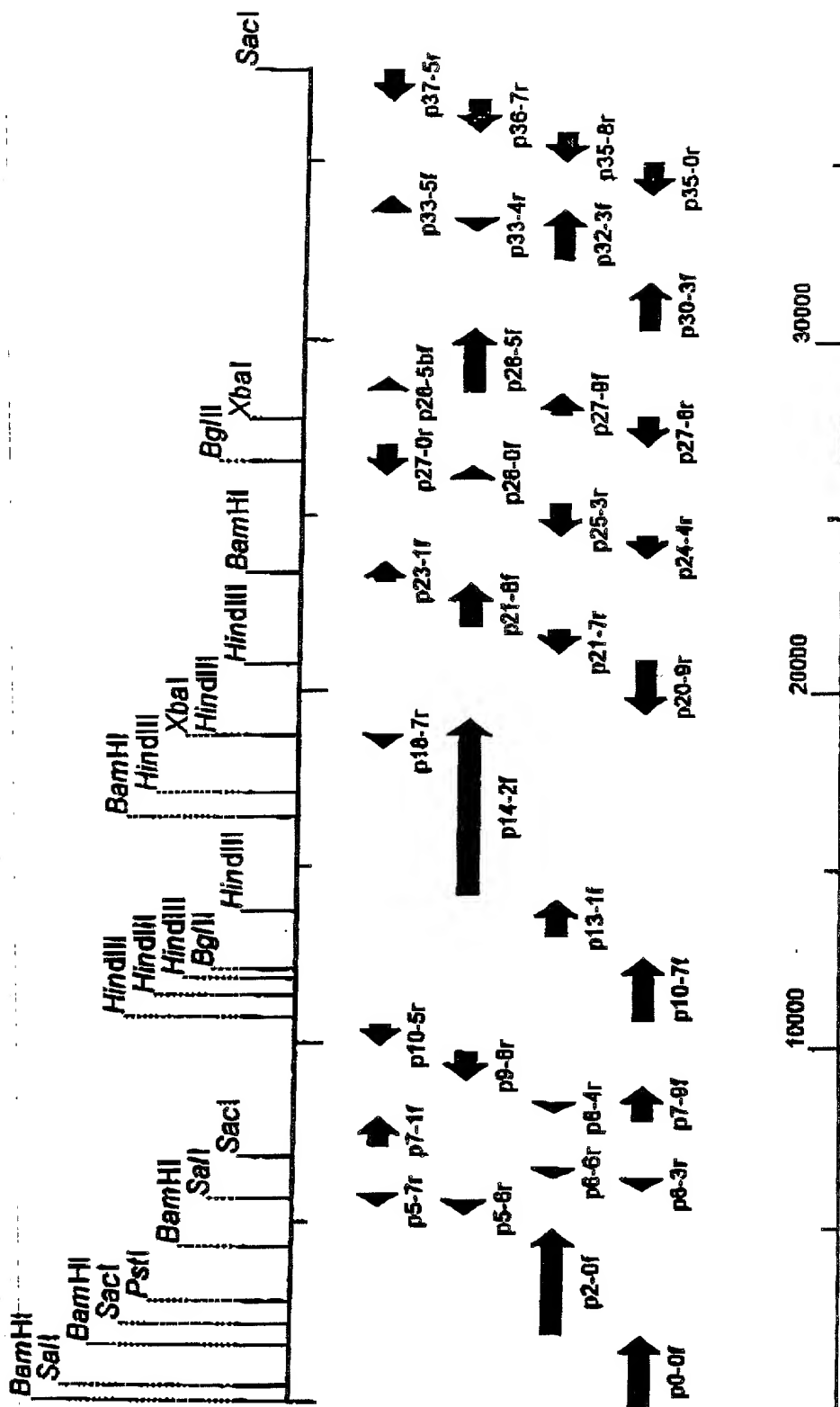
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PCT/GB00/00219

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chrom5ed2.seq (37544 bps) Fig. 3

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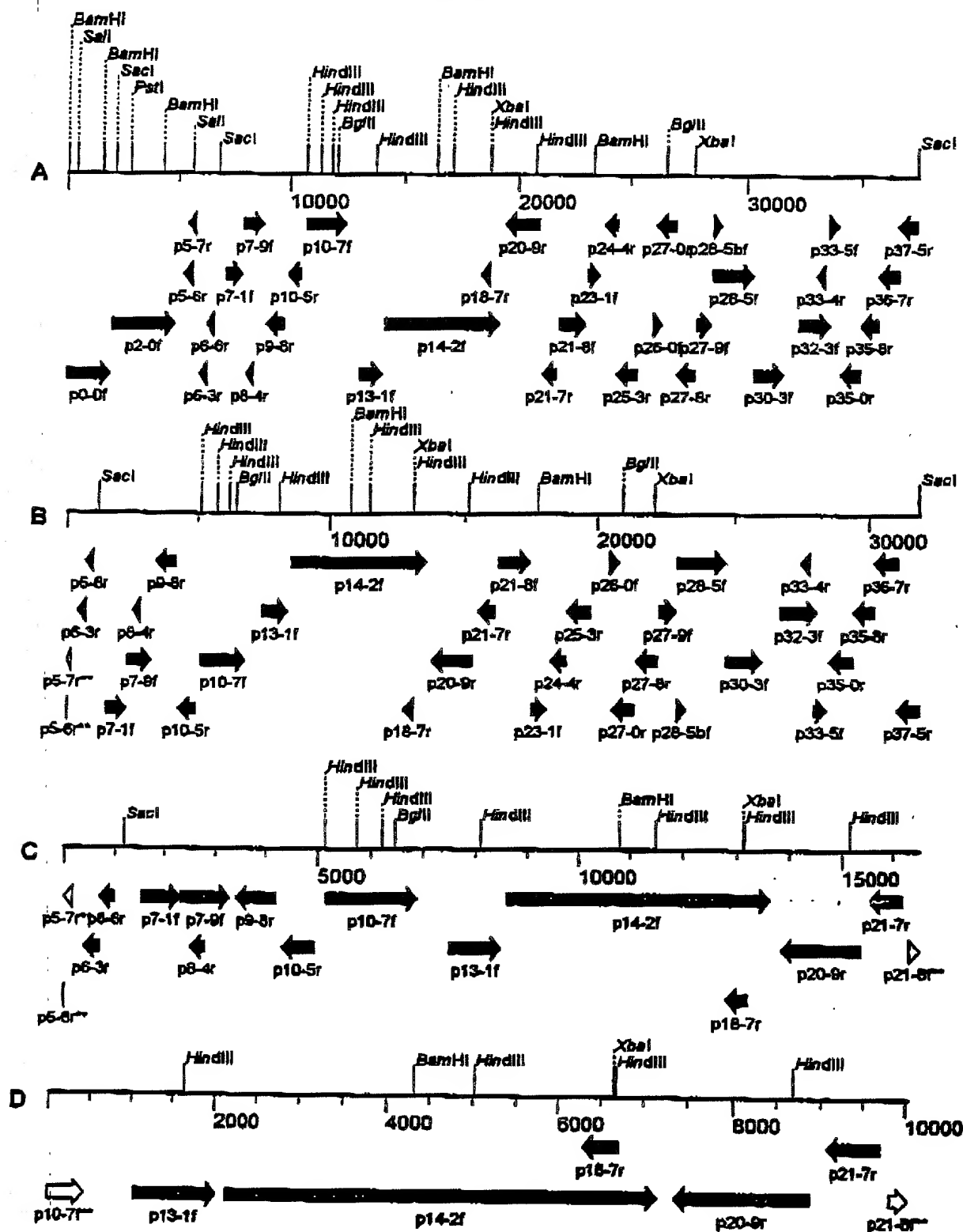


Fig. 4



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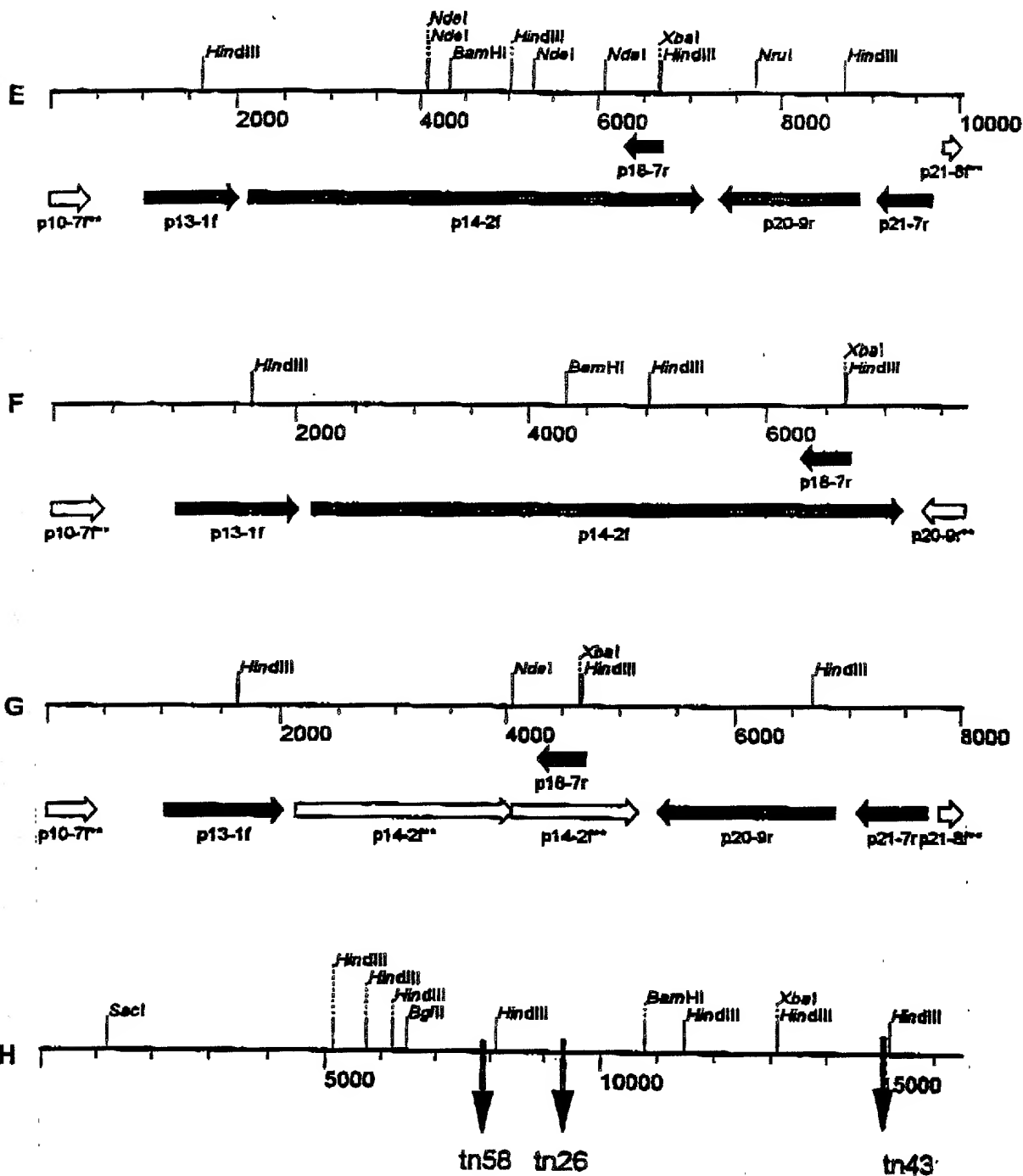


Fig. 4(cont'd)

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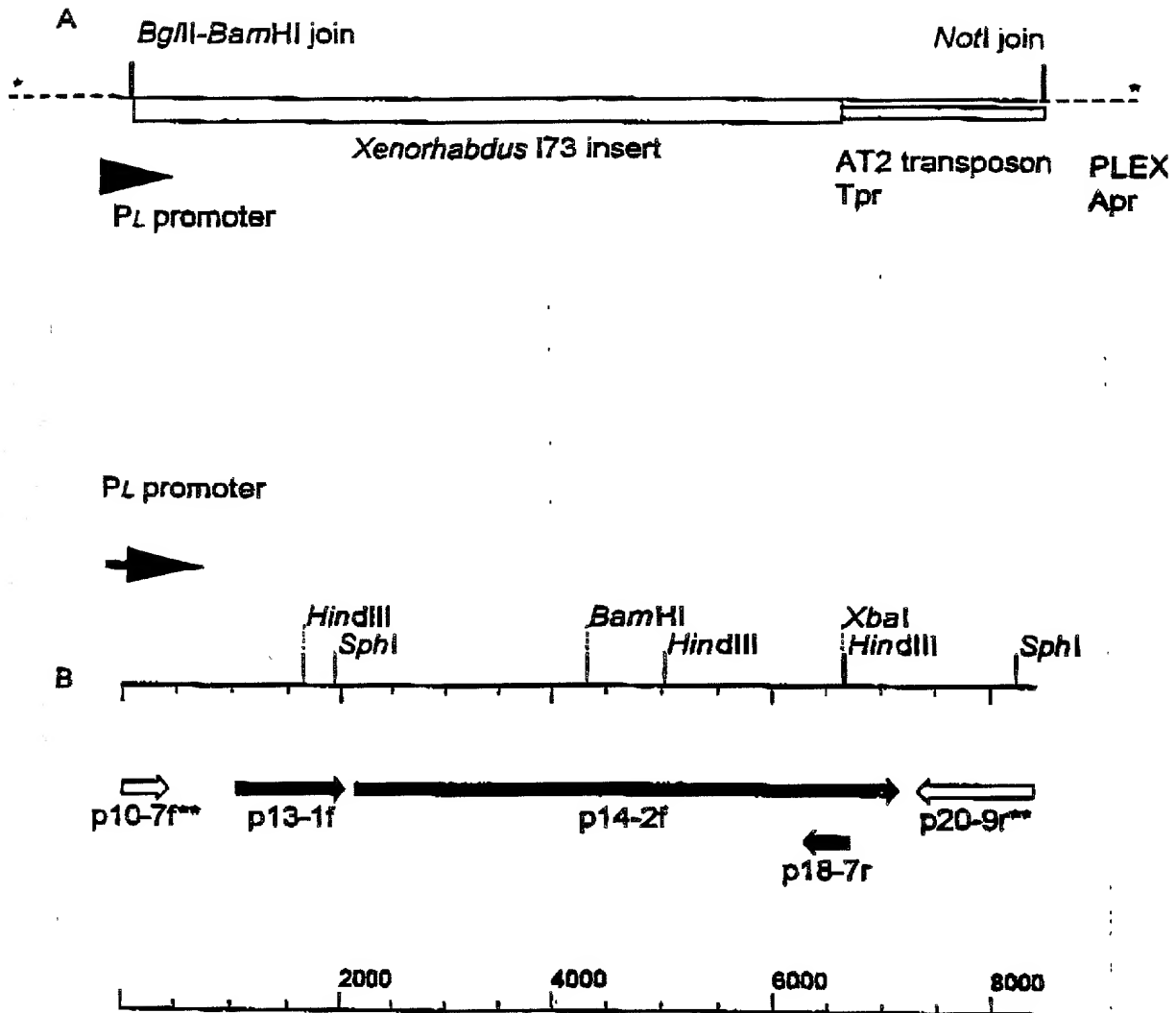


Fig. 5

## COMBINED DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled BIOLOGICAL CONTROL OF NEMATODES, the specification of which:

- ☐ is attached hereto.  
☐ was filed on \_ as Application Serial No. \_ and was amended on \_\_\_\_\_.  
☒ was described and claimed in PCT International Application No. PCT/GB00/00219 filed on 24 January 2000 and as amended under PCT Article 19 on \_\_\_\_\_.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information I know to be material to patentability in accordance with Title 37, Code of Federal Regulations, §1.56.

I hereby claim the benefit under Title 35, United States Code, §119(e)(1) of any United States provisional application(s) listed below:

U.S. Serial No.	Filing Date	Status
-----------------	-------------	--------

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose all information I know to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56(a) which became available between the filing date of the prior application and the national or PCT international filing date of this application:

U.S. Serial No.	Filing Date	Status
PCT/GB00/00219	January 24, 2000	Pending

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

Country	Application No.	Filing Date	Priority Claimed
Great Britain	9901499.5	22 January 1999	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Yes <input type="checkbox"/> No

I hereby appoint the following attorneys and/or agents to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

Timothy A. French, Reg. No. 30,175  
Anita Meiklejohn; Reg. No. 35,283  
John F. Hayden, Reg. No. 37,640

William E. Booth, Reg. No. 28,933  
David L. Feigenbaum, Reg. No. 30,378

**Combined Declaration and Power of Attorney**

Page 2 of 3 Pages

Address all telephone calls to ANITA L. MEIKLEJOHN, PH.D. at telephone number (617) 542-5070.

Address all correspondence to ANITA L. MEIKLEJOHN, PH.D. at:

FISH & RICHARDSON P.C.

225 Franklin Street

Boston, MA 02110-2804

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patents issued thereon.

Full Name of Inventor: <sup>u JAWM</sup> JAMES ALAN WYNNE MORGAN

Inventor's Signature: J.A.W. Morgan

Date: 4 October 2001

Residence Address: GREAT, BRITAIN

Citizenship: Great Britain

Post Office Address: Horticulture Research International

Wellesbourne, Warwick  
Warwickshire CV35 9EF  
GREAT BRITAIN

Full Name of Inventor: PAUL JARRETT

Inventor's Signature: P. Jarrett

Date: 4<sup>th</sup> October 2001

Residence Address: Wellesbourne, Warwick, Warwickshire

Citizenship: United Kingdom

Post Office Address: Horticulture Research International

Wellesbourne, Warwick  
Warwickshire CV35 9EF  
UNITED KINGDOM

Full Name of Inventor: DEBBIE ELLIS

Inventor's Signature: Debbie Ellis

Date: 4<sup>th</sup> October 2001

Residence Address: Wellesbourne, Warwick, Warwickshire

Citizenship: United Kingdom

Post Office Address: Horticulture Research International

Wellesbourne, Warwick  
Warwickshire CV35 9EF  
UNITED KINGDOM

## Page 3 of 3 Pages

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Charles

Wellesbourne, Warwick, Warwickshire

United Kingdom

Horticulture Research International

Wellesbourne, Warwick

Warwickshire CV35 9EF

UNITED KINGDOM

4 October 2001

GBX

# SECRET

## SEQUENCE LISTING

<110> Morgan, James Alun Wynne  
 Jarrett, Paul  
 Ellis, Debbie  
 Ousley, Margaret Anne

<120> BIOLOGICAL CONTROL OF NEMATODES

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<140> 09/889,874

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Ile	Pro	Tyr	Thr	Val	Met	Asp	Ser	Arg	Pro	Gln	Val	Arg	Leu	Leu	Thr
				85					90				95		
Gly	Leu	Pro	Gly	Asn	Ser	Pro	Thr	Val	Trp	Pro	Ser	Val	Ile	Glu	Gln
			100					105					110		
Arg	Thr	Trp	Gln	Tyr	Glu	Arg	Ile	Ala	Asp	Asp	Pro	Gln	Cys	His	Gln
			115				120					125			
Gln	Val	Val	Leu	Asn	Ser	Asp	Arg	Tyr	Gly	Phe	Pro	Arg	Glu	Thr	Val
			130			135					140				
Asp	Ile	Ala	Tyr	Pro	Arg	Arg	Pro	Lys	Pro	Ala	Val	Ser	Pro	Tyr	Pro
145					150					155				160	
Asp	Thr	Leu	Pro	Ala	Thr	Leu	Phe	Asp	Ser	Ser	Tyr	Asp	Glu	Gln	Gln
				165				170						175	
Gln	Gln	Leu	Arg	Leu	Thr	Arg	Gln	Arg	Gln	His	Tyr	His	His	Leu	Thr
			180				185						190		
Asp	Thr	Glu	His	Gln	Val	Leu	Gly	Leu	Pro	Asp	Val	Met	Arg	Ser	Asp

FOOTNOTES: 1-20000000

195	200	205
Ala Trp Gly Tyr Pro Ala	Ala Arg Val Pro Arg Glu Gly Phe Thr Leu	
210	215	220
Glu Asp Leu Leu Ala Glu Asn Ser Leu Ile Ala Pro Gly Thr Pro Leu		
225	230	235
Thr Tyr Leu Gly His Gln Arg Val Ala Tyr Thr Gly Thr Thr Gly Thr		240
245	250	255
Glu Glu Lys Pro Thr Arg Gln Ala Leu Val Ala Tyr Thr Glu Thr Ala		
260	265	270
Val Phe Asp Glu Leu Ala Leu Gln Ala Phe Asn Gly Thr Leu Ser Pro		
275	280	285
Glu Ala Leu Glu Lys Lys Leu Ile Glu Ser Gly Tyr Leu Ser Val Pro		
290	295	300
Arg Pro Phe Asn Thr Gly Ala Glu Ser Ala Val Trp Val Ala Arg Gln		
305	310	315
Gly Tyr Thr Asp Tyr Gly Gly Ser Glu Ala Phe Tyr Arg Pro Leu Ala		
325	330	335
Gln Arg Thr Thr Val Gln Ile Gly Lys Asn Thr Leu His Trp Asp Thr		
340	345	350
His Tyr Cys Ala Val Val Arg Met Gln Asp Ala Ala Gly Leu Tyr Thr		
355	360	365
Asp Ala Ala Tyr Asp Tyr Arg Phe Leu Thr Pro Val Gln Ile Thr Asp		
370	375	380
Ala Asn Asp Asn Gln Gln His Ile Thr Leu Thr Ala Leu Gly Gln Val		
385	390	395
Ser Ser Gly Arg Phe Trp Gly Thr Glu Glu Gly Thr Pro Gln Gly Tyr		
405	410	415
Thr Pro Pro Glu Asp Arg Pro Phe Thr Pro Pro Ser Ser Val Ala Glu		
420	425	430
Ala Leu Asp Leu Lys Pro Asp Leu Pro Val Ala Asn Cys Met Val Tyr		
435	440	445
Ala Pro Leu Ser Trp Met Pro Leu Ala His Thr Tyr Gln Glu Tyr Ile		
450	455	460
Ala Gly Phe Thr Trp Gln Ala Leu Leu Asp Ala Gly Val Val Thr Glu		
465	470	475
Asp Lys Arg Val Cys Ala Leu Gly Phe Arg Arg Trp Val Gln Arg Gln		
485	490	495
Gly Ile Val Leu Asn Gly Gln Ala Leu Ala Asp Ser Arg Glu Pro Val		
500	505	510
His Val Leu Thr Leu Ala Thr Asp Arg Tyr Asp Thr Asp Pro Asp Gln		
515	520	525
Gln Leu Arg Lys Ser Val Thr Tyr Ser Asp Gly Phe Gly Arg Leu Leu		
530	535	540
Gln Ser Ala Val Tyr His Ala Pro Gly Glu Ala Trp Gln Arg Ala Ala		
545	550	555
Asp Gly Ser Leu Ile Thr Asp Ala Lys Gly Ala Pro Leu Val Ala His		
565	570	575
Thr Ala Thr Arg Trp Ala Val Ser Gly Arg Thr Glu Tyr Asp Gly Lys		
580	585	590
Gly Gln Pro Val Arg Thr Tyr Pro Pro Phe Phe Leu Asn Ala Trp Gln		
595	600	605
Tyr Leu Ser Asp Asp Ser Ala Arg Gln Asp Leu Asn Ala Asp Thr His		
610	615	620
Arg Tyr Asp Pro Leu Gly Arg Glu Tyr Gln Val Arg Thr Ala Lys Gly		
625	630	635
Tyr Leu Arg Gln Asn Arg Leu Thr Pro Trp Phe Val Val Asn Glu Asp		
645	650	655

FOOT 4435550

Glu Asn Asp Thr Leu Ser  
660

<210> 2  
<211> 105  
<212> PRT  
<213> Xenorhabdus bovienii

<400> 2  
Tyr Leu Pro Gln Arg Gly Gln Cys Asp Met Leu Leu Val Val Ile Gly  
1 5 10 15  
Ile Gly Tyr Leu Asn Gly Gly Gln Glu Ala Val Ile Ile Gly Gly Ile  
20 25 30  
Arg Val Gln Thr Arg Arg Ile Leu His Thr Asp Asp Arg Thr Val Met  
35 40 45  
Gly Ile Pro Met Glu Gly Val Phe Ala Asn Leu His Arg Arg Pro Leu  
50 55 60  
Ser Gln Arg Thr Val Lys Arg Leu Arg Pro Ala Val Ile Gly Ile Ser  
65 70 75 80  
Leu Thr Gly Asp Pro Asp Arg Arg Phe Arg Thr Gly Ile Glu Trp Ala  
85 90 95  
Trp Asn Arg Gln Ile Thr Arg Leu Asp  
100 105

<210> 3  
<211> 971  
<212> PRT  
<213> Xenorhabdus bovienii

<400> 3  
Ser His Leu Pro Ala Arg Tyr Gly Gly Arg Leu Thr Thr Leu Ser Arg  
1 5 10 15  
Lys Gly Phe Met Thr Val Asn Arg Gly Asp Asn Leu His Gln Lys Thr  
20 25 30  
Pro Glu Val Thr Val Leu Asp Asn Arg Gly Leu Thr Val Arg Glu Leu  
35 40 45  
Arg Tyr His Arg His Pro Asn Thr Pro Thr Thr Thr Asp Glu Arg Ile  
50 55 60  
Thr Arg His Arg Phe Thr Leu Ser Gly Gln Leu Ala His Ser Ile Asp  
65 70 75 80  
Pro Arg Leu Phe Asp Leu Gln Gln Thr Asp Asn Thr Val Asn Pro Asn  
85 90 95  
Met Ile Tyr Asp Thr Ala Leu Thr Gly Glu Val Val Arg Thr Arg Ser  
100 105 110  
Val Asp Ala Gly Asn Asp Leu Ile Leu Asn Asp Ile Thr Gly Arg Pro  
115 120 125  
Val Leu Ala Ile Asn Ala Thr Glu Val Thr Arg Thr Trp Gln Tyr Glu  
130 135 140  
Asn Asp Thr Leu Pro Gly Arg Pro Leu Ser Ile Thr Glu Gln Pro Ala  
145 150 155 160  
Gly Glu Ala Gly Arg Ile Thr Glu Arg Phe Val Trp Ala Gly Asn Ser  
165 170 175  
Gln Ala Glu Lys Asn Ser Asn Leu Ala Gly Gln Cys Val Arg His Tyr  
180 185 190  
Asp Thr Ala Gly Leu Asn Gln Thr Asp Ser Ile Ala Leu Asn Gly Ile  
195 200 205  
Pro Leu Ser Val Thr Arg Gln Leu Leu Pro Asp Gly Thr Asp Ala Asp

T00001 4255555



210 215 220  
 Trp Gln Gly Asn Asn Glu Pro Ala Trp Asn Asp Arg Leu Ala Pro Glu  
 225 230 235 240  
 Asn Phe Thr Thr Leu Ser Thr Ala Asp Ala Thr Gly Ala Val Leu Thr  
 245 250 255  
 Thr Thr Asp Ala Ala Gly Asn Leu Gln Arg Val Ala Tyr Asp Val Ala  
 260 265 270  
 Gly Leu Leu Thr Gly Ser Trp Leu Arg Leu Ala Gly Gly Thr Glu Gln  
 275 280 285  
 Val Ile Val Lys Ser Leu Thr Tyr Ser Ala Ala Gly Gln Lys Leu Arg  
 290 295 300  
 Glu Glu His Gly Asn Gly Val Val Thr Thr Tyr Thr Tyr Glu Pro Glu  
 305 310 315 320  
 Thr Gln Arg Leu Val Gly Ile Lys Thr Lys Arg Pro Gln Gly His Ala  
 325 330 335  
 Gln Gly Thr Lys Val Leu Gln Asp Leu Arg Tyr Glu Tyr Asp Pro Val  
 340 345 350  
 Gly Asn Val Val Lys Val Thr Asn Asp Ala Glu Val Thr Arg Phe Trp  
 355 360 365  
 Arg Asn Gln Lys Val Val Pro Glu Asn Thr Tyr Val Tyr Asp Ser Leu  
 370 375 380  
 Tyr Gln Leu Val Ser Ala Thr Gly Arg Glu Met Ala Asn Ile Val Gln  
 385 390 395 400  
 Gln Ser Thr Leu Leu Pro Thr Pro Ser Leu Ile Asp Ser Ser Thr Tyr  
 405 410 415  
 Ser Asn Tyr Ser Arg Thr Tyr Asn Tyr Asp Arg Gly Asp Asn Leu Thr  
 420 425 430  
 Gln Ile Arg His Ser Ala Pro Ala Thr Gly Asn Ser Tyr Thr Thr Asp  
 435 440 445  
 Ile Thr Val Ser Asp His Ser Asn Arg Ala Val Leu Asp Thr Leu Thr  
 450 455 460  
 Asp Asp Pro Ala Lys Val Asp Ala Leu Phe Thr Ala Gly Gly His Gln  
 465 470 475 480  
 Ile Pro Leu Gln Pro Gly Gln Asn Leu Val Trp Thr Pro Arg Gly Glu  
 485 490 495  
 Leu Leu Lys Val Ala Pro Val Val Arg Asp Gly Gln Ile Ser Asp Gln  
 500 505 510  
 Glu Ser Tyr Arg Tyr Asp Ala Ala Ser Gln Arg Ile Ile Lys Thr His  
 515 520 525  
 Val Gln Gln Thr Ala Asn Ser Ser Gln Ala Gln Ser Thr Leu Tyr Leu  
 530 535 540  
 Pro Gly Leu Glu Arg His Thr Thr Ile Asn Gly Thr Thr Val Lys Glu  
 545 550 555 560  
 Val Leu His Val Ile Thr Ile Gly Glu Ala Gly Arg Ala Gln Val Arg  
 565 570 575  
 Val Leu His Trp Glu Asn Gly Lys Pro Gly Ala Ile Ser Asn Asn Gln  
 580 585 590  
 Met Arg Tyr Ser Tyr Asp Asn Leu Ile Gly Ser Ser Gly Leu Glu Val  
 595 600 605  
 Asp Gly Asp Gly Gln Ile Ile Ser Met Glu Glu Tyr Tyr Pro Tyr Gly  
 610 615 620  
 Gly Thr Ala Val Trp Thr Ala Arg Ser Gln Thr Glu Ala Asp Tyr Lys  
 625 630 635 640  
 Thr Val Arg Tyr Ser Gly Lys Glu Arg Asp Ala Thr Gly Leu Tyr Tyr  
 645 650 655  
 Tyr Gly Tyr Arg Tyr Tyr Gln Pro Trp Ala Gly Ser Trp Leu Ser Ala  
 660 665 670

123456789101112131415161718192021222324252627282930313233343536373839404142434445464748495051525354555657585960616263646566676869707172737475767778798081828384858687888990919293949596979899100

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<210> 4
<211> 108
<212> PRT
<213> Xenorhabdus bovienii
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<400> 4															
Pro	Ala	Ala	Glu	Tyr	Val	Arg	Asp	Phe	Thr	Ile	Thr	Cys	Ser	Val	Pro
1				5					10					15	
Pro	Ala	Ser	Arg	Ser	Gln	Leu	Pro	Val	Ser	Arg	Pro	Ala	Thr	Ser	Tyr
			20					25					30		
Ala	Thr	Arg	Cys	Arg	Leu	Pro	Ala	Ala	Ser	Val	Val	Val	Ser	Thr	Ala
		35				40						45			
Pro	Val	Ala	Ser	Ala	Val	Leu	Arg	Val	Val	Lys	Phe	Ser	Gly	Ala	Ser
	50					55					60				
Arg	Ser	Phe	Gln	Ala	Gly	Ser	Leu	Phe	Pro	Cys	Gln	Ser	Ala	Ser	Val
65					70					75					80
Pro	Ser	Gly	Ser	Ser	Trp	Arg	Val	Thr	Asp	Ser	Gly	Met	Pro	Leu	Ser
				85					90					95	

Ala Ile Leu Ser Val Trp Phe Ser Pro Ala Val Ser  
100 105

<210> 5

<211> 256

<212> PRT

<213> Xenorhabdus bovienii

<400> 5

Gln Arg Ala Leu Leu Asn Asp Ile Gly His Phe Ala Pro Gly Gly Thr  
1 5 10 15  
Asp Gln Leu Ile Gln Ala Val Ile Asp Ile Gly Val Leu Arg His His  
20 25 30  
Phe Leu Val Ala Pro Glu Ala Gly Asn Leu Arg Ile Val Arg His Phe  
35 40 45  
His His Val Pro His Arg Val Val Leu Ile Ala Gln Val Leu Gln His  
50 55 60  
Leu Arg Pro Leu Cys Met Ser Leu Trp Ala Phe Gly Phe Tyr Ala Asn  
65 70 75 80  
Lys Ala Leu Gly Leu Arg Leu Val Gly Val Gly Gly His His Ala Val  
85 90 95  
Ala Val Leu Phe Ala Gln Phe Leu Thr Arg Gly Gly Ile Arg Gln Gly  
100 105 110  
Phe His Asp Asn Leu Leu Cys Pro Ala Arg Lys Pro Gln Pro Thr Ala  
115 120 125  
Ser Gln Gln Ala Cys Tyr Val Ile Arg His Thr Leu Gln Val Thr Gly  
130 135 140  
Arg Ile Gly Gly Gly Gln Tyr Arg Ala Gly Gly Ile Arg Arg Ala Gln  
145 150 155 160  
Gly Gly Glu Val Phe Arg Cys Gln Pro Val Val Pro Gly Gly Phe Ile  
165 170 175  
Val Ser Leu Pro Val Cys Val Arg Thr Ile Arg Gln Gln Leu Ala Arg  
180 185 190  
Asp Gly Gln Arg Tyr Ala Val Lys Arg Asn Thr Val Arg Leu Val Gln  
195 200 205  
Ser Gly Gly Val Ile Val Thr His Ala Leu Ser Gly Gln Val Ala Val  
210 215 220  
Leu Leu Arg Leu Thr Val Pro Cys Pro Asp Lys Thr Leu Cys Asp Thr  
225 230 235 240  
Ala Cys Phe Ala Ser Arg Leu Phe Cys Asp Thr Glu Arg Ala Ser Gly  
245 250 255

<210> 6

<211> 316

<212> PRT

<213> Xenorhabdus bovienii

<400> 6

Ser Asp Arg Arg Gln Thr Gly Tyr Ala Tyr Ser Ala Asp His Tyr Arg  
1 5 10 15  
Ile Ser Gly Arg Ser Thr Val Cys Thr Val Arg Ala Gly Leu Met Asn  
20 25 30  
Tyr Gln Cys Trp Leu Gln His Ala Ala Thr Gln Leu Ser Glu Ser Asp  
35 40 45  
Ser Pro Lys Arg Asp Ala Glu Ile Leu Leu Gly Tyr Val Thr Gly Arg  
50 55 60  
Ser Arg Thr Tyr Leu Ile Ala Phe Asp Glu Thr Leu Ile Ser Ser Glu

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65                      70                      75                      80  
 Glu Leu His Gln Leu Asp Ser Leu Leu Val Arg Arg Ile Gln Gly Glu  
                                  85                      90                      95  
 Pro Val Ala Tyr Ile Ile Gly Glu Arg Glu Phe Trp Ser Leu Pro Phe  
                                  100                      105                      110  
 Ala Val Ser Pro Ala Thr Leu Ile Pro Arg Pro Asp Thr Glu Cys Leu  
                                  115                      120                      125  
 Val Glu Lys Ala Leu Glu Leu Leu Pro Asp Ser Pro Ala Arg Ile Leu  
                                  130                      135                      140  
 Asp Leu Gly Thr Gly Thr Gly Ala Ile Ala Leu Ala Leu Ala Ser Glu  
 145                                   150                      155                      160  
 Arg Asn Asp Cys Tyr Val Thr Gly Val Asp Ile Asn Ser Asp Ala Val  
                                  165                      170                      175  
 Met Leu Ala Gln His Asn Ala Glu Lys Asn Ala Gly Lys Leu Ala Ile  
                                  180                      185                      190  
 His Asn Val Asn Phe Leu Gln Ser Glu Trp Phe Ala Ala Val Gly Asn  
                                  195                      200                      205  
 Gln Gln Phe Asp Met Ile Val Ser Asn Pro Pro Tyr Ile Asp Glu Arg  
                                  210                      215                      220  
 Asp Pro His Leu Gln Glu Gly Asp Ile Arg Phe Glu Pro Ala Thr Ala  
 225                                   230                      235                      240  
 Leu Ile Ala Ala Gln Asn Gly Met Ala Asp Leu Gln Ala Ile Val Gly  
                                  245                      250                      255  
 Gln Ala Arg His Phe Leu Ser Pro Asn Gly Trp Leu Leu Leu Glu His  
                                  260                      265                      270  
 Gly Trp Lys Gln Gly Thr Val Val Arg Asn Leu Phe Leu Glu Lys Gly  
                                  275                      280                      285  
 Tyr Gln Gln Ile Ala Thr Phe Gln Asp Tyr Gly Gly Asn Glu Arg Ile  
                                  290                      295                      300  
 Thr Ile Gly Arg Trp Asn Lys Asn Glu Thr His Ser  
 305                                   310                      315

<210> 7

<211> 102

<212> PRT

<213> Xenorhabdus bovienii

<400> 7

Ala Arg Arg Ala Val Arg Arg Cys Gly Tyr Cys Thr Gly Arg Thr Glu  
 1                      5                      10                      15  
 Ser Arg Val Pro Ser Val Thr Thr Arg Cys Ala Thr Ala Met Ile Thr  
                                  20                      25                      30  
 Leu Ser Ala Ala Ala Val Trp Arg Trp Thr Val Thr Asp Lys Leu Ser  
                                  35                      40                      45  
 Val Trp Lys Asn Thr Thr Arg Thr Gly Ala Leu Arg Cys Gly Arg Arg  
                                  50                      55                      60  
 Gly Val Arg Gln Arg Leu Ile Thr Arg Leu Cys Val Thr Gln Ala Arg  
 65                                   70                      75                      80  
 Ser Gly Met Gln Arg Gly Cys Ile Ile Thr Ala Thr Gly Ile Thr Ser  
                                  85                      90                      95  
 Arg Gly Arg Gly Ala Gly  
                                  100

<210> 8

<211> 130

<212> PRT

<213> Xenorhabdus bovienii

F00001-129999

[illegible]

<211> 119

<212> PRT

<213> Xenorhabdus bovienii

[illegible]

<210> 10

<211> 138

<212> PRT

<213> Xenorhabdus bovienii

Val	His	Ser	Pro	Ser	Gly	Ala	Val	Ala	Pro	Gly	Lys	Phe	Phe	Ile	Glu
1				5					10					15	
Asn	Phe	Ala	Asp	Thr	Phe	Pro	Ala	Pro	Leu	Pro	Leu	His	Pro	Phe	Ile
			20					25					30		
Asp	Ala	Cys	Ile	Gln	Gln	Gly	Phe	Gln	Leu	Leu	Pro	Cys	Leu	Ile	Ala
		35					40					45			
Ile	Ala	His	Ser	Gly	Lys	Gln	Ala	Phe	Glu	Cys	Val	Leu	Leu	Asp	Arg

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<210> 11
<211> 110
<212> PRT
<213> Xenorhabdus bovienii
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<210> 12
<211> 103
<212> PRT
<213> Xenorhabdus bovienii
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<210> 13
<211> 265
<212> PRT
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&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 13

Asn Ala His Phe Leu Ile Val Ser Lys Thr Asn Val Val Met Ser Asn  
 1 5 10 15  
 Gln Asp Pro His Asn Lys Arg Asp Ser Leu Phe Ser Ala Pro Ile Ala  
 20 25 30  
 Asn Leu Gly Asp Trp Ser Phe Asp Glu Arg Val Ala Glu Val Phe Pro  
 35 40 45  
 Asp Met Val Lys Arg Ser Ile Pro Gly Tyr Ser Asn Ile Ile Ser Met  
 50 55 60  
 Ile Gly Met Leu Ala Ser Arg Phe Val Thr Pro Gly Ser Gln Ile Tyr  
 65 70 75 80  
 Asp Leu Gly Cys Ser Leu Gly Ala Ala Thr Leu Ser Ile Arg Arg Ser  
 85 90 95  
 Ile Asn Ala Asp Asn Cys Arg Ile Ile Ala Ile Asp Asn Ser Pro Ala  
 100 105 110  
 Met Ile Glu Arg Cys Arg Arg His Ile Asp Ser Phe Lys Ala Ser Thr  
 115 120 125  
 Pro Val Glu Val Ile Glu Gln Asn Ile Leu Asp Thr Asp Ile Gln Asn  
 130 135 140  
 Ala Ser Met Val Val Leu Asn Phe Thr Leu Gln Phe Leu His Pro Asp  
 145 150 155 160  
 Asp Arg Gln Lys Ile Leu Lys Lys Ile Tyr Ala Gly Leu Lys Pro Gly  
 165 170 175  
 Gly Val Leu Val Leu Ser Glu Lys Phe Asn Phe Glu Asp Gln Lys Ile  
 180 185 190  
 Gly Glu Leu Leu Phe Asn Met His Asp Phe Lys Arg Ala Asn Gly  
 195 200 205  
 Tyr Ser Glu Leu Glu Val Ser Gln Lys Arg Ser Met Leu Glu Asn Val  
 210 215 220  
 Met Arg Thr Asp Ser Val Asp Thr His Lys Ser Arg Leu Lys Glu Val  
 225 230 235 240  
 Gly Phe Gln His Val Glu Val Trp Phe Gln Cys Phe Asn Phe Gly Ser  
 245 250 255  
 Leu Leu Ala Ile Lys Gly Thr Glu Gln  
 260 265

&lt;210&gt; 14

&lt;211&gt; 324

&lt;212&gt; PRT

&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 14

Thr Met Ile Asp Phe Gly Asn Phe Tyr Gln Leu Ile Ala Lys His Pro  
 1 5 10 15  
 Leu Asn His Trp Leu Asp Ser Leu Pro Ala Gln Leu Ser His Trp Gln  
 20 25 30  
 Lys Thr Ser Gln His Gly Gln Phe Ser Ser Trp Val Lys Ile Leu Glu  
 35 40 45  
 Asn Leu Pro Glu Ile Lys Pro Ser His Leu Asp Leu Lys Asn Gly Val  
 50 55 60  
 Ile Ala Ile His Glu Pro Asp Leu Ser Lys Gly Glu Lys Ala Arg Leu  
 65 70 75 80  
 His Asn Ile Leu Lys Ile Leu Met Pro Trp Arg Lys Gly Pro Phe Ser  
 85 90 95  
 Leu Tyr Asp Val Glu Ile Asp Thr Glu Trp Arg Ser Asp Trp Lys Trp

100 105 110  
 Glu Arg Val Leu Pro His Ile Ser Pro Leu Glu Gly Lys Thr Val Leu  
 115 120 125  
 Asp Val Gly Cys Gly Ser Gly Tyr His Met Trp Arg Met Val Gly Glu  
 130 135 140  
 Gly Ala Gln Leu Val Val Gly Ile Asp Pro Thr Gln Leu Phe Leu Cys  
 145 150 155 160  
 Gln Phe Glu Ala Ile Arg Lys Leu Leu Gly Asn Asn Gln Arg Ala His  
 165 170 175  
 Leu Leu Pro Leu Gly Ile Glu Gln Leu Pro Glu Leu Gln Ala Phe Asp  
 180 185 190  
 Thr Val Phe Ser Met Gly Val Leu Tyr His Arg Arg Ser Pro Leu Asp  
 195 200 205  
 His Leu Trp Gln Leu Lys Asn Gln Leu Val Ser Asp Gly Glu Leu Val  
 210 215 220  
 Leu Glu Ser Leu Val Ile Glu Gly Asp Glu Asn Gln Cys Leu Ile Pro  
 225 230 235 240  
 Gly Glu Arg Tyr Ala Gln Met Arg Asn Val Tyr Phe Ile Pro Ser Ala  
 245 250 255  
 Lys Met Leu Lys Val Trp Leu Glu Lys Cys Gly Phe Val Asp Val Arg  
 260 265 270  
 Ile Val Asp His Ala Ala Thr Thr Pro Asp Glu Gln Arg Arg Thr Glu  
 275 280 285  
 Trp Met Lys Thr Glu Ser Leu Val Asp Phe Leu Asp Pro Ser Asp His  
 290 295 300  
 Ser Lys Thr Ile Glu Gly Tyr Pro Ala Pro Leu Arg Ala Val Leu Ile  
 305 310 315 320  
 Ala Arg Lys Pro

<210> 15  
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 <212> PRT  
 <213> Xenorhabdus bovienii

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 Pro Ile Glu Arg Leu Arg Gln Pro Cys Ala Thr Cys Asp Asn His Cys  
 20 25 30  
 His Ser Arg His Gln Val Arg Phe Leu Leu Lys Glu Lys Tyr Gly  
 35 40 45  
 Ala Ala Leu Ala Pro Ile Ser Ser Gln Ser Ala Ile Arg Tyr Gln Phe  
 50 55 60  
 Gln Arg His Thr Met Lys Lys Gly Leu Phe Ala Met Ala Ser Ile Phe  
 65 70 75 80  
 Ser Gly Tyr Cys Gly Gly Glu Leu Phe His Leu Leu Thr Asp Pro Ala  
 85 90 95  
 His Glu Ser Gln  
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<210> 16  
 <211> 267  
 <212> PRT  
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<400> 16

T000001-42353560



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<210> 17
<211> 189
<212> PRT
<213> Xenorhabdus bovienii
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Arg	Asn	Phe	Tyr 20	Gly	Leu	Phe	Asn	Gly 25	Glu	Glu	Met	Ser	His 30	Phe	Ser
Lys	Ile	Ser 35	Glu	Leu	Gln	Asp	Leu 40	Val	Ala	Asp	Leu	Ala 45	Gly	Phe	Glu
Gln 50	Lys	Leu	Lys	Gln	Phe	Glu 55	Gly	His	Leu	Gly	Leu 60	His	Phe	Glu	Gln
Tyr 65	Ser	Ala	Asp	His 70	Ile	Ser	Leu	Arg	Cys	Asn 75	Glu	Ser	Lys	Ile 80	Ala
Asp	Arg	Trp	Arg 85	Lys	Gly	Phe	Leu	Gln 90	Cys	Gly	Gln	Leu	Ile 95	Ser	Glu
Ser	Ile	Ile	Asn 100	Gly	Arg	Pro	Ile	Cys 105	Leu	Phe	Asp	Leu	Asn 110	Gln	Pro
Ile	Val	Leu 115	Leu	Asp	Trp	Lys	Ile 120	Asp	Cys	Val	Glu	Leu 125	Pro	Tyr	Pro

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<210> 18
<211> 579
<212> PRT
<213> Xenorhabdus bovienii
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			20					25					30		
Gln	Ser	Ala	Lys	Ala	Gln	Phe	Gly	Asp	Tyr	Gln	Ala	Asn	Gly	Val	Met
		35					40					45			
Ala	Ala	Ala	Lys	Lys	Val	Gly	Ile	Pro	Pro	Arg	Gln	Leu	Ala	Glu	Lys
	50					55					60				
Val	Val	Ser	Gln	Leu	Asp	Leu	Gln	Gly	Ile	Ala	Ser	Lys	Val	Glu	Ile
65					70					75					80
Ala	Gly	Pro	Gly	Phe	Ile	Asn	Ile	Phe	Leu	Asp	Lys	Ala	Trp	Val	Ala
				85					90					95	
Ala	Asn	Ile	Glu	Thr	Thr	Leu	Lys	Asp	Glu	Lys	Leu	Gly	Ile	Thr	Pro
			100					105					110		
Val	Glu	Pro	Gln	Thr	Ile	Val	Ile	Asp	Tyr	Ser	Ala	Pro	Asn	Val	Ala
		115					120					125			
Lys	Gln	Met	His	Val	Gly	His	Leu	Arg	Ser	Thr	Ile	Ile	Gly	Asp	Ala
	130					135					140				
Ala	Ala	Arg	Thr	Leu	Glu	Phe	Leu	Gly	His	Lys	Val	Ile	Arg	Ala	Asn
145					150					155					160
His	Val	Gly	Asp	Trp	Gly	Thr	Gln	Phe	Gly	Met	Leu	Ile	Ala	Tyr	Leu
				165					170					175	
Glu	Lys	Ile	Gln	Asn	Glu	Asn	Ala	Asn	Asp	Met	Ala	Leu	Ala	Asp	Leu
			180					185					190		
Glu	Ala	Phe	Tyr	Arg	Glu	Ala	Lys	Lys	His	Tyr	Asp	Glu	Asp	Glu	Glu
		195					200				205				
Phe	Ala	Ile	Arg	Ala	Arg	Asn	Tyr	Val	Val	Lys	Leu	Gln	Gly	Gly	Asp
	210					215					220				
Glu	Tyr	Cys	Arg	Lys	Met	Trp	Arg	Lys	Leu	Val	Asp	Ile	Thr	Met	Ser
225					230					235				240	
Gln	Asn	Gln	Glu	Thr	Tyr	Asn	Arg	Leu	Asn	Val	Thr	Leu	Thr	Glu	Lys
				245					250					255	
Asp	Val	Met	Gly	Glu	Ser	Leu	Tyr	Asn	Asp	Met	Leu	Pro	Gly	Ile	Val
			260					265					270		
Ala	Asp	Leu	Lys	Gln	Arg	Gly	Ile	Ala	Val	Lys	Ser	Asp	Gly	Ala	Thr
		275					280					285			
Val	Val	Tyr	Leu	Asp	Glu	Phe	Lys	Asn	Lys	Glu	Gly	Glu	Pro	Met	Gly

Leu Tyr Tyr Ile Asp Ser Arg Gln His Gln His Leu Met Gln Ala Trp  
 340 345 350  
 Ala Ile Val Arg Lys Thr Gly Tyr Ile Pro Glu Ser Met Ser Leu Glu  
 355 360 365  
 His His Met Phe Gly Met Met Leu Gly Lys Asp Gly Lys Pro Phe Lys  
 370 375 380  
 Thr Arg Ala Gly Gly Thr Val Arg Leu Ser Asp Leu Leu Asp Glu Ala  
 385 390 395 400  
 Ile Glu Arg Ala Asp Thr Leu Ile Arg Glu Lys Asn Pro Asp Met Pro  
 405 410 415  
 Glu Asp Glu Leu Lys Lys Val Val Glu Ala Val Gly Ile Gly Ala Val  
 420 425 430  
 Lys Tyr Ala Asp Leu Ser Lys Ser Arg Thr Thr Asp Tyr Val Phe Asp  
 435 440 445  
 Trp Asp Asn Met Leu Ala Phe Glu Gly Asn Thr Ala Pro Tyr Met Gln  
 450 455 460  
 Tyr Ala Tyr Thr Arg Val Ser Ser Ile Phe Lys Arg Ala Asp Ile Asp  
 465 470 475 480  
 Glu Asn Ser Leu Thr Leu Pro Val Met Leu Asn Glu Glu Arg Glu Gln  
 485 490 495  
 Ala Leu Ala Thr Arg Leu Leu Gln Phe Glu Glu Thr Ile Thr Thr Val  
 500 505 510  
 Ala Arg Glu Gly Thr Pro His Val Met Cys Ala Tyr Leu Tyr Asp Leu  
 515 520 525  
 Ala Gly Leu Phe Ser Gly Phe Tyr Glu His Cys Pro Ile Leu Asn Ala  
 530 535 540  
 Asp Ser Glu Glu Leu Arg Gln Ser Arg Leu Lys Leu Ala Leu Leu Thr  
 545 550 555 560  
 Ala Lys Thr Leu Lys Gln Gly Leu Asp Thr Leu Gly Ile Gln Thr Val  
 565 570 575  
 Glu Arg Met

<210> 19  
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 <212> PRT  
 <213> Xenorhabdus bovienii

<400> 19  
 Ala Gln Val Ser Asn Met His Leu Leu Gly Asp Ile Arg Cys Gly Ile  
 1 5 10 15  
 Ile Asp Asn Asp Gly Leu Arg Phe His Trp Gly Asp Thr Glu Leu Phe  
 20 25 30  
 Ile Phe Gln Gly Ser Phe Tyr Ile Cys Cys Asn Pro Arg Phe Ile Lys  
 35 40 45  
 Lys Asn Ile Asp Lys Thr Trp Ala Cys Asn Phe Asn Phe Ala Gly Asn  
 50 55 60  
 Ser Leu Gln Ile Gln Leu Ala Asp Asp Phe Phe Cys Gln Leu Ser Arg  
 65 70 75 80  
 Arg Tyr Ser His Leu Phe Ser Gly Ser His His Thr Ile Arg Leu Ile  
 85 90 95  
 Val Thr Lys Leu Cys Phe Gly Arg Leu Thr Asp Val Ser Phe Thr Val  
 100 105 110  
 Gly Trp Ser Ala Ser Phe Asn Gln Arg Ile Ala Asp Phe Phe  
 115 120 125

<210> 20

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<400> 22
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 1          5          10          15
Phe Asn Val Ile Leu Glu Thr Gly Leu Asp Asn Cys Asn Ile Tyr Ala
          20          25          30
Asn Gly Leu Asn Met Ile Gly Val Ile Ile Asn Ile Thr Pro Thr Asp
          35          40          45
Asp Glu Gly Asn Phe Val Asp Ile Asp Asp Val Thr Leu Asn Asp Asn
 50          55          60

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Ile Lys Ile Val Asp Tyr Ile Asp Gly Ser Asp Ile Asp Gly Ser Asp  
 65 70 75 80  
 Gly Trp Phe Tyr Thr Gly Asn Pro Asn Glu Tyr Asn Thr Ile Pro Asn  
 85 90 95  
 Ser Gln Ser Tyr Ser Leu Leu Lys Ser Glu Asn Ser Gln Ile Thr Gln  
 100 105 110  
 Ile Lys Arg Tyr Val Ser Cys Ser Asn Thr Ser Arg Leu Arg Thr Lys  
 115 120 125  
 Ser Phe Ser Ala Lys Val Thr Thr Thr Ser Gly Lys Val Ile Ser Ile  
 130 135 140  
 Thr Gln Asn Ser Ile Asn Ser Ser Arg Val Val Ile Asn Ala Ile Asp  
 145 150 155 160  
 Ala Thr Asn Phe Thr Asp Asp Glu Leu Arg Thr Thr Lys Glu Thr Arg  
 165 170 175  
 Phe Glu Asn Gln Ser Tyr Thr Ser His Lys Ser Ser Thr Asn Ser Leu  
 180 185 190  
 Tyr Val His Thr Trp Thr Ile Pro Arg Ser Leu Lys Leu Gln Asn Trp  
 195 200 205  
 Arg Trp Glu Asp Tyr Asn Asn Gly Trp Thr Trp Ala Gln Ser Cys Tyr  
 210 215 220  
 Tyr Lys Thr Gly Ala Asp Gly Gly Ser Glu Ser Thr Arg Trp Leu Ala  
 225 230 235 240  
 Ala Gly Ser Ile Phe Pro Pro Gly Asn Tyr Asp Gly Leu Trp Leu Asp  
 245 250 255  
 Asn Asp Ile Ala Leu Ser Gly Met Ala His Lys Ser Tyr Asn Val Asp  
 260 265 270  
 Thr Gly Ile Asn Gln Leu Ser Phe Thr Arg Ile Ile Gly Lys Gly Phe  
 275 280 285  
 Ser Trp Val Tyr Asn Ile Ser Gly Leu Asp Arg Gly His Ala Val Ile  
 290 295 300  
 Ile Ile Asp Gln Tyr Gly Asn Lys Tyr Arg Ile Leu Phe His Ala Gly  
 305 310 315 320  
 Tyr Glu Asn Ser Asp Pro Tyr Leu Ser Ser Ser Ile Val Tyr  
 325 330

<210> 23  
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 <212> PRT  
 <213> Xenorhabdus bovienii

<400> 23  
 Val Tyr Ile Lys Phe Leu Lys Leu Phe Arg Arg Ile Thr Met Ser Asp  
 1 5 10 15  
 Asn Asn Glu Phe Phe Thr Gln Ala Asn Asn Phe Thr Ser Ala Val Ser  
 20 25 30  
 Gly Gly Val Asp Pro Arg Thr Gly Leu Tyr Asn Ile Gln Ile Thr Leu  
 35 40 45  
 Gly His Ile Val Gly Asn Gly Asn Leu Gly Pro Thr Leu Pro Leu Thr  
 50 55 60  
 Leu Ser Tyr Ser Pro Leu Asn Lys Thr Asp Ile Gly Phe Gly Ile Gly  
 65 70 75 80  
 Phe Asn Phe Gly Leu Ser Val Tyr Asp Arg Lys Asn Ser Leu Leu Ser  
 85 90 95  
 Leu Ser Thr Gly Glu Asn Tyr Lys Val Ile Glu Thr Asp Lys Thr Val  
 100 105 110  
 Lys Leu Gln Gln Lys Lys Leu Asp Asn Leu Arg Phe Glu Lys Asp Leu  
 115 120 125

F00007 4255350

Lys	Glu	Asn	Cys	Tyr	Arg	Ile	His	Lys	Ser	Gly	Asp	Ile	Glu	Val	
130						135				140					
Leu	Thr	Gly	Phe	Asn	Asn	Asn	Ala	Phe	Asp	Leu	Lys	Val	Pro	Lys	Lys
145					150					155					160
Leu	Leu	Asn	Pro	Ala	Gly	His	Ala	Ile	Tyr	Ile	Asp	Trp	Asn	Phe	Glu
				165					170					175	
Ala	Thr	Gln	Pro	Arg	Leu	Asn	Arg	Ile	Tyr	Asp	Asp	Leu	Asp	Gly	His
			180					185					190		
Asp	Ile	Pro	Leu	Leu	Asn	Leu	Glu	Tyr	Gln	Gly	Leu	Ile	Lys	Thr	Ile
		195					200					205			
Leu	Thr	Leu	Phe	Pro	Gly	Gln	Lys	Glu	Gly	Tyr	Arg	Thr	Glu	Leu	Arg
		210				215					220				
Phe	Leu	Asn	Arg	Gln	Leu	Asn	Ser	Ile	His	Asn	Phe	Ser	Leu	Gly	Asn
225					230					235					240
Glu	Asn	Pro	Leu	Thr	Trp	Ser	Phe	Gly	Tyr	Thr	Pro	Ile	Gly	Lys	Asn
				245					250					255	
Gly	Ile	Leu	Gly	Gln	Trp	Ile	Thr	Ser	Met	Thr	Ala	Pro	Gly	Gly	Leu
			260					265					270		
Lys	Glu	Thr	Val	Asn	Tyr	Ser	Asn	Asn	Asn	Gln	Gly	His	His	Phe	Pro
		275					280					285			
Gln	Ser	Ala	Asn	Leu	Pro	Val	Leu	Pro	Tyr	Val	Thr	Leu	Met	Lys	Gln
		290				295					300				
Val	Pro	Gly	Ala	Gly	Gln	Pro	Ala	Ile	Gln	Ala	Glu	Tyr	Ser	Tyr	Thr
305					310					315					320
Ser	His	Asn	Tyr	Val	Gly	Gly	Gly	Ser	Asn	Gly	Ile	Trp	Asn	Asn	Lys
				325					330					335	
Leu	Asp	Asn	Leu	Tyr	Gly	Leu	Met	Thr	Glu	Tyr	Asn	Tyr	Gly	Ser	Thr
			340					345					350		
Glu	Ser	Arg	Arg	Tyr	Lys	Asp	Lys	Glu	Gly	His	Asp	Gln	Ile	Val	Arg
		355					360					365			
Ile	Glu	Arg	Thr	Tyr	Asn	Asn	Tyr	His	Leu	Leu	Thr	Ser	Glu	Cys	Lys
		370				375					380				
Gln	Gln	Asn	Gly	Tyr	Ile	Gln	Thr	Thr	Glu	Thr	Ala	Tyr	Tyr	Ala	Ile
385					390					395					400
Ile	Gly	His	Asn	Phe	Asp	Ser	Gln	Pro	Ser	Gln	Phe	Gln	Leu	Pro	Lys
				405					410					415	
Thr	Lys	Thr	Glu	Thr	Trp	Arg	Ser	Ala	Asp	Asn	Ser	Tyr	Arg	Ser	Glu
			420					425					430		
Ile	Thr	Glu	Thr	Thr	Phe	Asp	Glu	Ser	Gly	Asn	Pro	Leu	Thr	Lys	Val
		435					440					445			
Ile	Lys	Asp	Lys	Lys	Thr	Gln	Lys	Ile	Ile	Ser	Pro	Ser	Thr	His	Trp
		450				455					460				
Glu	Tyr	Tyr	Pro	Pro	Ala	Gly	Glu	Val	Asp	Asn	Cys	Pro	Pro	Glu	Pro
465					470					475					480
Tyr	Gly	Phe													

			580					585				590				
Asn	Phe	Thr	Ile	His	Arg	Ser	Gln	Val	Arg	Ser	Arg	Tyr	Thr	Gly	Arg	
		595					600					605				
Leu	Phe	Ser	Asp	Thr	Asp	Thr	Lys	Asp	Ile	Val	Thr	Gln	Met	Ser	Tyr	
	610					615					620					
Asp	Lys	Leu	Gly	Arg	Leu	Leu	Thr	Arg	Thr	Leu	Asn	Ser	Gly	Thr	Pro	
625					630					635					640	
Tyr	Ala	Asn	Thr	Leu	Thr	Tyr	Asp	Tyr	Glu	Leu	Asn	Asn	Leu	Gln	Asp	
				645					650					655		
Asp	Asn	Arg	Pro	Pro	Phe	Val	Ile	Thr	Thr	Thr	Asp	Val	Asn	Gly	Asn	
			660					665					670			
Gln	Leu	Arg	Asn	Glu	Phe	Asp	Gly	Ala	Gly	Arg	His	Val	Ser	Gln	Cys	
		675					680					685				
Leu	Lys	Asp	Ser	Asp	Gly	Asp	Gly	Lys	Phe	Tyr	Thr	Ile	His	Thr	Gln	
	690					695					700					
Gln	Tyr	Asp	Glu	Gln	Gly	Arg	His	His	Thr	Ser	Thr	Tyr	Ser	Asp	Tyr	
705					710					715					720	
Leu	Thr	Asn	Gly	Arg	Gln	Gln	Thr	Asp	Pro	Asp	Lys	Val	His	Leu	Ser	
				725				730						735		
Met	Ser	Lys	Ser	Tyr	Asp	Asn	Trp	Gly	Gln	Ile	Ala	Asn	Thr	His	Trp	
			740					745					750			
Ser	Tyr	Gly	Val	Ser	Glu	Lys	Ile	Thr	Val	Asp	Pro	Ile	Thr	Leu	Thr	
		755					760					765				
Ala	Thr	Lys	Gln	Leu	Gln	Ser	Asn	Ser	Asn	Asn	Val	Gln	Thr	Gly	Lys	
	770					775					780					
Glu	Val	Thr	Thr	Tyr	Thr	Pro	Ser	Gln	Gln	Pro	Ile	Gln	Ile	Thr	Leu	
785					790					795					800	
Phe	Asp	Glu	Ala	Gly	His	Leu	Gln	Ser	Cys	His	Thr	Leu	Thr	Arg	Asp	
				805					810					815		
Gly	Trp	Asp	Arg	Val	Arg	Lys	Glu	Thr	Asp	Ala	Ile	Gly	Gln	Cys	Thr	
			820					825					830			
Ile	Tyr	Gln	Tyr	Asp	Asn	Tyr	Asn	Arg	Val	Ile	Gln	Ile	Thr	Leu	Pro	
		835					840					845				
Asp	Gly	Thr	Ile	Val	Asn	Arg	Lys	Tyr	Ala	Pro	Phe	Ser	Thr	Asp	Thr	
	850					855					860					
Leu	Ile	Thr	Asp	Ile	Arg	Val	Asn	Gly	Ile	Ser	Leu	Gly	Gln	Gln	Thr	
865					870					875					880	
Phe	Asp	Gly	Leu	Ser	Arg	Leu	Thr	Gln	Ser	Gln	Asp	Gly	Gly	Arg	Val	
				885					890					895		
Trp	Ala	Tyr	Thr	Tyr	Ser	Ala	Gly	Asn	Asp	Gln	Cys	Pro	Ser	Thr	Val	
			900					905					910			
Ile	Thr	Pro	Asp	Gly	Gln	Phe	Ile	His	Tyr	Gln	Tyr	Gln	Pro	Glu	Leu	

Thr	Asp	Leu	Ala	Thr	Gly	His	Met	Leu	Thr	Thr	Val	Glu	Phe	Asp	
				1045					1050				1055		
Gly	Leu	Asn	Arg	Glu	Ile	Gly	Arg	Lys	Leu	Cys	Asp	Ser	Ser	Gly	His
			1060					1065					1070		
Thr	Leu	Asp	Ile	Gln	Gln	Ser	Trp	Leu	Lys	Thr	Gln	Gln	Leu	Ala	Asn
		1075					1080					1085			
Arg	Ile	Val	Lys	Leu	Asn	Gly	Val	Leu	Gln	Arg	Thr	Glu	Gln	Tyr	Ser
	1090					1095					1100				
Tyr	Asp	Ser	Arg	Asn	Arg	Leu	Asn	Gln	Tyr	Lys	Cys	Asp	Gly	Ala	Glu
1105					1110					1115					1120
Cys	Pro	Thr	Asp	Lys	Tyr	Gly	His	Ser	Ile	Val	Thr	Gln	Asn	Phe	Thr
			1125						1130					1135	
Tyr	Asp	Ile	Tyr	Gly	Asn	Ile	Thr	Ala	Cys	His	Thr	Thr	Phe	Ala	Asp
		1140						1145					1150		
Gly	Thr	Glu	Asp	His	Ala	Thr	Phe	Lys	Phe	Ala	Asn	Pro	Thr	Asp	Pro
		1155					1160					1165			
Cys	Gln	Leu	Thr	Glu	Val	His	His	Thr	His	Pro	Asp	Met	Pro	Asp	Asn
	1170					1175					1180				
Ile	Arg	Leu	Lys	Tyr	Asp	Lys	Ala	Gly	Arg	Val	Ile	Asn	Ile	Thr	Asp
1185					1190					1195					1200
Asn	His	Gly	Asn	Thr	Glu	Asn	Phe	Thr	Tyr	Asp	Thr	Leu	Gly	Arg	Leu
			1205						1210					1215	
Gln	Asn	Gly	Gln	Gly	Ser	Val	Tyr	Gly	Tyr	Asp	Pro	Leu	Asn	Arg	Leu
		1220						1225					1230		
Val	Ser	Gln	Lys	Thr	Asp	Thr	Leu	Asp	Cys	Glu	Leu	Tyr	Tyr	Arg	Glu
		1235					1240					1245			
Thr	Met	Leu	Val	Asn	Glu	Val	Arg	Asn	Gly	Glu	Met	Ile	Arg	Leu	Leu
	1250					1255					1260				
Arg	Thr	Gly	Glu	Thr	Ile	Ile	Ala	Gln	Gln	Arg	Ala	Ser	Lys	Val	Leu
1265					1270					1275					1280
Leu	Thr	Gly	Thr	Asp	Ser	Gln	Gln	Ser	Val	Ile	Leu	Thr	Ser	Asp	Lys
			1285						1290					1295	
Gln	Asn	Leu	Ser	Gln	Glu	Ala	Tyr	Ser	Ala	Tyr	Gly	Lys	His	Lys	Ser
		1300						1305					1310		
Thr	Ala	Asn	Asp	Ala	Ser	Ile	Leu	Gly	Tyr	Asn	Gly	Glu	Arg	Ala	Asp
		1315					1320					1325			
Pro	Val	Ser	Gly	Val	Thr	His	Leu	Gly	Asn	Gly	Tyr	Arg	Ser	Tyr	Asp
	1330					1335					1340				
Pro	Thr	Leu	Met	Arg	Phe	His	Thr	Pro	Asp	Ser	Leu	Ser	Pro	Phe	Gly
1345					1350				1355						1360
Ala	Gly	Gly	Ile	Asn	Pro	Tyr	Ser	Tyr	Cys	Leu	Gly	Asp	Pro	Ile	Asn
			1365						1370					1375	
Arg	Ser	Asp	Pro	Ser	Gly	His	Leu	Ser	Trp	Gln	Ala	Trp	Thr	Gly	Ile
		1380													



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<210> 24
<211> 105
<212> PRT
<213> Xenorhabdus bovienii
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<210> 25
<211> 129
<212> PRT
<213> Xenorhabdus bovienii
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<400> 25  
Ser Ala Gln Cys Ile Val Gly Lys Val Phe Arg Ile Ser Met Val Ile  
1 5 10 15  
Ser Asp Ile Tyr Tyr Ser Thr Ser Leu Ile Ile Phe Gln Pro Asp Ile  
20 25 30  
Ile Arg His Ile Trp Met Ser Val Tyr Leu Cys Gln Leu Ala Trp  
35 40 45

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<210> 26
<211> 141
<212> PRT
<213> Xenorhabdus bovienii
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<210> 27
<211> 101
<212> PRT
<213> Xenorhabdus bovienii
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<400> 27  
Ala His Cys His Ile Ala Leu Phe Pro Cys Trp His Asn Pro Gln Tyr  
   1              5           10          15  
Cys Gln Gln His Pro Asp His His Ser Asn Cys His His Gln Phe Lys  
               20        25         30  
Gln Glu Tyr Pro Pro Ser Arg Gln Arg Arg Glu Asn Ile Thr Leu Thr  
      35            40             45  
Gln Leu Pro Ile Lys His Thr Gly Ile Glu Ala Gly Ser Gln Thr Asn  
    50            55             60  
Arg Lys Arg Gln Thr Cys Met Phe Gln Arg Ala Asn Glu Ser Lys Val  
65       70           75  
His Gln Leu Gly Gln Asn Gln Gly Arg Asp Arg Asn Phe Tyr Trp Cys  
     85           90          95  
Phe Asp Ile Leu Thr
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100

<210> 28  
 <211> 117  
 <212> PRT  
 <213> Xenorhabdus bovienii

&lt;400&gt; 28

Pro	Gln	Ser	Thr	Pro	Ser	Ser	Gln	Asn	Ser	Arg	Gln	Leu	Thr	Pro	Ala
1				5					10					15	
Glu	Ser	Ser	Gln	His	Gln	Lys	Gln	Lys	Ser	Asp	His	Ile	Glu	Ile	Met
			20					25				30			
Ile	Pro	Ser	Glu	Ala	Pro	Arg	Glu	Tyr	Arg	Glu	Gln	Leu	His	Lys	Ala
		35					40					45			
Thr	Pro	Ala	Arg	Asn	Arg	Asp	Val	Ala	Pro	Asn	Pro	Ser	Val	Phe	Asp
	50					55					60				
Ile	Leu	Arg	Asp	Tyr	His	Trp	Lys	Asn	Phe	Ser	Pro	Val	Lys	Ala	Ala
65					70					75					80
Lys	Ser	Ser	Leu	Thr	Pro	His	Pro	Val	His	Gln	Lys	Ala	Ile	Pro	Leu
				85					90					95	
Asn	Asp	Gln	Arg	Asn	Thr	Ser	Met	Lys	Gln	Ser	Leu	Lys	Pro	Glu	Met
			100					105						110	
Arg	Gln	Lys	Leu	Tyr											
			115												

<210> 29  
 <211> 124  
 <212> PRT  
 <213> Xenorhabdus bovienii

&lt;400&gt; 29

Gly	Lys	Asn	Cys	Ile	Asn	Asp	Gln	Gly	Asn	Leu	Pro	Asp	Arg	Tyr	Thr
1				5					10					15	
Gln	Asn	Cys	Arg	Pro	His	Leu	Thr	Asp	Asn	Pro	Pro	Tyr	Gly	Thr	Val
			20					25					30		
Thr	Glu	Arg	Asn	Pro	Arg	Gln	Tyr	Gln	His	Ala	Asp	Leu	Phe	Gln	Met
		35					40					45			
Arg	Lys	Leu	Ile	Gly	Gln	Leu	Gln	Asn	Pro	Ser	Gly	Asn	Asn	Gly	Pro
	50					55					60				
Thr	Gln	Arg	Gln	His	Trp	Arg	Ile	Ala	Ile	Arg	Ser	His	Lys	Gln	Cys
65					70					75					80
Lys	Asn	Asp	His	Thr	Asp	Ile	Glu	Gln	Cys	Arg	Ser	Lys	Ser	Arg	His
				85					90					95	
Arg	Lys	Ala	Val	Pro	Cys	Ile	Lys	Asn	Cys	Ala	Ser	Gln	Arg	Ser	Gln
			100					105						110	
Arg	Asn	Gln	Lys	Asp	Ile	Arg	Lys	Arg	Asn	Ser	Lys				
			115				120								

<210> 30  
 <211> 515  
 <212> PRT  
 <213> Xenorhabdus bovienii

&lt;400&gt; 30

Asn	Asn	Thr	Met	Asn	Leu	Leu	Lys	Ser	Leu	Ala	Ala	Val	Ser	Ser	Met
1				5					10					15	
Thr	Met	Phe	Ser	Arg	Val	Leu	Gly	Phe	Ile	Arg	Asp	Ala	Ile	Ile	Ala

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			20					25					30		
Arg	Ile	Phe	Gly	Ala	Gly	Met	Ala	Thr	Asp	Ala	Phe	Phe	Val	Ala	Phe
		35					40					45			
Lys	Leu	Pro	Asn	Leu	Leu	Arg	Arg	Ile	Phe	Ala	Glu	Gly	Ala	Phe	Ser
	50					55					60				
Gln	Ala	Phe	Val	Pro	Ile	Leu	Ala	Glu	Tyr	Lys	Asn	Gln	Gln	Gly	Asp
65					70					75					80
Glu	Ala	Thr	Arg	Thr	Phe	Ile	Ala	Tyr	Ile	Ser	Gly	Met	Leu	Thr	Leu
				85					90					95	
Ile	Leu	Ala	Ile	Val	Ser	Val	Ile	Gly	Val	Ile	Ala	Ala	Pro	Trp	Ile
			100					105					110		
Ile	Tyr	Val	Thr	Ala	Pro	Gly	Phe	Thr	Asp	Thr	Pro	Asp	Lys	Phe	Val
		115					120					125			
Leu	Thr	Arg	Asp	Leu	Leu	Arg	Ile	Thr	Phe	Pro	Tyr	Ile	Phe	Leu	Ile
	130					135					140				
Ser	Leu	Ala	Ser	Leu	Ala	Gly	Ala	Ile	Leu	Asn	Thr	Trp	Asn	Arg	Phe
145					150					155					160
Ser	Val	Pro	Ala	Phe	Ala	Pro	Thr	Leu	Leu	Asn	Val	Ser	Met	Ile	Ile
				165					170					175	
Phe	Ala	Leu	Phe	Val	Ala	Pro	Tyr	Cys	Asn	Pro	Pro	Val	Leu	Ala	Leu
			180					185					190		
Gly	Trp	Ala	Val	Val	Ala	Gly	Gly	Val	Leu	Gln	Leu	Ala	Tyr	Gln	Leu
		195					200					205			
Pro	His	Leu	Lys	Lys	Ile	Gly	Met	Leu	Val	Leu	Pro	Arg	Ile	Ser	Phe
	210					215					220				
Arg	Asp	Ser	Ala	Val	Trp	Arg	Val	Ile	Arg	Gln	Met	Gly	Pro	Ala	Ile
225					230					235					240
Leu	Gly	Val	Ser	Val	Gly	Gln	Ile	Ser	Leu	Ile	Ile	Asn	Thr	Ile	Phe
				245					250					255	
Ala	Ser	Phe	Leu	Val	Ser	Gly	Ser	Val	Ser	Trp	Met	Tyr	Tyr	Ala	Asp
			260					265					270		
Arg	Leu	Met	Glu	Leu	Pro	Ser	Gly	Val	Leu	Gly	Val	Ala	Leu	Gly	Thr
		275					280						285		
Ile	Leu	Leu	Pro	Ser	Leu	Ala	Lys	Ser	Phe	Ser	Ser	Gly	Asn	His	Glu
	290					295					300				
Glu	Tyr	Arg	Lys	Leu	Met	Asp	Trp	Gly	Leu	Arg	Leu	Cys	Phe	Leu	Leu
305					310					315					320
Ala	Leu	Pro	Cys	Ala	Val	Ala	Leu	Gly	Ile	Leu	Ala	Glu	Pro	Leu	Thr
				325					330					335	
Val	Ser	Leu	Phe	Gln	Tyr	Gly	His	Phe	Ser	Ala	Phe	Asp	Ala	Glu	Met
			340					345					350		
Thr	Gln	Arg	Ala	Leu	Ile	Ala	Tyr	Cys	Phe	Gly	Leu	Met	Gly	Leu	Ile
		355					360					365			
Val	Val	Lys	Val	Leu	Ala	Pro	Gly	Phe	Tyr	Ser	Arg	Gln	Asp	Ile	Lys
	370														

Leu Leu Arg Leu Met Gly Val Val Ile Ala Gly Ala Gly Ser Tyr Phe  
                   485                  490                  495  
 Ala Val Leu Ala Leu Met Gly Phe Arg Leu Lys Asp Phe Ala His Arg  
                   500                  505                  510  
 Gly Leu Gln  
                   515

<210> 31  
 <211> 216  
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 <213> Xenorhabdus bovienii

<400> 31  
 Ala Ile Ile Leu Ile Arg Asp Lys Leu Ser Arg Ile Phe Ser Arg Gln  
   1                  5                  10                  15  
 Ile Ser Gly Glu Gly Met Phe Gly Tyr Arg Ser Ala Ser Pro Lys Ile  
                   20                  25                  30  
 Arg Phe Ile Thr Asp Arg Met Val Val Arg Leu Val Tyr Glu Arg Asp  
                   35                  40                  45  
 Ala Tyr Arg Leu Ala Glu Tyr Tyr Ser Glu Asn Lys Asp Phe Leu Lys  
                   50                  55                  60  
 Pro Trp Glu Pro Thr Arg Asp Gly Ser Phe Tyr Gln Pro Ser Gly Trp  
   65                  70                  75                  80  
 Thr Asn Arg Leu Asn Tyr Ile Ala Glu Leu Gln Arg Gln Asn Ala Thr  
                   85                  90                  95  
 Phe Asn Phe Val Leu Leu Asp Ser Asp Glu Arg Glu Ile Met Gly Val  
                   100                  105                  110  
 Ala Asn Phe Thr Asn Val Val Arg Gly Ala Phe His Ser Cys Tyr Leu  
                   115                  120                  125  
 Gly Tyr Ser Leu Ala Glu Lys Leu Gln Gly Gln Gly Leu Met Tyr Glu  
                   130                  135                  140  
 Ala Leu Gln Pro Ala Ile Arg Tyr Met Gln Arg Tyr Gln Arg Met His  
   145                  150                  155                  160  
 Arg Ile Met Ala Asn Tyr Met Pro His Asn His Arg Ser Gly Asn Leu  
                   165                  170                  175  
 Leu Lys Lys Leu Gly Phe Glu Gln Glu Gly Tyr Ala Lys Asn Tyr Leu  
                   180                  185                  190  
 Met Ile Asp Gly Val Trp Gln Asp His Val Leu Thr Ala Leu Thr Asp  
                   195                  200                  205  
 Asp Ala Trp Gly Lys Val Gly Leu  
                   210                  215

<210> 32  
 <211> 404  
 <212> PRT  
 <213> Xenorhabdus bovienii

<400> 32  
 Trp Cys Ala Met Ser Leu Val Ser Gln Ala Arg Ser Leu Gly Lys Tyr  
   1                  5                  10                  15  
 Phe Leu Leu Phe Asp Asn Leu Leu Val Val Leu Gly Phe Phe Val Val  
                   20                  25                  30  
 Phe Pro Leu Ile Ser Ile Arg Phe Val Glu Gln Leu Gly Trp Ala Ala  
                   35                  40                  45  
 Leu Ile Val Gly Phe Ala Leu Gly Leu Arg Gln Leu Val Gln Gln Gly  
                   50                  55                  60  
 Leu Gly Ile Phe Gly Gly Ala Ile Ala Asp Arg Phe Gly Ala Lys Pro

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65 70 75 80  
 Met Ile Val Thr Gly Met Leu Leu Arg Ala Leu Gly Phe Ala Leu Met  
 85 90 95  
 Ala Met Ala His Glu Pro Trp Ile Leu Leu Ser Cys Val Leu Ser  
 100 105 110  
 Gly Leu Gly Gly Thr Leu Phe Asp Pro Pro Arg Ala Ala Leu Val Ile  
 115 120 125  
 Lys Leu Thr Arg Pro His Glu Arg Gly Arg Phe Tyr Ser Ile Leu Met  
 130 135 140  
 Met Gln Asp Ser Ala Gly Ala Val Val Gly Ala Leu Ile Gly Ser Trp  
 145 150 155 160  
 Leu Leu Gln Tyr Asp Phe Asn Ile Val Cys Trp Ile Gly Ala Ser Ile  
 165 170 175  
 Phe Val Leu Ala Ala Leu Phe Asn Ala Trp Leu Leu Pro Ala Tyr Arg  
 180 185 190  
 Ile Ser Thr Ile Arg Thr Pro Ile Lys Glu Gly Met Met Arg Val Ile  
 195 200 205  
 Arg Asp Arg Arg Phe Leu Tyr Tyr Val Leu Thr Leu Thr Gly Tyr Phe  
 210 215 220  
 Val Leu Ser Val Gln Val Met Leu Met Phe Pro Ile Ile Ile His Glu  
 225 230 235 240  
 Ile Thr Gly Thr Pro Thr Ala Val Lys Trp Met Tyr Ala Ile Glu Thr  
 245 250 255  
 Ala Ile Ser Leu Thr Leu Leu Tyr Pro Ile Ala Arg Trp Ser Glu Lys  
 260 265 270  
 His Phe Arg Leu Glu Gln Arg Leu Met Ala Gly Leu Phe Leu Met Ser  
 275 280 285  
 Ile Cys Met Phe Pro Ile Gly Trp Val Asn Gln Leu His Thr Leu Phe  
 290 295 300  
 Gly Leu Leu Cys Leu Phe Tyr Leu Gly Leu Val Thr Ala Asp Pro Ala  
 305 310 315 320  
 Arg Glu Thr Leu Ser Ala Ser Leu Ser Asp Pro Arg Ala Arg Gly Ser  
 325 330 335  
 Tyr Met Gly Phe Ser Arg Leu Gly Leu Ala Leu Gly Gly Ala Ile Gly  
 340 345 350  
 Tyr Thr Gly Gly Gly Trp Leu Tyr Asp Thr Gly Arg Asp Leu Asn Met  
 355 360 365  
 Pro Gln Leu Pro Trp Ile Leu Leu Gly Leu Ser Gly Leu Ile Thr Ile  
 370 375 380  
 Tyr Ala Leu His Arg Gln Phe Asn Gln Lys Lys Ile Asp Pro Val Met  
 385 390 395 400  
 Leu Gly Arg His

&lt;210&gt; 33

&lt;211&gt; 191

&lt;212&gt; PRT

&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 33

Lys Gly Ala Asn Met Lys Arg Phe Phe Leu Gly Ala Ala Leu Val Leu  
 1 5 10 15  
 Val Gly Leu Val Ser Gly Cys Asp Gln Phe Lys Asp Phe Ser Ile Asn  
 20 25 30  
 Glu Gly Leu Met Asn Asp Tyr Leu Lys Lys Val His Tyr Gln Lys  
 35 40 45  
 Lys Ile Ser Ile Pro Gly Ile Ala Asn Ala Asn Ile Thr Leu Gly Asp

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<210> 34
<211> 205
<212> PRT
<213> Xenorhabdus bovienii
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<210> 35
<211> 315
<212> PRT
<213> Xenorhabdus bovienii
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<400> 35

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<210> 36
<211> 132
<212> PRT
<213> Xenorhabdus bovienii
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<400> 36																
Lys	Thr	Ser	Gln	Gly	Phe	Thr	Ser	Thr	Thr	Cys	Ser	Asn	Gly	Asn	Val	
1				5					10					15		
Leu	Lys	Ile	Cys	Gly	Leu	Ile	Thr	Pro	Cys	Ser	Ser	Leu	Ile	Gln	Arg	
			20					25					30			
Thr	Tyr	Pro	Asn	Asn	Met	Thr	Ile	Gly	Ile	Phe	Ser	Lys	Glu	Ser	Thr	
		35				40						45				
Ala	Lys	Asn	Phe	Gly	Met	Gly	Phe	Leu	Tyr	Tyr	Phe	Asp	Leu	Arg	Val	
	50					55					60					
Leu	Ser	Pro	Phe	Phe	Lys	Ala	Pro	Ile	Asn	Ile	Phe	Thr	Gly	Trp	Gln	
65					70					75					80	



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<210> 37
<211> 289
<212> PRT
<213> Xenorhabdus bovienii
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<210> 38
<211> 270
<212> PRT
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&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 38

Lys Gly Asn Gln Ile Thr Met Ile Leu Tyr Lys Gly Ser Lys Asn Tyr  
 1 5 10 15  
 Leu Phe Asn Gln Leu Asn Tyr Asp Ser Cys Val Leu Leu Glu Val Asp  
 20 25 30  
 Glu Ser Val Asn Leu Asn Gly Trp Asp Glu Leu Ser Arg Ala Gln Arg  
 35 40 45  
 Leu Leu Phe Leu Met Glu Ile Leu Arg Arg Tyr His Phe Pro Val Gln  
 50 55 60  
 Gly Lys Val Leu Ala Gln Lys Leu Asn Ile Ser Leu Arg Thr Leu Tyr  
 65 70 75 80  
 Arg Asp Ile Ala Ser Leu Gln Ala Gln Gly Ala Ile Ile Glu Gly Glu  
 85 90 95  
 Pro Gly Ile Gly Tyr Val Leu Arg Pro Gly Phe Val Leu Pro Pro Leu  
 100 105 110  
 Met Phe Thr Gln Asn Glu Ile Glu Ala Leu Ala Leu Gly Ala Asn Trp  
 115 120 125  
 Val Ala Lys Arg Ala Asp Pro Gln Leu Lys Glu Ser Ala Asn Asn Ala  
 130 135 140  
 Ile Ser Lys Ile Ala Ala Val Ile Pro Ala Glu Leu Lys Gln Met Leu  
 145 150 155 160  
 Glu Ala Ser Ser Leu Leu Ile Gly Pro Ala Ala Thr Ala Val Gln Pro  
 165 170 175  
 Val Val Glu Ile Gln Gln Ile Arg Gln Ala Ile Asn Thr Arg His Lys  
 180 185 190  
 Ile Thr Leu Ala Tyr Leu Asp Ile Lys Asp Ile Pro Ser Glu Arg Thr  
 195 200 205  
 Ile Trp Pro Phe Ala Leu Gly Tyr Phe Glu Asn Ile Ser Ile Val Ile  
 210 215 220  
 Gly Trp Cys Glu Leu Arg Glu Glu Phe Arg His Phe Arg Ser Asp Arg  
 225 230 235 240  
 Ile Met Arg Leu Lys Ile Glu Asn Gln Cys Tyr Pro Arg Ser Arg Gln  
 245 250 255  
 Val Leu Leu Lys Glu Trp Arg Ala Met Glu Lys Ile Ser Arg  
 260 265 270

&lt;210&gt; 39

&lt;211&gt; 209

&lt;212&gt; PRT

&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 39

Arg Lys Met Thr Ile Tyr Asp Leu Lys Pro Arg Phe Gln Asn Leu Leu  
 1 5 10 15  
 Arg Pro Ile Val Ile Tyr Leu Tyr Lys Gln Gly Ile Thr Ala Asn Gln  
 20 25 30  
 Val Thr Leu Thr Ala Leu Phe Leu Ser Ile Phe Ala Gly Ser Leu Leu  
 35 40 45  
 Ser Leu Phe Pro Ser Pro His Leu Tyr Trp Leu Leu Pro Val Phe Leu  
 50 55 60  
 Phe Ile Arg Met Ala Leu Asn Ala Ile Asp Gly Met Leu Ala Arg Glu  
 65 70 75 80  
 His Asn Gln Lys Ser His Leu Gly Ala Ile Tyr Asn Glu Leu Gly Asp  
 85 90 95  
 Val Ile Ser Asp Val Ala Leu Tyr Leu Pro Phe Cys Leu Leu Pro Asp

100 105 110  
 Val Asn Ser Leu Ser Leu Leu Ile Ile Leu Phe Leu Thr Ile Leu Thr  
 115 120 125  
 Glu Phe Ile Gly Val Leu Ala Gln Thr Ile Gly Ala Ser Arg Arg Tyr  
 130 135 140  
 Asp Gly Pro Ile Gly Lys Ser Asp Arg Ala Phe Ile Phe Gly Ala Tyr  
 145 150 155 160  
 Gly Leu Ile Ile Ala Ile Phe Pro Leu Ala Leu Gly Trp Ser Ile Ser  
 165 170 175  
 Leu Phe Ala Phe Met Ile Ile Leu Leu Leu Val Thr Cys Tyr Gln Arg  
 180 185 190  
 Val Val Lys Ala Leu Arg Glu Ile Arg Leu Ala Glu Gln Ser His Ser  
 195 200 205

Lys

<210> 40

<211> 592

<212> PRT

<213> Xenorhabdus bovienii

<400> 40

Gly Val Asn Met Thr Pro Gln Leu Asp Gln Arg Ile Ala Glu Glu His  
 1 5 10 15  
 Tyr Phe Thr Thr Ser Asp Asn Ala Ser Leu Phe Tyr Arg Tyr Trp Pro  
 20 25 30  
 Gln Gln Gln Ala Asn Pro Asp Arg Ala Ile Ile Ile Phe His Arg Gly  
 35 40 45  
 His Glu His Ser Gly Arg Ile Gln His Val Val Asp Gly Leu Asp Leu  
 50 55 60  
 Pro Asp Val Pro Met Phe Ala Trp Asp Ala Arg Gly His Gly Lys Thr  
 65 70 75 80  
 Glu Gly Pro Arg Gly Tyr Ser Pro Ser Met Gly Thr Ser Ile Arg Asp  
 85 90 95  
 Val Asp Glu Phe Val Arg Phe Ile Ala Thr Gln Tyr Gly Ile Ala Met  
 100 105 110  
 Glu Asn Ile Val Val Ile Gly Gln Ser Val Gly Ala Val Leu Val Ser  
 115 120 125  
 Ala Trp Val His Asp Tyr Ala Pro Lys Ile Arg Ala Met Ile Leu Ala  
 130 135 140  
 Ala Pro Ala Phe Asp Ile Lys Leu Tyr Ile Pro Phe Ala Thr Gln Gly  
 145 150 155 160  
 Leu Gln Leu Met Gln Lys Ala Arg Gly Ile Phe Phe Val Asn Ser Tyr  
 165 170 175  
 Val Lys Ala Arg Tyr Leu Thr His Asp Glu Thr Arg Ile Ala Ser Tyr  
 180 185 190  
 Asn Ser Asp Pro Leu Ile Thr Arg Glu Ile Ala Val Asn Ile Leu Leu  
 195 200 205  
 Asp Leu Tyr Gln Thr Ala Glu Arg Val Val Lys Asp Ala Ala Ala Ile  
 210 215 220  
 Thr Leu Pro Thr Leu Leu Phe Ile Ser Gly Ser Asp Tyr Val Val Asn  
 225 230 235 240  
 Lys Lys Pro Gln His Gln Phe Tyr Gln Gln Leu Asn Thr Pro Ile Lys  
 245 250 255  
 Glu Lys His Val Met Asp Gly Phe Tyr His Asp Thr Leu Gly Glu Lys  
 260 265 270  
 Asp Arg His Leu Val Phe Asp Lys Ile Arg Val Phe Ile Glu Arg Ile

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<210> 41
<211> 121
<212> PRT
<213> Xenorhabdus bovienii
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<400>	41														
His	His	Asn	Ser	Ile	Asn	Val	Leu	Leu	Lys	Asn	Ile	Ile	Ser	Pro	His
1				5					10					15	
Gln	Ile	Met	Leu	Leu	Cys	Phe	Thr	Val	Thr	Gly	His	Asn	Asn	Arg	Pro
			20					25					30		
Ile	Gln	Thr	Glu	Arg	Ser	Leu	Phe	Phe	Thr	Val	Val	Met	Ser	Thr	Gln
		35					40					45			
Asp	Val	Ser	Ser	Met	Ser	Leu	Thr	Asp	Ser	Ile	Cys	Leu	Met	Phe	Leu
	50					55					60				
Cys	Ser	Ser	Arg	Gly	Met	Pro	Val	Asp	Thr	Val	Arg	Gln	Lys	Gly	Arg
65					70						75				80
Val	Thr	Ala	His	Pro	Trp	Glu	Arg	Arg	Phe	Val	Met	Leu	Met	Asn	Leu

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<210> 42
<211> 444
<212> PRT
<213> Xenorhabdus bovienii
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<400>	42															
Ile	Asn	Lys	Tyr	Lys	Met	Glu	His	His	Met	His	Ser	Ser	Leu	Asp	Ser	
1				5					10					15		
Arg	Arg	Arg	Leu	Trp	Leu	Thr	Gly	Val	Ile	Trp	Leu	Leu	Phe	Leu	Ala	
			20					25					30			
Pro	Phe	Phe	Phe	Leu	Thr	Tyr	Gly	Gln	Val	Asn	Gln	Phe	Thr	Ala	Gln	
		35					40					45				
Arg	Ser	Asp	Val	Gly	Thr	Val	Met	Phe	Gly	Trp	Glu	His	Asn	Ile	Pro	
	50					55					60					
Phe	Trp	Ser	Trp	Ser	Ile	Ile	Pro	Tyr	Trp	Ser	Ile	Asp	Leu	Phe	Tyr	
65					70					75					80	
Gly	Ile	Ser	Leu	Phe	Ile	Cys	Thr	His	Arg	Arg	Glu	Gln	Trp	Leu	His	
				85					90					95		
Gly	Trp	Arg	Leu	Met	Thr	Ala	Ser	Leu	Ile	Ala	Cys	Val	Gly	Phe	Leu	
			100					105					110			
Leu	Phe	Pro	Leu	Lys	Phe	Ser	Phe	Ser	Arg	Pro	Thr	Thr	Glu	Gly	Leu	
		115					120					125				
Phe	Gly	Trp	Leu	Phe	Asn	Gln	Leu	Glu	Leu	Phe	Asp	Leu	Pro	Tyr	Asn	
	130					135					140					
Gln	Ala	Pro	Ser	Leu	His	Ile	Ile	Leu	Leu	Trp	Leu	Leu	Trp	Leu	Arg	
145					150					155					160	
Tyr	Ser	Ala	Tyr	Val	Ser	Gly	Tyr	Trp	Arg	Gly	Leu	Leu	His	Ile	Trp	
				165					170					175		
Ser	Val	Leu	Ile	Ala	Leu	Ser	Val	Leu	Thr	Thr	Trp	Gln	His	His	Phe	
			180					185					190			
Ile	Asp	Val	Leu	Thr	Gly	Phe	Ala	Val	Gly	Val	Ile	Leu	Ser	Tyr	Leu	
	195						200					205				
Leu	Pro	Val	Ser	Tyr	Arg	Trp	Arg	Trp	Gln	Pro	Asn	Gln	Asp	Arg	Tyr	
	210					215					220					
Ala	Arg	Lys	Leu	Phe	Gly	Tyr	Tyr	Leu	Thr	Gly	Ser	Ala	Leu	Phe	Ala	
225					230					235					240	
Leu	Ile	Ala	Ser	Leu	Leu	Gly	Gly	Ser	Phe	Trp	Ile	Leu	Leu	Trp	Pro	
				245					250					255		
Ala	Val	Ser	Leu	Leu	Met	Ile	Ala	Leu	Gly	Tyr	Ala	Gly	Leu	Gly	Ser	
			260					265					270			
Ser	Val	Phe	Gln	Lys	Gln	Pro	Asp	Gly	Arg	Met	Ser	Leu	Ser	Ala	Arg	
		275					280					285				
Trp	Leu	Leu	Ala	Pro	Tyr	Gln	Leu	Gly	Ala	Trp	Leu	Ser	Tyr	Leu	Trp	
	290					295					300					
Phe	Arg	Arg	Lys	Ser	Ala	Pro	Phe	Asn	His	Ile	Thr	Glu	Gly	Ile	Ile	
305					310					315					320	
Leu	Gly	Ser	Leu	Pro	Cys	Gln	Pro	Val	Thr	Ala	Val	Ser	Val	Leu	Asp	
				325												

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<210> 43
<211> 174
<212> PRT
<213> Xenorhabdus bovienii
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<210> 44
<211> 466
<212> PRT
<213> Xenorhabdus bovienii
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<400> 44															
Met	Asn	Thr	Arg	Lys	Ile	Asn	Gly	Ile	Arg	Pro	Phe	Ser	Ala	Phe	Ile
1				5					10					15	
Asp	Ser	Cys	Leu	Lys	Glu	Ser	Tyr	Ser	Phe	Pro	Arg	Phe	Ile	Arg	Asp
			20					25					30		
Ile	Ile	Ala	Gly	Ile	Thr	Val	Gly	Val	Ile	Ala	Ile	Pro	Leu	Ala	Met
		35					40					45			
Ala	Leu	Ala	Ile	Gly	Ser	Gly	Val	Ala	Pro	Gln	Tyr	Gly	Leu	Tyr	Thr
	50					55				60					
Ala	Ala	Ile	Ala	Gly	Ile	Val	Ile	Ala	Met	Thr	Gly	Gly	Ser	Arg	Tyr
65					70					75					80

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<210> 45
<211> 125
<212> PRT
<213> Xenorhabdus bovienii
<400> 45
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<210> 48
<211> 308
<212> PRT
<213> Xenorhabdus bovienii
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<400> 48																
Leu 1	Ser	Cys	Ile	Arg 5	Phe	Ile	Phe	Leu	Leu 10	Ile	Gln	Gln	Ile	Tyr 15	Leu	
Pro	Leu	Thr	Arg 20	Glu	Gly	Ile	Ser 25	Met	Gln	Gln	Lys	Val 30	Val	Asn	Ile	
Gly	Asp	Ile 35	Lys	Val	Ala	Asn 40	Asp	Leu	Pro	Phe	Val 45	Leu	Phe	Gly	Gly	
Met	Asn 50	Val	Leu	Glu	Ser 55	Arg	Asp	Leu	Ala	Met 60	Arg	Ile	Cys	Glu	His	
Tyr 65	Val	Thr	Val	Thr 70	Gln	Lys	Leu	Gly	Ile 75	Pro	Tyr	Val	Phe	Lys	Ala 80	
Ser	Phe	Asp	Lys 85	Ala	Asn	Arg	Ser	Ser 90	Ile	Arg	Ser	Tyr 95	Arg	Gly	Pro	
Gly	Leu	Glu 100	Glu	Gly	Met	Lys	Ile 105	Phe	Gln	Glu	Leu	Lys 110	Gln	Thr	Phe	
Gly	Val 115	Lys	Ile	Ile	Thr	Asp 120	Val	His	Glu	Pro	Ala 125	Gln	Ala	Gln	Pro	
Val	Ala 130	Asp	Val	Val	Asp 135	Val	Ile	Gln	Leu	Pro	Ala 140	Phe	Leu	Ala	Arg	
Gln 145	Thr	Asp	Leu	Val 150	Glu	Ala	Met	Ala	Lys	Thr 155	Gly	Ala	Val	Ile	Asn 160	
Val	Lys	Lys 165	Pro	Gln	Phe	Val	Ser	Pro	Gly 170	Gln	Met	Gly 175	Asn	Ile	Val	
Glu	Lys	Phe 180	Lys	Glu	Gly	Gly	Asn 185	Asp	Gln	Val	Ile	Leu 190	Cys	Asp	Arg	
Gly	Ser 195	Asn	Phe	Gly	Tyr	Asp 200	Asn	Leu	Val	Val	Asp 205	Met	Leu	Gly	Phe	
Gly	Val 210	Met	Gln	Gln	Ala 215	Thr	Gln	Gly	Ala	Pro	Val 220	Ile	Phe	Asp	Val	
Thr 225	His	Ala	Leu	Gln 230	Cys	Arg	Asp	Pro	Leu	Gly 235	Ala	Ala	Ser	Gly	Gly 240	
Arg	Arg	Ala 245	Gln	Val	Ala	Glu	Leu	Ala	Arg 250	Ala	Gly	Met	Ala	Val	Gly 255	
Ile	Ala	Gly	Leu	Phe	Leu	Glu	Ala	His	Pro	Asp	Pro	Glu	Asn	Ala	Lys	

	260		265		270										
Cys	Asp	Gly	Pro	Ser	Ala	Leu	Pro	Leu	Ala	Lys	Leu	Glu	Ser	Phe	Leu
	275		280		285										
Met	Gln	Ile	Lys	Ala	Ile	Asp	Asp	Val	Val	Lys	Asn	Phe	Pro	Glu	Leu
	290		295		300										
Asp	Thr	Ser	Lys												
305															

&lt;210&gt; 49

&lt;211&gt; 274

&lt;212&gt; PRT

&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 49

Val	Asp	Gly	Ile	Lys	Met	Lys	Pro	Ile	Val	Asn	Tyr	Glu	Phe	Asn	Asn
1				5				10						15	
Thr	Pro	Leu	Ile	Asp	Gly	Ile	Ile	Leu	Val	Ser	Lys	Ile	Ile	Arg	Pro
			20					25					30		
Asp	Phe	Pro	Gln	Thr	Leu	Val	Ser	Glu	Gln	Leu	Thr	Ala	Leu	Val	Glu
		35					40					45			
Glu	Ala	Arg	Gln	Arg	Leu	Ser	Ser	Ile	Thr	Asp	Ser	Lys	Val	Lys	Leu
	50					55				60					
Asp	Ser	Leu	Leu	Thr	Leu	Phe	Tyr	Arg	Glu	Trp	Lys	Phe	Gly	Gly	Ala
65					70				75					80	
Asn	Gly	Val	Tyr	Cys	Leu	Ser	Asp	Thr	Leu	Trp	Leu	Asp	Arg	Leu	Leu
			85						90					95	
His	Ser	Arg	Gln	Gly	Ser	Pro	Val	Ser	Leu	Gly	Thr	Val	Phe	Thr	His
			100					105					110		
Ile	Ala	Gln	Ala	Leu	Gly	Leu	Ser	Val	Gln	Pro	Val	Ile	Phe	Pro	Ile
		115						120					125		
Gln	Leu	Ile	Leu	Arg	Ile	Asp	Leu	Leu	Asp	Gln	Pro	Thr	Trp	Phe	Ile
	130					135					140				
Asn	Pro	Leu	Asn	Gly	Asp	Thr	Leu	Asn	Glu	His	Thr	Leu	Asp	Val	Trp
145					150					155					160
Leu	Lys	Gly	Asn	Ile	Gly	Pro	Thr	Val	Arg	Leu	Lys	Lys	Gln	Asp	Leu
			165						170					175	
Gln	Glu	Ala	Asp	Asn	Val	Ser	Leu	Val	Arg	Lys	Ile	Thr	Asp	Thr	Ile
			180					185					190		
Lys	Val	Ser	Leu	Met	Glu	Glu	Lys	Lys	Met	Glu	Leu	Ala	Leu	Lys	Ala
		195					200					205			
Ser	Glu	Val	Val	Leu	Thr	Phe	Asp	Pro	Asp	Asp	Pro	Tyr	Glu	Ile	Arg
	210					215					220				
Asp	Arg	Gly	Leu	Ile	Tyr	Ala	Gln	Leu	Asp	Cys	Asn	His	Ile	Ala	Val
225					230					235					240
Ser	Asp	Leu	Ser	Tyr	Phe	Val	Glu	His	Cys	Pro	Glu	Asp	Pro	Ile	Ser
			245						250					255	
Glu	Met	Ile	Lys	Met	Gln	Ile	Asn	Thr	Ile	Glu	Gln	Arg	Leu	Ile	Val
			260					265					270		
Leu	His														

&lt;210&gt; 50

&lt;211&gt; 316

&lt;212&gt; PRT

&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 50

Ser Asp Arg Arg Gln Thr Gly Tyr Ala Tyr Ser Ala Asp His Tyr Arg  
 1 5 10 15  
 Ile Ser Gly Arg Ser Thr Val Cys Thr Val Arg Ala Gly Leu Met Asn  
 20 25 30  
 Tyr Gln Cys Trp Leu Gln His Ala Thr Gln Leu Ser Glu Ser Asp  
 35 40 45  
 Ser Pro Lys Arg Asp Ala Glu Ile Leu Leu Gly Tyr Val Thr Gly Arg  
 50 55 60  
 Ser Arg Thr Tyr Leu Ile Ala Phe Asp Glu Thr Leu Ile Ser Ser Glu  
 65 70 75 80  
 Glu Leu His Gln Leu Asp Ser Leu Leu Val Arg Arg Ile Gln Gly Glu  
 85 90 95  
 Pro Val Ala Tyr Ile Ile Gly Glu Arg Glu Phe Trp Ser Leu Pro Phe  
 100 105 110  
 Ala Val Ser Pro Ala Thr Leu Ile Pro Arg Pro Asp Thr Glu Cys Leu  
 115 120 125  
 Val Glu Lys Ala Leu Glu Leu Leu Pro Asp Ser Pro Ala Arg Ile Leu  
 130 135 140  
 Asp Leu Gly Thr Gly Thr Gly Ala Ile Ala Leu Ala Leu Ala Ser Glu  
 145 150 155 160  
 Arg Asn Asp Cys Tyr Val Thr Gly Val Asp Ile Asn Ser Asp Ala Val  
 165 170 175  
 Met Leu Ala Gln His Asn Ala Glu Lys Asn Ala Gly Lys Leu Ala Ile  
 180 185 190  
 His Asn Val Asn Phe Leu Gln Ser Glu Trp Phe Ala Ala Val Gly Asn  
 195 200 205  
 Gln Gln Phe Asp Met Ile Val Ser Asn Pro Pro Tyr Ile Asp Glu Arg  
 210 215 220  
 Asp Pro His Leu Gln Glu Gly Asp Ile Arg Phe Glu Pro Ala Thr Ala  
 225 230 235 240  
 Leu Ile Ala Ala Gln Asn Gly Met Ala Asp Leu Gln Ala Ile Val Gly  
 245 250 255  
 Gln Ala Arg His Phe Leu Ser Pro Asn Gly Trp Leu Leu Leu Glu His  
 260 265 270  
 Gly Trp Lys Gln Gly Thr Val Val Arg Asn Leu Phe Leu Glu Lys Gly  
 275 280 285  
 Tyr Gln Gln Ile Ala Thr Phe Gln Asp Tyr Gly Gly Asn Glu Arg Ile  
 290 295 300  
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 305 310 315

&lt;210&gt; 51

&lt;211&gt; 289

&lt;212&gt; PRT

&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 51

Val Glu Met Arg Glu Met Ala Gln Glu Glu Leu Lys Glu Ala Lys Ile  
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 Arg Asn Glu Glu Leu Glu Gln Gln Leu Gln Leu Leu Leu Leu Pro Lys  
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 Asp Pro Asp Asp Glu Arg Asn Cys Phe Leu Glu Val Arg Ala Gly Thr  
 35 40 45  
 Gly Gly Asp Glu Ala Ala Ile Phe Ala Gly Asp Leu Phe Arg Met Tyr  
 50 55 60  
 Ser Arg Tyr Ala Glu Ala Arg Arg Trp Arg Val Glu Ile Ile Ser Ala  
 65 70 75 80

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Asn Glu Gly Glu His Gly Gly Tyr Lys Glu Val Ile Ala Lys Val Ser  
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 Gly Asp Gln Val Tyr Gly His Leu Lys Phe Glu Ser Gly Gly His Arg  
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 Val Gln Arg Val Pro Glu Thr Glu Ser Gln Gly Arg Ile His Thr Ser  
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 Asp Ile Ser Pro Gly Asp Leu Lys Ile Asp Thr Phe Arg Ser Ser Gly  
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 His Leu Pro Thr Gly Ile Val Val Glu Cys Gln Asp Glu Arg Ser Gln  
 180 185 190  
 His Lys Asn Lys Ala Lys Ala Met Ser Val Leu Ala Ala Arg Ile Arg  
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&lt;210&gt; 52

&lt;211&gt; 37544

&lt;212&gt; DNA

&lt;213&gt; Xenorhabdus bovienii

&lt;400&gt; 52

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